

PROKARYOTIC REVERSE TRANSCRIPTASE

RELATED CASES

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while This is a continuation-in-part of prior copending U.S. patent application Serial No. 07/315,427, filed February 24, 1989 and since issued as U.S. Patent No. 5,079,151 on January 7, 1992, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/315,316, filed February 24, 1989 and since issued as U.S. Patent No. 5,320,958 on June 14, 1994, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/315,432, filed on February 24, 1989 and since abandoned, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/517,946, filed on May 2, 1990, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/518,749, filed on March 2, 1990, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/753,110, filed on August 30, 1991, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/817,430, filed January 6, 1992, which is a continuation-in-part of prior copending U.S. patent application Serial No. 07/979,447, filed November 20, 1992, respectively which are incorporated herein by reference.

FIELD OF THE INVENTION

The invention relates to bacterial RT enzymes which are capable of synthesizing a hybrid RNA-DNA molecule, called msDNA together with the genes which synthesize the DNA and RNA portion of the molecule.

Another aspect of the invention relates to the isolation and purification of RTs from bacterium which is capable of synthesizing msDNA. The invention deals with groups of prokaryotes & ASSOCIATES e.g., bacteria which are capable of synthesizing msDNAs by means of a reverse transcriptase. The

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bacterium capable of synthesizing msDNAs is identified by testing positive by an appropriate screening test.

This is the first time that, as taught in the subject parent patent applications, reverse transcriptase has been found and isolated from a prokarvote.

BACKGROUND OF THE INVENTION

Previously, there was described a chromosomal region of the bacterium Myxococcus xanthus which coded for the RNA and DNA portions of an msDNA. Dhundale et al. (Dhundale '87) "Structure of msDNA from Myxococcus xanthus: Evidence for a Long, Self-Annealing RNA precursor for the Covalently Linked, Branched RNA", Cell, Vol. 51, pages 1105-1112 (December 24, 1987). Dhundale et al. speculated that an Alu I nucleotide fragment contained all the essential coding regions to produce an msDNA. This speculation turned out to be in error.

The Alu I fragment of Dhundale et al., in fact, and inherently did not contain the gene sequence coding for an RT. The Alu I fragment was too short to code for the gene sequence coding for an RT. This was proven by way of sequence analysis by a computer program which searches for open reading frames that can potentially code for a protein. The print-out of the sequence analysis clearly shows that there is no translational reading frame in the Dhundale et al. fragment open across a stretch of DNA sufficiently long enough to encode any reverse transcriptase.

What is reported in Dhundale et al. in 1987 with respect to a bacterial reverse transcriptase was totally contrary to accepted dogma at that time about the distribution of these enzymes, i.e., that they were present only in viruses which infect eukaryotic organisms.

For the 20 years since the discovery of reverse transcriptase, it was believed that these enzymes were restricted to viruses which infect eukaryotic cells. Now, in accordance with the invention, reverse transcriptases have been identified in bacteria.

SUMMARY OF THE INVENTION

In accordance with the invention, it is shown that various bacteria have nucleotide sequences named "retrons" which encode reverse transcriptases (RTs) which are capable of synthesizing msDNAs. The invention also relates to the isolated and purified bacterial RTs. It has also been determined that the RTs of the bacteria which synthesize msDNAs possess common conserved nucleotide sequences and amino acid residues.

Representative members of the <u>Enterobacteriaceae</u>, <u>Rhizobiaceae</u> and <u>Mycobacteriaceae</u> families are demonstrated to be capable of synthesizing msDNA. These bacteria can be screened for the capability of synthesizing msDNA by an RT labeling or extension <u>in vitro</u> test.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the restriction map of the 3.4 kb fragment around <u>msd</u> and downstream of <u>msr</u>.

Figure 2 shows the nucleotide sequence of the chromosomal region encompassing the msDNA and msd RNA coding regions and an ORF region downstream of msr and the amino acid sequence of Mx162-RT.

Figure 3 shows the amino acid sequence alignment of the msDNA-Mx162 ORF with a portion of the retroviral Pol sequences from HIV and HTLV1 and the ORF of msDNA-Ec67.

Figure 4 shows the sequence similarity of the msDNA-Mx162 reverse transcriptase with other retroelements.

Figure 5 shows the sequence comparison of the regions around the YXDD box of various reverse transcriptases.

Figure 6 shows the detection of msDNA in a clinical isolate of E. coli.

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Figure 7 shows the complete primary and proposed secondary structure of msDNA-Ec67.

Figure 8 shows the determination of the RNA nucleotide sequence for the branched RNA linked to msDNA.

Figure 9 shows the southern blot analysis of <u>E. coli</u> Cl-1 Chromosomal DNA(A) and analysis of msDNA synthesis by pCl-1E and pCl-1P(B).

Figure 10 shows the restriction map of the 11.6 kb Eco RI fragment.

Figure 11 shows the nucleotide sequence of the region from the <u>E. coli</u> Cl-1 chromosome encompassing the msDNA, <u>msd</u> RNA and ORF coding regions and the amino acid sequence of Ec67-RT.

Figure 12 shows the amino acid sequence alignment of the <u>E. coli</u> msDNA ORF with a portion of the retroviral Pol sequence from HIV and HTLV1.

Figure 13 shows the detection of RT activity from various cell extracts.

Figure 14 shows the amino acid sequence alignment of bacterial RTs.

Figure 15 shows the nucleotide and amino acid sequence of Mx65-RT.

Figure 16 shows the nucleotide and amino acid sequence of Sa163-RT.

Figure 17 shows the nucleotide and amino acid sequence of Ec73-RT.

Figure 18 shows the nucleotide and amino acid sequence of Ec86-RT.

Figure 19 shows the nucleotide and amino acid sequence of Ec107-RT.

Figure 20 shows the msDNAs from total RNA prepared from each bacterial strain were specifically labeled with ³²P by the RT extension method (12, 14).

Figure 21 shows a collection of 63 rhizobial isolates screened for the presence of msDNA by the RT extension method.

DETAILED DESCRIPTION OF THE DRAWINGS

Figure 1. Restriction Map of the 3.4-kb fragment Around <u>msd</u> and Downstream of <u>msr</u>.

The locations and the orientation of msDNA and msdRNA are indicated by a small arrow and an open arrow, respectively. A large solid arrow represents an ORF and its orientation. The only two AluI sites (A and B) are shown and the DNA sequence between AluI (A) and AluI (B) was determined previously by Yee et al. (1984).

Figure 2. Nucleotide Sequence of the Chromosomal Region Encompassing the Seq. ID NO. I WASTED TONO. 2 msDNA and msdRNA Coding Regions and an ORF Region Downstream of msr.

The upper strand beginning at the Alu I (A) site (see Figure 1) and ending just beyond the ORF is shown. Only a part of the complementary lower strand is shown from base-301 to -600. The boxed region of the upper strand (332-408) and the boxed region of the lower strand (401-562) correspond to the sequences of msdRNA and msDNA respectively (Dhundale et al., 1987). The starting sites for DNA and RNA and the 5' to 3' orientations are indicated by open arrows. The msdRNA and msDNA regions overlap at their 3' ends by 8 bases. The circled G residue at position 351 represents the branched rG of RNA linked to the 5' end of the DNA strand in msDNA. Long solid arrows labeled a1 and a2 represent inverted repeat sequences proposed to be important in the secondary structure of the primary RNA transcript involved in the synthesis of msDNA (Dhundale et al., 1987). The ORF begins with the initiation codon at base 640. Single letter designations are given for amino acids. The YXDD amino acid sequence highly conserved among known RT proteins is boxed. Numbers on the right hand column enumerate the nucleotide bases and numbers with a* enumerate amino acids. Small vertical arrows labeled Alu I and Smal locate the Alu I and Smal restriction cleavage sites, respectively. The DNA sequence was determined by the chain termination method (Sanger et al., 1977) using synthetic oligonucleotides as primer.

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Figure 3. Amino acid Sequence Alignment of the msDNA-Mx162ORF with a 5 1 Seq. FD, NO; Seq. FD, N

Amino acid sequences are compared with matching residues assigned as follows: (o) amino acid residues shared by all four proteins; (o) amino acid residues shared by msDNA-Mx162 and msDNA-Ec67 RTs; (x) amino acid residues shared by msDNA-Mx162 RT with HIV or HTLV1 RTs. Amino acid sequences showed are from residue-177 to -439 for HIV RT (Ratner et al., 1985); residue-15 to -277 for HTLV1 RT (Seiki et al., 1983); residue-32 to -291 for Ec-67 RT (Lampson et al., 1989); and residue-170 to -435 for Mx-162 RT (this work). The YXDD consensus sequence is outlined with a box.

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Figure 4. Sequence Similarity of the msDNA-Mx162 Reverse Transcriptase with Other Retroelements. A. Sequence similarity of the region from residue-18 to -128 of the msDNA-Sea FD NO. 67
Mx162 RT (see Figure 2) with a carboxyl terminal region of integrase of Moloney murine leukemia virus (Mo-MLV) (residue-1070 to -1179; Shinnick et al., 1981). B. Comparison of the sequence from residue-411 to -485 of the msDNA-Mx162 RT (see Figure 2) with the sequence from residue-396 to -461 of the gap protein of human immunodeficiency virus (HIV; Ratner et al., 1985).

Figure 5. Sequence Comparison of the Regions Around the YXDD Box of Various Reverse Transcriptases.

The region from residue-304 to residue-371 of the msDNA-Mx162 RT (see Figure 2) is aligned with various RTs from different sources. The identical amino acid residues with the msDNA-Mx162 RT are indicated by open circles. The YXDD sequences are boxed. The residue numbers for the amino terminal residues and for the carboxyl terminal residues are indicated by the left and the right hand sides of the sequences, respectively. Mx-162 RT from this work (Figure 2);

Seq. ID No. 11/2

Ec-67 RT from Lampson et al. (1989); Ec-86 RT from Lim and Maas (1989); HIV RT from Ratner 10 No. 11/4

et al. (1985); HTLV1 RT from Seiki et al. (1983); Mo-MLV RT from Shinnick et al. (1981); RSV (Rous sarcoma virus) RT from Dickson et al. (1982); BLV (bovine leukemia virus) RT from Rice

illustrated.

et al. (1985); Mt. plasmid (Neurospora mitochondrial plasmid) RT from Nargang et al. (1984); 17.6

Drosophila retrotransposon from Saigo et al. (1984); gypsy Drosophila retrotransposon from Yuki et

Sea. ID No. 26

Al. (1986); Tal-3 plant (Arabidopsis thaliana) retrotransposon from Voytas and Ausubel (1988); and

Ty912 yeast retrotransposon from Clare and Farabaugh (1985). Small arrows in Copia, Tal-3 and

Ty912 indicate positions of insertions of extra sequences of 18, 18 and 13 residues, respectively. B,

Phylogenetic relationships among various RTs listed in A. The branching positions are arbitrarily

Figure 6. Detection of msDNA in a clinical isolate of <u>E</u>. coli. Total RNA, prepared (Maniatis et al., 1982) from a 5-ml culture, was added to 50 μl of a reaction mixture containing: 50 mM Tris-HCl (pH8.3); 6 mM MgCl₂; 40 mM KCl; 5 mM DTT; 1 μM dATP, dTTP, and dGTP; 0.04 μM dCTP; 0.2 μM [α-³²P]dCTP; and 10 units of AMV-RT (Boehringer Mannheim). The reaction mixture was incubated at 37°C for 30 min. followed by extraction with 50 μl phenol-chloroform (1:1) and ethanol precipitation. The samples were electrophoresed on a 4% acrylamide -8 M urea gel. Lanes: (S) molecular weight markers; Mspl digest of pBR322 end-labeled with [α-³²P]dCTP and the Klenow fragment of DNA polymerase I, (1) <u>E</u>. coli K-12 strain C600, (2) the same as in lane 1 except the sample was treated with RNase A (5 μg, 10 min at 37 °C) just prior to electrophoresis, (3) clinical isolate Cl-1, (4) clinical isolate Cl-1 treated with RNase A. The clinical isolate was identified as <u>Escherichia coli</u> (The clinical <u>E</u>. coli strains were urinary tract isolates kindly provided by Dr. Melvin Weinstein from the microbiology laboratory, R.W. Johnson Hospital, New Brunswick, NJ. The clinical strain Cl-1 was identified using the API-20E identification system (API laboratory products) and gave a typical <u>E</u>. coli profile number of 5044552).

Figure 7. The complete primary and proposed secondary structure of msDNA-Ec67. The DNA sequence was determined by the Maxam and Gilbert method (Maxam et al., 1980) using 3'-end labeled msDNA. The RNA sequence (msdRNA; boxed region) was determined using base-specific RNases as previously described (Dhundale et al., 1987). The 2',5' Branched linkage between the 15th rG residue and the 5' end of the DNA strand was determined using the debranching enzyme from HeLa cells as described previously (Dhundale et al., 1987; Furuichi et al., 1987; Ruskin et al., 1985; Arenas et al., 1987; the debranching enzyme was a gift from Jerard Hurwitz). The branched rG at position 15 is circled, and both RNA and DNA are numbered from their 5' ends.

Determination of the RNA nucleotide sequence for the branched RNA

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Figure 8. linked to msDNA. Total RNA was prepared from the clinical strain Cl-1 and fractionated on a 5% acrylamide gel. msDNA containing full length RNA was eluted from the gel. This fraction was then labeled at the 5' end of the RNA with [Y 32P]ATP and T4 polynucleotide kinase. The 5' end labeled RNA linked to msDNA was again purified on an 18% acrylamide - 8M urea sequencing gel. The labeled RNA was then sequenced using limited digestion with base-specific RNases as described previously (Dhundale et al., 1987). Lanes: OH-, partial alkaline hydrolysis ladder; (0.5 M sodium bicarbonate/carbonate pH9.2); -E, no enzyme treatment of the labeled RNA linked to msDNA; T1, RNase T1 (1U/reaction, 55°, 15 min.); U2, RNase U2 (1U and 0.5U/reaction, 55°, 15 min.); PhyM, RNase PhyM (1U/reaction, 55°, 15 min); Bc, RNase B. cerus (2U/reaction, 55°, 15 min.); CL3, RNase CL3 (2U/reaction, 37°, 15 min.). The large gap in the sequence gel is due to msDNA linked at the rG residue at position 15 by a 2',5' phosphodiester linkage (Furuichi et al., 1987). The RNA sequence at the 3'-end region from the branched rG residue (the upper part of the gel) was determined from 6% gel (data not shown).

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Figure 9. Southern blot analysis of E. coli Cl-1 chromosomal DNA(A) and analysis of msDNA synthesis by pLl-1E and pCl-1P(B). A: The chromosomal DNA was digested with EcoRI (lane 1), HindIII (lane 2), BamHI (lane 3), PstI (lane 4), and BglII (lane 5). For each lane, 3 µg of the DNA digest was applied to a 0.7% agarose gel. After electrophoresis the gel was blotted to a nitrocellulose filter, and hybridization analysis was carried out according to Southern (Southern, 1975) using msDNA labeled by AMV-RT with $[\alpha^{-32}P]dCTP$ as a probe. Numbers at the left represent the molecular weights in kb. B: Total DNA prepared from each strain was treated with RNase A,

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separated on a 5% acrylamide gel and stained with ethidium bromide. Lane S, pBR322 digested with MspI used for molecular size markers; lane 1, DNA prepared from the host strain CL-83(recA); lane 2, CL-83 (recA⁻) transformed with plasmid pCl-1E (11.6 kb EcoRI fragment; see Figure 5); lane 3, with plasmid pCl-1P (2.8-kb PstI(a)-PstI(b) fragment; see Figure 5). An arrow indicates the position of msDNA.

Restriction map of the 11.6-kb EcoRI fragment. In the Cl-1E map, Figure 10. the left-hand half (EcoRI to HindIII) was not mapped. In the Cl-1EP5 map, the locations and the orientations of msDNA and msdRNA are indicated by a small arrow and an open arrow, respectively. A large solid arrow represents an ORF and its orientation.

Nucleotide sequence of the region from the E. coli Cl-1 chromosome Figure 11. encompassing the msDNA and msdRNA coding regions and an ORF downstream of the msdRNA The entire upper strand beginning at the Ball site (see Figure 5) and ending just beyond the ORF is shown. Only a part of the complementary lower strand is shown from base 241 to 420. The long boxed region of the upper strand (249-306) corresponds to the sequence of the branched RNA (msdRNA; see Figure 7) portion of the msDNA molecule. The boxed region of the lower strand corresponds to the sequence of the DNA portion of msDNA (see Figure 7). The starting site for DNA and RNA and the 5' to 3' orientations are indicated by large open arrows. The msdRNA and msDNA regions overlap at their 3' ends by 7 bases. The circled G residue at position 263 represents the branched rG of RNA linked to the 5' end of the DNA strand in msDNA. Long solid arrows labeled a1 and a2 represent inverted repeat sequences proposed to be important in the secondary structure of the primary RNA transcript involved in the synthesis of msDNA (Dhundale et al., 1987). Note that the nucleotide at position 257 (U on the RNA transcript) and the nucleotide at position 373 (G on the RNA transcript) form a U-G pair in the stem between sequence a1 and a2. The proposed promoter elements (-10 and -35 regions) for the primary RNA transcript are also boxed. The ORF begins with SOLIFE 500
SOLIFIE THE ST. ADELEMBA, PA 19102 the initiation codon at base 418. Single letter designations are given for amino acids. The YXDD

Figure 13.

amino acid sequence conserved among known RT proteins is boxed. Numbers on the right hand column enumerate the nucleotide bases and numbers with a* enumerate amino acids. Small vertical arrows labeled H and P locate the <u>HindIII</u> and <u>PstI</u> restriction cleavage sites, respectively. The DNA sequence was determined by the chain termination method (Sanger et al., 1977) using synthetic oligonucleotides as primers.

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Figure 12. Amino acid sequence alignment of the E. coli msDNA ORF with a portion of the retroviral Pol sequence from HIV and HTLV1. Amino acid sequences are compared with matching residues assigned as follows: (+) amino acid common to msDNA and HIV RTs; (o) amino acid shared by msDNA and HTLV1 RTs; and (o) amino acid shared by all three proteins. Arrows divide the protein sequences into three functional domains (Toh et al., 1983; Geng et al., 1985; Varmus, 1985, Tanese et al., 1988): An amino terminal RT domain, a carboxy terminal RNase H region, and a central "tether" region. The specific amino acid residues for the RT, tether, and RNase H domains, for each protein are: HIV, 177-439, 440-600, 601-722 respectively; HTLV1, 15-277, 278-462, 463-592 respectively; msDNA ORF, 32-290, 291-465, 466-586 respectively. The YXDD polymerase consensus sequence is outlined with a box.

were prepared from <u>E</u>. <u>coli</u> strain C2110 (polA⁻) (Tanese <u>et al</u>., 1985; Tanese <u>et al</u>., 1986. <u>E</u>. <u>coli</u> strain C2110 (polA1⁻) was a gift from M. Roth and S. Goff) containing plasmid pCl-1EP5 encoding the msDNA-ORF (see Figure 10) as well as the vector plasmid (pUC9; Yanisch-Perron <u>et al</u>., 1985) alone. Extracts were also prepared from the <u>E</u>. <u>coli</u> strain PRTS7-1 (<u>pol</u>A+) containing the cloned M-MuLV RT gene (Varmus <u>et al</u>., 1985; Tanese <u>et al</u>., 1977; Tanese <u>et al</u>., 1985; Tanese <u>et al</u>., 1986. Crude extracts were prepared essentially as described (Roth <u>et al</u>., 1985; Hizi <u>et al</u>., 1988). Crude extract equivalent to 15 μg total protein was added to a 50 μl reaction cocktail (50 mM tris-HCl pH7.8, 10 mM DTT, 60 mM NaCl, 0.05% NP-40, 10 mM MgCl₂, 0.5 μg poly(rC)-oligo(dG), and 0.1 μM [α-³²P]dGTP and incubated at 37°C for one hour. Five μl of the reaction mixture was then spotted onto

Detection of RT activity from various cell extracts. Crude cell extracts

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DEAE paper (DE81; Whatman Inc.). The paper was washed to remove unincorporated label (Tanese et al., 1985; Tanese et al., 1986) and then exposed to an X-ray film. In row (A) all reactions contain added template primer (poly rC-dG). Row (B) contains control reactions in which no template-primer is added. Columns contain the designated cell extracts: M-MuLV, cloned Moloney Murine Leukemia Virus RT gene; pGB2 (Churchward et al., 1984), vector plasmid in strain C2110; pCl-1EP5, recombinant plasmid with the cloned msDNA gene. The large amount of background activity observed with the M-MuLV control extract is due to the activity of DNA Polymerase I since this extract is obtained from a PolA⁺ strain (HB101).

Figure 14 shows the amino acid sequence alignment of bacterial RT carried out according to Xiong and Eickbush (1990). Amino acids highly conserved in eukaryotic RTs are shown at the top of the sequences. These amino acids include largely unvaried residues or chemically similar residues. (h) Hydrophobic residue; (p) small polar residues; (c) charged residue. Amino acids conserved in all seven bacterial RTs (identical residues plus functional conserved residues indicated by h for hydrophobic residues or p for polar residues) are marked by solid dots at the bottom of the sequences. The consensus sequence shown at the bottom of the sequences is determined when five out of seven sequences contain an identical or a chemically similar residue (h, hydrophobic residue; p, charged and polar residue). The subdomains 1 to 7 are according to Xiong and Eickbush (1990), which are boxed and indicated by numbers. The highly conserved YXDD sequences are also boxed.

Numbers on the right indicate the amino acid positions from the amino terminus for each RT.

Sources for the sequences are Sal63 (Hsu et al. 1992b), Mx162 (Inouye et al. 1989), Mx65 (Inouye et al. 1990), Ec67 (Lampson et al. 1989b), Ec86 (Lim and Maas 1989), Ec73 (Sun et al. 1991), and Ec107 (Herzer et al. 1992).

Figure 15 shows nucleotide sequence of the chromosomal region encompassing the Soq. II No. 37, Wo Mx65-msDNA and msdRNA coding regions and an ORF region downstream of msr. The sequence covers from the Alu I(A) site to 78 bp downstream of the ORF. The complementary strand is only

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shown from bases 121-300. The boxed region of the upper strand (positions 143-191) and the boxed region of the lower strand (positions 186-250) correspond to the sequences of msdRNA and msDNA, respectively. The starting sites for DNA and RNA and the 5' to 3' orientation are indicated by open arrows. The msdRNA and msDNA regions overlap at their 3' ends by 6 bases. The circled G residue at position 206 represents the branched guanosine of RNA linked to the 5' end of the DNA strand in msDNA. Long solid arrows labeled a1 and a2 represent inverted repeat sequences proposed to be important in the secondary structure of the primary RNA transcript involved in the synthesis of msDNA. The ORF begins with the initiation codon at base 279. The YXDD amino acid sequence highly conserved among known RT proteins is boxed. Numbers on the right-hand column enumerate the nucleotide bases, and numbers with asterisks enumerate amino acids (single-letter code). The DNA sequence was determined by the chain-termination method using synthetic oligonucleotides as primers.

Figure 16 shows nucleotide sequences of 3,060 bases encompassing msr, msd, and the TD No 32 2 42 ca. The sequence from base 421 to base 720 which contains msr and msd is shown double stranded. The boxed regions of the upper strand (bases 440 to 540) and the lower strand (bases 508 to 670) correspond to the sequences of msdRNA and msDNA, respectively. The starting sites for msDNA and msdRNA are indicated by open arrows. The circled G at the position 458 is the branched rG of msdRNA linked to the 5' end of msDNA. Long solid arrows labeled with al and all represent inverted repeated sequences proposed to form the secondary structure in the primary RNA transcript which serves to prime msDNA synthesis. Amino acids are indicated by single letters. The YXDD sequence highly conserved among known RTs is boxed. Xe and Bf sites are indicated by arrows. Numbers on the right-hand side and numbers with asterisks represent numbers for bases and amino acids, respectively.

> SEO 1D NO: 43 SEO ID NO: 44 and SEO ID Mo: 45 of msdRNA and msDNA which are boxed and their Figure 17 shows the sequences

orientations are indicated by open arrows. The branched G residue at position 10425 is circled. The

al and a2. Amino acid residues of Ec73-RT are shown by a single-letter code put at the center of each codon.

Figure 18 shows the restriction map of the 3.5 kb insert of pDB808 and nucleotide sequence of chromosomal determinants of the msDNA-RNA compound of E. coli B. (A) Restriction map of the 3.5 kb insert of clone pDB808. The solid bar represents the region whose sequence is presented in (B). Transcription is from left to right. Restriction enzymes are: P, Pstl, H, Hpal; B, Bglll; X, Xhol. (B) Nucleotide sequences of the chromosomal determinants. Only the strand corresponding to the transcript is shown. Nucleotides are numbered starting from the first base observed in the msdRNA. The mdsRNA coding region is overlined, and the msDNA coding region is underlined. The msDNA sequence is complementary to the sequence shown in this figure. Inverted repeats are indicated by double-dashed lines. The G at position 14 is the branched guanylate of msdRNA in the msDNA-RNA compound. IR, 12 bp inverted repeat.

Figure 19 shows sequence of the retron and flanking regions of Ec107. The sequences corresponding to the K-12 genomic DNA are shown in lower case letters from bases 1-99 and 1400-1540. The msRNA and msDNA regions are boxed. Also indicated are the a1-a2 conserved inverted repeats (indicated by arrows) and the branched G, which is circled. The RT consists of 319 amino acids and contains the YXDD sequence (boxed) which is highly conserved among known RTs. The transcription start site occurs at base 170; a possible terminator is indicated by head-to-head arrows following the RT coding region. Primer extension was utilized in order to determine the transcription start site. These sequence data will appear in the EMBL/GenBank/DDJB Nucleotide Sequence Data Libraries under the accession number X62583.

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DETAILED DESCRIPTION OF THE INVENTION

The description which follows describes msDNA and RT from Myxococcus xanthus. This is a typical bacterium which belongs to a genus of bacteria, whose representative members possess an RT capable of synthesizing msDNA.

The existence of a peculiar branched RNA-linked DNA molecule called msDNA (multicopy single-stranded) has been demonstrated in various myxobacteria, Gram-negative soil bacteria (Yee et al., 1984; Dhundale et al., 1985; Furuichi et al., 1987a,b; Dhundale et al., 1987; Dhundale et al., 1988b). msDNA (msDNA-Mx162) from Myxococcus xanthus consists of 162-base single stranded DNA, the 5' end of which is linked to the 2' position of the 20th rG residue of a 77base RNA molecule (msdRNA) by a 2', 5'-phosphodiester linkage (Dhundale et al., 1987). It exists at a level of approximately 700 copies per genome. Stigmatella aurantiaca also possesses an msDNA (msDNA-Sal63) which is highly homologous to msDNA-Mx162 (Furuichi et al., 1987b). In addition to msDNA-Mx162, M. xanthus has another smaller species of msDNA (mrDNA or msDNA-Mx65), which has no primary sequence homology with msDNA-Mx162 or msDNA-Sal63 (Dhundale et al., 1988b). However, all msDNAs so far characterized share key structural features such as a branched rG residue, stem-and-loop structures in RNA and DNA molecules, and a DNA-RNA hybrid at the 3' ends of DNA and RNA molecules.

Previously it was predicted that reverse transcriptase is required for msDNA biosynthesis on the basis of the finding that msdRNA is derived from a much longer precursor, which can form a very stable stem-and-loop structure (Dhundale et al., 1987). This precursor molecule was proposed to serve as a primer for initiating msDNA synthesis as well as a template to form the branched RNA-linked msDNA. The latter reaction requires reverse transcriptase activity. In M. xanthus, the region coding for the RNA molecule (msr) is located on the chromosome in the opposite orientation to the msDNA coding region (msd) with the 3' ends overlapping by 6 bases for msDNA-Mx65 (Dhundale et al., 1988b) or by 8 bases for msDNA-Mx162 (Dhundale et al., 1987). In addition, SO. FIFTEENTH ST.
DELPHA, PA 19102 as in all the msDNAs found in myxobacteria, there is an inverted repeat comprised of a 14-base (215) 875-8883

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sequence for msDNA-Mx65 (Dhundale et al., 1988b) or a 34-base sequence for msDNA-Mx162 (Dhundale et al., 1987) and a 33-base sequence for msDNA-Sal63 (Furuichi et al., 1987b) immediately upstream of the branched G residue and a sequence immediately upstream of the msDNA coding region. As a result of this inverted repeat, a longer primary transcript beginning upstream of the RNA coding region and extending through the msDNA coding region is considered to self-anneal and form a stable secondary structure. When three base mismatches were introduced into the secondary structure immediately upstream of the branched rG residue, msDNA synthesis was almost completely blocked. However, if three additional base substitutions were made on the other strand to resume the complementary base pairing, msDNA production was restored (Hsu et al., 1989). This result strongly supports the proposed model for msDNA synthesis.

It was also shown that a deletion mutation at the region 100 base pairs (bp) upstream of the DNA coding region (msd) and an insertion mutation at a site 500 bp upstream of msd caused a significant reduction in msDNA production (Dhundale et al., 1988a). This indicates that there is a cis- or trans-acting positive element required for msDNA synthesis in this region. In this report we determined the DNA sequence of this region and found an opening reading frame (ORF) coding for 485 amino acid residues beginning with an initiation codon, ATG, which is located 77 bp upstream of msd (or 231 bp downstream of msr). The very close proximity between msd and the ORF suggests that they may be transcribed as a single transcript. The amino acid sequence of the ORF shows similarity with retroviral reverse transcriptases. We discuss a possible origin of the reverse transcriptase gene as well as a possible relationship between the msDNA system and retroviruses. Recently, some strains of Escherichia coli were found to produce msDNA and the gene for reverse transcriptase which is essential for msDNA production, is linked to the msd region, (Lim and Maas, 1989; Lampson et al., 1989b). Comparison of the msDNA systems of M. xanthus and E. coli raises an intriguing question as to how the extensive diversity found in msDNA systems has emerged in bacteria and what possible functions msDNA may have.

In a preceding paper, it was demonstrated that msDNA is in fact synthesized by reverse transcriptase in a cell-free system in M. xanthus (Lampson et al., 1989a).

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Reverse transcriptases are isolated, and if desired, purified, and biological characterization carried out, if desired, by known methods such as those described in Lampson, B.C., M. Viswanathan, M. Inouye and S. Inouye, "Reverse Transcriptase from Escherichia coli Exists as a Complex with msDNA and is Able to Synthesize Double-stranded DNA", J. Biol. Chem. 265: 8490-8496 (1990), which is incorporated by reference as if fully set forth herein.

RESULTS AND DISCUSSION

Identification of an ORF Associated with msd

On the basis of mutations closely associated with msd which significantly reduce msDNA production, it was assumed that in this region there is a cis- or trans-acting element which is essential for msDNA synthesis (Dhundale et al., 1988a). Figure 1 shows a restriction map around msd. The msDNA coding region is shown by a thin arrow from right to left (msd), and the msdRNA coding region by a thick open arrow (msr). In the previous work (Dhundale et al., 1988a), two mutations were constructed; one, a deletion mutation in which the sequence from Alu I(b) to SmaI was replaced by a gene for kanamycin resistance (see Figure 1), and the other an insertion mutation at the Smal site by a gene for kanamycin resistance (see Figure 1).

In order to elucidate the properties of the element required for msDNA production, the DNA sequence of the region upstream of msd was determined as shown in Figure 2. A long open reading frame (ORF) beginning with an initiation codon was found 77 bases upstream of msd. The ORF is preceded by a ribosome binding sequence of AGG (residue 630 to 632) 7 bases upstream of the initiation codon. The ORF codes for a polypeptide of 485 amino acid residues. The Alu I(b) and Smal sites (see Figure 1), where mutations inhibiting msDNA synthesis were created, are located at amino acid residue-12 and -142 of the ORF, respectively or at the nucleotide sequence from residue -672 to -675, and from residue-1061 to -1066, respectively (Figure 2). In Figure 2, msd or the DNA sequence corresponding to the msDNA sequence is indicated by the closed box on the lower strand SUITE 500.
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DELEMA, PA 19102 and the orientation is from right to left. Similarly, the msdRNA sequence (msr) is also indicated by

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the closed box on the upper strand and the orientation is from left to right. The msd and msr regions overlap by 8 bases. An inverted repeat is also indicated by arrows with letters a1 and a2. This inverted repeat comprises a 34-base sequence immediately upstream of the branched G residue (residue 317 to 350; sequence a2 in Figure 2) and another 34-base sequence at the 3' end (residue 597 to 564; sequence a1). This inverted repeat is essential to form a stem structure which provides a stable secondary structure in a long primary transcript. This secondary structure is considered to serve as the primer as well as the template for msDNA synthesis (Dhundale et al., 1987; Hsu et al., 1989).

Sequence Similarity with Retroviral Reverse Transcriptases

When the amino acid sequence of the ORF was compared with known proteins, a striking similarity was found between the sequence from Leu-308 to Ser-351 and retroviral reverse transcriptases (RT). In particular, this region contains the YXDD sequence, the highly conserved sequence in all known RTs. This sequence (Tyr-344 to Asp-347) is boxed in Figure 2. In Figure 3, the ORF sequence of 266 amino acid residues from Ala-170 to Lys-435 is compared with RTs from HIV (human immunodeficiency virus; Ratner et al., 1986) and HTLV1 (human T-cell leukemia virus type 1; Seiki et al., 1983). As mentioned above, within the sequence of 44 amino residues from Leu-308 to Ser-351, there are 14 and 12 identical residues with HIV (32%) and HTLV1 (27%), respectively. The entire RT domains of HIV and HTLV can also be aligned with the ORF sequence from Ala-170 to Lys-435, with much less similarity as shown in Figure 3. However, the same region was found to be extremely well aligned with the RT which was recently found in a clinical strain of Escherichia coli (Lampson et al., 1989b). This E. coli RT consists of 586 amino acid residues, and its amino terminal domain (residue-32 to -291) and the carboxyl terminal domain (residue-466 and -586) have been demonstrated to have sequence similarity with retroviral RT and ribonuclease H. This RT gene from E. coli was shown to be required for the production of msDNA (msDNA-Ec67) and to have reverse transcriptase activity (Lampson et al., 1989b). Figure 3 shows that the sequence similarity between E. coli and M. xanthus RTs is distributed within almost the entire RT region; in 200 SO. FIFTEENTH ST. PHILADELPHIA, PA 19102 particular in the region from Tyr-181 to Ser-212, 15 out of 32 residues are identical (47% similarity); (215) 875-8383 ELECOPIER (215) 875-8394

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in the region from Gly-226 to Gly-265, 19 out of 40 residues (48% similarity); in the region from Leu-308 to Ser-351, 26 out of 44 residues (59% similarity); and in the region from Lys-354 to Asn-408, 21 out of 55 residues (38% similarity). Overall, similarity from Ala-170 to Lys-435 is 32% (85 out of 266 residues are identical). In spite of these similarities, the M. xanthus ORF does not have the domain, which shows apparent sequence similarity with ribonuclease H (RNase H). The RNase H domain is found to be located in the carboxyl terminal region of the same polypeptide in which the RT domain exists in the amino terminal region in the case of the E. coli RT and other retroviral RTs. In the preceding paper, it was shown that there is a precise coupling between RT and RNase H activity (Lampson et al., 1989a). Therefore, RNase H may still reside with the ORF, or RNase H may be encoded by a separate gene.

Sequence Similarity with Other Proteins

In contrast to the E. coli RT and other retroviral RTs, the ORF found in M. xanthus has a long amino terminal extra domain consisting of approximately 170 residues. Interestingly, this region shows some sequence similarities with the carboxyl terminal region associated with integration protein of Mo-MLV (Moloney murine leukemia virus; Shinnick et al., 1981) (see Figure 4A); the sequence from Pro-18 to Leu-128 of the ORF shows 22% similarity (24 out of 111 residues) with the region from Pro-1070 to Leu-1179 of the gag-pol polyprotein of Mo-MLV. It should be noted that this region of Mo-MLV is unique for Mo-MLV integration protein and does not share sequence similarity with other retroviral endonucleases (Johnson et al., 1986). It is also interesting to notice that in Ty retrotransposon, this domain is located in front of the RT domain in contrast to the retroviral endonuclease domain (Clare and Farabaugh, 1985).

As pointed out above, the ORF does not have homology to E. coli or retroviral RNase H. Instead, it has a short sequence of approximately 80 residues after the RT domain. In this region, one can also find sequence similarity with a part of the gag region of HIV. As shown in Figure 4B, the sequence from Gly-411 to Glu-485 has 22 identical amino acid residues (31% similarity) with the SUITE 500 229 SO. FITE FEBRUTH ST. PHILADELPHIA, PA 19102 [215) 875-8383 [ELECOPIER [215) 875-8393 [ELECOPIER [215) 875-8393]

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Requirement of Reverse Transcriptase

The fact that disruption of the ORF significantly reduced msDNA production in M. xanthus (Dhundale et al., 1988a) and the fact that the ORF has sequence similarity with retroviral RTs strongly supports the previous hypothesis that RT is required for the synthesis of msDNA (Dhundale et al., 1987). Recently, we were able to demonstrate that msDNA is indeed synthesized by reverse transcriptase activity in a cell-free system (Lampson et al., 1989a). The fact that a small amount of msDNA (3% of the wild type level) is still produced in the ORF mutants (Dhundale et al., 1988a) is most likely due to another RT associated with smaller msDNA (msDNA-Mx65; previously assigned mrDNA; Dhundale et al., 1988b). In fact, an ORF has been found to be associated with the region responsible for msDNA-Mx65 production.

At present it is unknown if the ORF is transcribed together with msdRNA from a common upstream promoter or if the ORF has its own independent promoter. Previously, a major RNA transcript of approximately 375 bases by S1 mapping (Dhundale et al., 1987) was identified. This transcript covers the region from approximately 75 bases upstream of msr (at around residue-256 in Figure 2) to approximately 70 bases upstream of msd (at around residue-632 in Figure 2). This indicates that this RNA transcript ends at the ribosome binding site (AGG, 630-632) of the ORF. It is possible that the primary RNA transcript covers not only the msr-msd region but also the entire ORF. This transcript of approximately at least 2 kilobases (kb) is then used as the mRNA for the ORF to produce RT. At the same time, the 5' untranslated region of 350 bases forms a stable secondary structure which serves as a primer and a template for msDNA synthesis as previously proposed (Dhundale et al., 1987). Because of the secondary structure, the 5' end region is probably much more stable than the ORF mRNA region. As a result, only the 375-base RNA from the 5' end of the transcript was detected in the previous work. In E. coli, the RT gene was shown to be transcribed from a single promoter for the msr region (Lampson et al., 1989b).

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Evolution of Reverse Transcriptase

All of the RTs so far identified are from eukaryotic origins, and associated with either retroviruses or retrotransponsons. DNA synthesis for retroviruses and transposition events for retrotransponsons occur via RNA which is used as a template for RTs (see review by Varmus, 1985). From amino acid similarity in various RTs, possible evolutionary relationships among these RTs has been proposed (Yuki et al., 1986).

The present invention demonstrates that RTs are not specific to eukaryotes but exist in prokaryotes as well. An intriguing question arises as to the evolutionary relationship between prokaryotic and eukaryotic RTs and the origin of RT. In order to compare the amino acid sequences of these RTs, the sequence of the M. xanthus RT from Gly-304 to Leu-371 was chosen, since this sequence includes the YXDD box, the most conserved region among different RTs. In Figure 5A this sequence is compared with 13 other representative RTs from bacteria, yeast, plant, mitochondrial plasmid, and animal retroviruses. Within these 14 sequences, the D-D sequence (residues-346 and -347) is completely conserved, and both G-311 and Y-344 are also well conserved except for Ty-RT. Besides these residues, L-308, P-309, Q-310, S-315, P-316, L-330, S-351, and L-371 are fairly well conserved among these sequences. On the basis of the numbers of identical amino acid residues, $\underline{\mathbf{M}}$. xanthus RT has the following similarities with other RTs: 47% (32 amino acid residues) with E. coli C1-1 RT; 41% (28) with E. coli B RT; 24% (16) with HIV, BLV, and mitochondrial plasmid RTs; 22% (15) with Mo-MLV RT; 21% (14) with RSV, 17.6, gypsy, and Tal-3 RTs; 19% (13) with HTLV1 RT; 15% (10) with Ty912 RT; and 9% (6) with Copia RT. On the basis of the phylogenetic relationships among RTs proposed by Yuki et al. (1986), and the present data, a dendrogram of homology of various RTs may be constructed as shown in Figure 5B. As proposed earlier (Yuki et al., 1986), modern RTs are composed to two major groups I and II. One group (group II) consists of retrotransponsons found in yeast (Ty912), plant (Tal-3), and Drosophila (Copia). Bacterial RTs seem to belong to the other group (group I) together with other retrotransponsons from Drosophila such as 17.6 and gypsy, mitochondrial plasmid RT, and retroviral RTs. This indicates that both prokaryotic SUITE 500
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Origin of the M. xanthus Reverse Transcriptase

In addition to the sequence similarity between the M. xanthus RT and RTs from retroviruses and retrotransponsons, msDNA shares other interesting similarities with retroviruses and retrotransponsons; msDNA (synthesis of single-stranded DNA) starts at a site 77 bases upstream of the RT gene and the orientation of DNA synthesis is opposite to the direction of translation of the RT gene. In the case of retroviruses and retrotransponsons, single-stranded DNA synthesis proceeds at the 5'-end untranslated region of an RNA molecule which serves as the mRNA for RT as well (Weiss et al., 1985). The orientation of DNA synthesis is also opposite to the direction of translation of the RT gene. In the case of msDNA synthesis an RNA transcript itself serving as a template also serves as a primer by self-annealing to form a stable secondary structure (Dhundale et al., 1987), whereas in the case of retroviruses and retrotransponsons tRNAs are recruited from the cell for the priming reaction. At present it is unknown if branched RNA-linked msDNA is the final product of an unknown function or if it is a stable intermediate leading to other products.

Furthermore, it is of great interest whether the M. xanthus RT is associated with a complex such as virus-like particles such as those found for yeast Ty1 element (Eichinger and Boeke, 1988). In a preliminary experiment, msDNA of M. xanthus exists as a complex with proteins in the cell which sediments as a 22S particle. Characterization of this complex may shed light on questions concerning the relationship between msDNA and retrocomponents as well as the functions of msDNA.

At present, there is no information to support the possibility that msDNA may be a transposable element or an element associated with a provirus (or prophages). It is important to point out that the RT gene from M. xanthus appears to be as old as other genomic genes for the following reasons: (a) Nine independent natural isolates of M. xanthus from various sites (including Fiji Island and eight different sites in the United States) contained mutually hybridizable msDNA (Dhundale et al., 1985). Since under the same hybridization condition, msDNA-Mx162 did not hybridize with msDNA-Sa163 [which has extensive homology in both DNA and RNA sequences with msDNA-Mx162; Dhundale et al., (1987)], the nine independent strains M. xanthus are assumed to contain SUITE 500
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in other M. xanthus genes (Table 1). M. xanthus is known to have a very high G+C content (70%; Johnson and Ordal, 1968) and as a result, all the genes so far characterized have very high G+C contents at the third positions of codons used; 85.4% for vegA (Komano et al., 1987), 85.7% of ops (Inouye et al., 1983), 87.2% for tps (Inouye et al, 1983), 88.4% for mbhA (Romeo et al., 1986), and 93.9% for sigma factor. The average G+C content of the third positions is calculated to be 90.0% for these genes (Table 1). Surprisingly, the G+C content of the third positions of the RT codons is highest among these genes (95.5%; Table 1).

In contrast, the E. coli msDNA system including the RT gene is considered to have been acquired much later in the evolution of E. coli. Reasons for this conclusion include: (a) Only four strains out of 89 independent clinical E. coli strains were found to produce msDNAs (Lampson et al., 1989b). (b) The codon usage of the E. coli RT is significantly different from the general codon usage of E. coli genes obtained from 199 E. coli genes (Maruyama et al., 1986). In particular, out of 62 arginine codons used in the E. coli RT, 40 (65%) use AGA or AGG in contrast to 2.7% for the AGA+AGG usage among all arginine codons in 199 E. coli genes (see Table 1). The AGA and AGG codons are the least used codons in E. coli (Maruyama et al., 1986). In addition to AGA and AGG codons, many other codons, GCC and GCG for Ala, CGU and CGC for Arg, CAG for Gln, GGC and GGA for Gly, CAC for His, AUC and AUA for Ile, UUA, CUU and CUG for Leu, UUC for Phe, CCU and CCG for Pro, UCG for Ser, ACC and ACA for Thr, and GUC for Val. (c) Although the E. coli msDNAs share little sequence homology, they all share the key secondary structures of a branched rG residue, a DNA-RNA hybrid at the 3' ends of the msDNA and msdRNA, and stem-andloop structures in RNA and DNA strands (Lampson et al., 1989b; Lim and Maas, 1989).

These results clearly demonstrate distinct differences between the msDNA systems of E. coli and M. xanthus. Myxobacteria are common organisms in soil and are found all over the world regardless of climate, and considered to diverge from their nearest bacterial relatives about $2x10^9$ years ago when the atmosphere became aerobic (see a review by Kaiser, 1986). Since it is reasonable to assume that the M. xanthus RT gene is as old as other genomic genes, the RT gene existed much Sulties of the surface of the surfac

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prokaryotic and eukaryotic RTs as shown in Figures 5A and B strongly supports the existence of a single ancestral gene for all RTs. It is possible that such an ancestral RT gene was independently recruited into different systems such as the msDNA system, the retrotransposon system, and the retroviral system. Alternatively, the msDNA system may be a primitive ancestral system from which retrotransposons and retroviruses originated. In this regard, it is intriguing to point out other sequence similarities between the M. xanthus RT-ORF and other retroelements (see Figure 4) other than RT itself as well as the similar mode of initiation of DNA synthesis by RT as discussed earlier.

At present, it is beyond our speculation why the <u>E. coli</u> msDNA systems are so diverged in contrast to the <u>M. xanthus</u> msDNA system and how they were acquired into the genomes of some <u>E. coli</u> strains. However, it should be noted that the <u>E. coli</u> RTs are most related to the <u>M. xanthus</u> RT indicating that they were not derived from eukaryotic origins. Possible origins of retroviruses have been discussed (Temin, 1980). The recent finding of an imposon in a genetic component for a mouse gene also raises an interesting question concerning the evolution of retroelements (Stavenhagen and Robins, 1988). Further characterization of the prokaryotic RTs and the msDNA systems will provide clues to the origins of RT and other retroelements.

EXPERIMENTAL PROCEDURE

DNA Manipulation and Plasmids

DNA manipulation was performed as described by Maniatis et al. (1982). The plasmid isolation was as originally described by Birnboim and Dolly (1979). Plasmid pmsSB7 containing the 5 kb Sall-BamHI fragment shown between the Sall and BamHI sites of pUC9 (Vieira and Messing, 1982) was used. After the 2.2 kb Sall-Smal fragment from pmsSB7 was subcloned between the Sall and Smal sites of pUC9, all Rsal fragments were gel-purified and cloned into pUC9 for DNA sequence.

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DNA sequence

DNA sequence was determined by the chain termination method (Sanger et al., 1977) using single-stranded or double-stranded DNA as templates with synthetic oligonucleotides.

Other Material and Methods

Restriction enzymes were purchased from either Bethesda Research Laboratories or New England BioLabs. [α-35S] dATP was from Amersham. Sequenase, Version 2.0 Kit was purchased from United States Biochemical Corporation for DNA sequences.

Cyborg program from International Biotechnologies Inc. was used to search sequence homology in GenBank Release 55.

Screening of bacteria for retron synthesized msDNAs was performed by the methods of Lampson et al. J. Bacteriol, 173:5363-5370 (1991), or Yee et al, Cell, 38, 203-209 (1984).

RTs were identified and isolated by the method of Lampson et al, J. Biol. Chem, 265:8490-8496.

msDNA in Escherichia coli

The recent serendipitous finding of msDNA (msDNA-Ec86) in E. coli B by Dongbin Lim and Werner Maas (D. Lim et al., 1989) prompted a to search for msDNA in other E. coli strains. Previously established by Yee et al. (T. Yee et al., 1984), msDNA is not found in the common laboratory strain K12, however, to our surprise, it was in a clinical E. coli strain isolated from a patient with a urinary tract infection. Fifty independent E. coli urinary tract isolates were examined for the presence of msDNA (The clinical E. coli strains were urinary tract isolates kindly provided by Dr. Melvin Weinstein from the microbiology laboratory, R.W. Johnson Hospital, New Brunswick, NJ. The clinical strain Cl-1 was identified using the API-20E identification system (API laboratory products) and gave a typical E. coli profile number of 5044552.). The screening method involved treatment of total RNA prepared from each strain with (AMV) RT in the presence of $[\alpha^{-32}P]dCTP$ sure soon in the sound of the sound in the s

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a DNA-RNA duplex structure, the 3' end of the DNA molecule serves as an intramolecular primer and the RNA molecule as a template for RT. When RNA prepared from one of the clinical strains, E. coli Cl-1, was labeled in this manner, two distinct, low molecular weight bands of about 160 bases became labeled with 32P and are shown in Figure 6. If the labeled sample is digested with ribonuclease (RNase) A prior to loading on the gel, a single band corresponding to 105 bases of single-stranded DNA is detected (lane 4). This indicates that both bands in lane 3 contain a singlestranded DNA of identical size. The two labeled bands observed prior to RNase treatment (lane 3) are due to two species of msDNA comprised of a single species of single-stranded DNA linked to RNA molecules of two different sizes. RNA molecules of two different sizes have been observed at the 5' ends of msDNA from myxobacteria in which a precursor molecule contains a longer RNA which is processed into a smaller mature form (Dhundale et al., 1987; Furuichi et al., 1987). Among the 89 clinical isolates screened, three other strains produced msDNA-like molecules of varying size and quantity, suggesting extensive diversity among these molecules. As previously reported (Dhundale, 1985), msDNA was not observed in the E. coli K-12 strain, C600 (lanes 1 and 2, Figure 6).

Nucleotide sequence of msDNA Ec-67

To determine the base sequence of the DNA molecule, the RNA-DNA complex isolated from the clinical strain was labeled at the 3' end of the DNA molecule with AMV-RT and $[\alpha^{-32}P]dATP$. By adding dideoxy-CTP, ddTTP, and ddGTP to the reaction mixture, a single labeled adenine is added to the 3' end of the DNA molecule. RNA is removed with RNase A+ T1 and the end-labeled DNA is subjected to the Maxam and Gilbert sequencing method (Maxam et al., 1980). Figure 7 shows that msDNA consists of a single-stranded DNA of 67 bases and, as in the case of msDNAs from myxobacteria (Yee, 1984; Dhundale, 1987), it can form a secondary hair-pin structure. The primary sequence, however, is not homologous to any of the myxobacterial msDNAs, nor to the msDNA from E. coli B (msDNA-Ec86; Lim and Maas, personal communication).

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The sequence of the RNA molecule was determined using the RNA-DNA complex purified from E. coli Cl-1. The RNA sequence was determined using base specific RNases as described previously (Dhundale et al., 1988). As shown in Figure 8, a large gap is observed in the RNA sequence "ladder". This gap is due to the DNA strand branched at the 2' position of the 15th rG residue of the RNA strand which produces a shift in mobility of the sequence ladder (see Figure 7). The RNA consists of 58 bases with the DNA molecule branched at the G residue at position 15 by a 2',5'-phosphodiester linkage. The branched G structure was determined as previously described for msDNAs from myxobacteria (Dhundale, 1987; Furuichi et al., 1987). After RNase (A and T1) treatment, msDNA retains a small oligoribonucleotide linked to the 5' end of the DNA molecule due to the inability of RNases to cleave in the vicinity of the branched linkage. The 5' end was labeled with [Y-32P]ATP using T4 polynucleotide kinase and the labeled RNA molecule was detached from the DNA strand by a debranching enzyme purified from HeLa cells (Ruskin et al. 1985; Arenas et al., 1987; the debranching enzyme was a gift from Jerard Hurwitz). This small RNA was found to be a tetraribonucleotide which could be digested with RNase T1 to yield a labeled dinucleotide (not shown). Since RNase T1 could not cleave the RNA molecule at the G residue before debranching enzyme treatment, it was concluded that the single-stranded DNA is branched at the G residue via a 2',5'-phosphodiester linkage. In addition, partial RNase U2 digestion cleaved the RNA molecule to yield a ³²P-labeled mono- and a ³²P-labeled trinucleotide (not shown). Thus, the sequence of the tetranucleotide is 5'A-G-A-(U or C)3'. Based on these data, the complete structure of msDNA-Ec67 from E. coli Cl-1 is presented in Figure 7. Despite a lack of primary structural homology, msDNA-Ec67 displays all the unique features found in msDNAs from myxobacteria. These include a singlestranded DNA with a stem-and-loop structure, a single-stranded RNA with a stem-and-loop structure, a 2',5'-phosphodiester linkage between the RNA and DNA, and a DNA-RNA hybrid at their 3' ends. This hybrid structure was confirmed by demonstrating sensitivity of the RNA molecule to RNase H (not shown).

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Cloning of the locus for msDNA-Ec67

In order to identify the DNA fragment which is responsible for msDNA synthesis in E. coli Cl-1, Southern blot hybridization was carried out with various restriction enzyme digests of total chromosomal DNA prepared from E. coli Cl-1, using msDNA-Ec67 labeled with AMV-RT (the same preparation as shown in lane 3, Figure 6) as a probe. The result is shown in Figure 9A. EcoRI (lane 1), HindIII (lane 2), BamHI (lane 3), PstI (lane 4) and Bg11II (lane 5) digestions showed single band hybridization signals corresponding to 11.6, 2.0, 22, 2.8 and 2.5 kilobase pairs (kb), respectively. The upper band appearing in the EcoRI digestion is due to incomplete digestion of the chromosomal DNA. Analysis of total chromosomal DNA prepared from E. coli Cl-1 by agarose gel electrophoresis revealed that the strain contains two plasmids of different size. However, neither plasmid hybridized with the ³²P- labeled probe, indicating that the fragments detected in Figure 9A are derived from chromosomal DNA. Furthermore, there is only one location for the msDNA-coding region on the chromosome, since various restriction enzyme digestions gave only one band of varying sizes. Similar results were observed for the msDNAs of myxobacteria (Yee et al., 1984; Furuichi et al., 1987; and Dhundale et al., 1988).

The 11.6-kb EcoRI fragment and the 2.8-kb PstI fragment were each cloned into pUC9 (Yanisch-Perron et al., 1985) and E. coli CL83 (a recA transductant of strain JM83), an msDNA-free K-12 strain (lane 1, Figure 9B), was transformed with the plasmids. Cells transformed with the 11.6-kb EcoRI clone (pCl-1E) were found to produce msDNA (lane 2, Figure 9B), whereas cells transformed with the 2.8-kb PstI clone (pCl-1P) failed to produce any detectable msDNA (lane 3, Figure 9B). A map of the 11.6-kb fragment is shown in Figure 10. Southern blot analysis of the fragment revealed that a 1.8-kb PstI - HindIII fragment hybridized with the msDNA probe. When the DNA sequence of this fragment was determined, a region identical to the sequence of the msDNA molecule was discovered. The DNA sequence corresponding to the sequence of msDNA is indicated by the enclosed box on the lower strand in Figure 11 and the orientation is from right to left. The location of this sequence is also indicated by a small arrow in Figure 10. As is the case for all other known myxobacterial msDNAs (Dhundale et al., 1987; Furuichi et al., 1987; and Dhundale et al.,

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1988), a sequence identical to that of the RNA linked to msDNA (see Figure 7) was found downstream of the msDNA-coding region in opposite orientation and overlapping with that region by 7 bases. This sequence is indicated by the enclosed box on the upper strand in Figure 11 and the branched G residue is circled. Again, as in all the msDNAs found in myxobacteria, there is an inverted repeat comprised of a 13-base sequence immediately upstream of the branched G residue (residue 250 to 262; sequence a2 in Figure 11) and a sequence at the 3' end shown by an arrow in Figure 11 (residue 368 to 380; sequence a1). As a result of this inverted repeat, a putative longer primary RNA transcript beginning upstream of the RNA coding region and extending through the msDNA coding region would be able to self-anneal and form a stable secondary structure, which is proposed to serve as the primer as well as the template for msDNA synthesis (Dhundale et al., 1987).

Existence of an essential gene for msDNA synthesis

The 2.8-kb PstI fragment (from PstI(a) to PstI(b) in Figure 10) was not able to synthesize msDNA. However, an overlapping 3.9-kb fragment from Ball (1.0 kb downstream of PstI(a); see Figure 10) to the following EcoRI site contains all the information required for synthesis of msDNA. This indicates that a region downstream of the PstI(b) site (Figure 10) is required for msDNA production. The nucleotide base sequence from this region revealed a long open reading frame (ORF) of 586 amino acid residues, starting with the initiation codon ATG at nucleotide 418 to 420 as shown in Figure 11. A distance of only 51 bases separates the initiation codon from the region which encodes msDNA. A putative Shine-Dalgarno sequence (GGA) can be found 10 bases upstream of the initiation codon. When the lacZ gene was fused in frame at the HindIII site (within the ORF) at amino acid residue-126, \(\beta\)-galactosidase activity was detected (not shown). Thus the region encompassing the ORF is indeed transcribed and the gene product encoded by the ORF is essential for msDNA synthesis. In a preliminary experiment, both msdRNA and the ORF appeared to be transcribed as the same transcription unit, since a deletion mutation removing the sequence from residue 1 to 181 blocked the expression of the lacZ gene fused at the HindIII site. A putative SUITE 500
1-SO. FIFTEENTH ST. ADELPHAST. Promoter can be found in the deleted sequence as boxed in Figure 11. These -35 and -10 regions (215) 875-83894

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probably serve as the promoter for both msdRNA synthesis and the ORF.

Sequence similarity with retroviral reverse transcriptases

When the amino acid sequence of the ORF was compared with known proteins, a striking similarity was found with retroviral RTs. In Figure 12, the ORF is compared with RTs from HIV (human immunodeficiency virus; Ratner et al., 1985; and Johnson et al., 1986), and HTLV1 (human T-cell leukemia virus type I; Seiki et al., 1983; and Patarca et al., 1984). The first domain (Asn-32 to Val-291) matches well with the RT domains of HIV and HTLV1. In particular, the sequences around the polymerase consensus "Asp-Asp" sequence (Toh et al., 1983; and Geng et al., 1985; boxed in Figures 11 and 12) are well conserved. Out of 260 amino acid residues in this domain, 44 and 38 residues are identical with HIV and HTLV1, respectively. Between HIV-RT and HTLV1-RT, there are 78 identical amino acid residues in this domain.

The <u>pol</u> gene of retroviruses is known to produce a protein consisting of RT and RNase H activities; the former at the amino-terminal and the latter at the carboxyl-terminal region of the <u>pol</u> gene product (Ratner <u>et al.</u>, 1985; Johnson <u>et al.</u>, 1986; Varmus, 1985; and Tanese <u>et al.</u>, 1988). These domains have been shown to be separated by a poorly conserved "tether" domain of approximately 160 to 190 amino acid residues (Ratner <u>et al.</u>, 1985; Johnson <u>et al.</u>, 1986). On the basis of the HIV sequence, the similarities (only identical amino acid residues) between HIV and HTLV1 are 29.5 and 16.8% for the RT domain and the tether domain, respectively. The similarities between HIV and msDNA are 16.9 and 10.3% for the RT domain and the tether domain, respectively. The similarities between HTLV1 and msDNA are 14.6 and 15.5% for the RT domain and the tether domain, respectively. These results indicate that in addition to the RT region, there are reasonable similarities in the tether domain between retroviruses and msDNA. An alignment of the RNase H domains also revealed that there are similarities between retroviruses and msDNA (15.7 and 17.4% with HIV and HTLV, respectively; see Figure 12). The similarity between HIV and HTLV1 in this region is 18.0%.

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Cell extracts were prepared and assayed for the presence of RT activity associated with the production of msDNA as predicted from the amino acid homologies. Only the E. coli strain (C2110, polA) (Tanese et al., 1985; Tanese et al., 1986; E. coli strain C2110 (polA1) was a gift from M. Roth and S. Goff) harboring the plasmid, pCl-1EP5, containing the msDNA ORF displayed RT activity (Figure 13). The polA strain was used to eliminate high background activity in the RT assay due to DNA polymerase I. No RT activity was detected in extracts containing the vector plasmid alone, or when the template-primer (poly rC-dG) was absent from the reaction mix (Figure 13). It is interesting to note that the PstI(b) site is located at amino acid residue-430, which is between the tether domain and the RNase H domain. A plasmid lacking sequences downstream of the PstI(b) site did not produce msDNA. This suggests that the RNase H domain may be essential for msDNA synthesis, or alternatively that PstI disruption may result in inactivation of RT.

In addition to the similarity between msDNA-Ec67 RT and retroviral RT, there is an interesting similarity between msDNA and retroviruses; DNA synthesis starts at a site upstream of the RT-RNase H gene, and the orientation of DNA synthesis is opposite to the direction of transcription of the RT-RNase H gene. In the case of retroviruses, tRNAs are recruited from the cell for the priming reaction (Weiss et al., 1985), whereas for msDNA an RNA transcript serving as, template also serves as a primer by self-annealing to form a stable secondary structure (Dhundale et al., 1987; Furuichi et al., 1987).

Origin of the E. coli Reverse Transcriptase

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At present the relationship between msDNA and retroviruses is an open question. It is possible that the study of msDNA may shed light on the question of the origin and evolution of retroviruses. It is an intriguing question to consider why some of the clinical E. coli strains, isolated from human patients produce msDNA. Our preliminary data indicate that msDNAs produced by four independent E. coli strains, isolated from urinary track infections, share little homology. This suggests that there may be enormously large numbers of species of msDNA in E. coli. In contrast to SUITE 500 SO. FIFTEENTH ST. ADELPHIA, PA 19102 msDNAs found in \underline{E} . \underline{coli} , msDNA-Mx162 from \underline{M} . $\underline{xanthus}$ is highly conserved, since nine (215) 875-8883

independent M. xanthus strains isolated from various sites have msDNA which hybridizes with the original msDNA-Mx162 (Dhundale et al., 1985). Furthermore, msDNA from another myxobacterium, S. aurantiaca (msDNA-Sa163; Furuichi et al., 1987), also shows a high degree of homology to msDNA-Mx162 (Furuichi et al., 1987).

Several lines of evidence suggest that the RT gene found in the E. coli strain Cl-1 is

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not likely to have originated in E. coli, but rather was recently acquired from some other source. For example, only about 4% of E. coli strains tested were found to produce msDNA. In addition, the RT gene from strain Cl-1 does not cross hybridize to chromosomal DNA from four other E. coli strains which produce msDNA molecules, indicating that there is extensive diversity among these RT genes. In contrast, a DNA fragment from the E. coli-K-12 sigma factor gene can hybridize to chromosomal DNA from all five msDNA producing, E. coli strains, indicating the conserved nature of sigma factors. An analysis of the E. coli RT gene indicates that the codon usage for this gene is remarkably different from most E. coli proteins. In particular, AGA and AGG, the least frequently (2.7%) used codons for arginine among 199 E. coli genes (Maruyama et al., 1986), occurs at a frequency of 64.5% in the E. coli RT gene. Similarly, CUG is the most commonly used codon for leucine (61.3%; Maruyama et al., 1986) in E. coli genes, while its prevalence in the RT gene is only 9.1%. The AT base pair content of the E. coli RT gene was calculated to be 67.6%, which is substantially higher than the AT content of the E. coli genome (45%; Fasman, 1976). The AT contents of HIV and HTLV1 RT genes are 62.1% and 47.8%, respectively. These facts pose an intriguing question as to how and when the RT gene, as well as the msDNA coding region, were integrated into the genome of the clinical strain.

There are many questions to be answered, including (a) are there any particles associated with msDNA, (b) is the msDNA region transposable like the Ty element of yeast (Boeke et al., 1985; Eichinger et al., 1988), (c) can the element responsible for the production of msDNA be transferred from cell to cell, (d) can a RT from one strain (E. coli or myxobacteria) complement the production of msDNA of other strains, (e) does the promoter for the RNA transcript have any

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similarities to the retroviral LTR, (f) are there any specific integration sites for the msDNA element SUITE 500 230 SO. FIFTEENTH ST. PHILADELPHIA, PA 19102 (215) 875-8383 ELECOPIER (215) 875-8394

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on the E. coli chromosome, (g) why is the branched G residue conserved, (h) is there an enzyme responsible for priming DNA synthesis at the 2'-OH position of the rG residue, (i) why and how does msDNA synthesis stop at one distinct site on the RNA template, and (j) how different biochemically are the msDNA RTs from retroviral RTs?

The existence of reverse transcriptase in prokaryotes, previously speculated upon (Dhundale et al., 1987), is now evident. This fact raises intriguing questions concerning possible roles of this enzyme in the prokaryotes other than a role in msDNA production. Recently we also found that M. xanthus, in which msDNA was originally discovered, has a long ORF in the same manner as found for msDNA-Ec67. This ORF has a high degree of similarity to the E. coli RT. Since eight independent isolates of \underline{M} . $\underline{xanthus}$ produce homologous msDNA, the \underline{M} . $\underline{xanthus}$ RT is likely to have been acquired at a very early stage of its evolution in contrast to the E. coli RT. The determination of the structures of both M. xanthus and other E. coli RTs will shed light on the key question of the origin of RT and its role in prokaryotes.

An important embodiment of the invention relates to the discovery of msDNAproducing retron elements in a number of diverse bacterial groups. Thus, retron elements appear to be widely prevalent, at least amongst the purple bacteria or proteobacteria including Proteus, Klebsiella and Salmonella of the gamma subdivision; Rhizobium and Bradyrhizobium from the alpha subdivision; and Nannocystis (a myxobacterium) from the delta subdivisions. These are representatives of the three of the four major subdivisions of the purple bacteria of proteobacteria. As shown above the retron-encoded RT is responsible for the synthesis of msDNAs.

The retron elements were discovered by detecting the presence of msDNA by one of two classic methods: the so-called "RT extension method", described by Lampson, B.C., M. Inouye and S. Inouye, 1991. Survey of multicopy single-stranded DNAs and reverse transcriptase genes among natural isolates of Myxococcus xanthus. J. Bacteriol. 173:5363-5370 and in Lampson, B.C., M. Viswanathan, M. Inouye and S. Inouye, 1990. Reverse transcriptase from Escherichia coli exists as a complex with msDNA and is able to synthesize double-stranded DNA. J. Biol. Chem. 265:8490-SUITE SOO. SECTION ST. 8496 or polyacrylamide gel electrophoresis of a chromosomal DNA extract followed by staining with ADELPHIA, PA 19102

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ethidium bromide as described by Yee, T., T. Furuichi, S. Inouye, 1984. Multicopy Single-Stranded DNA Isolated from a Gram-Negative Bacterium, Myxococcus xanthus. Cell, Vol. 38, 203-209. Both of these publications are incorporated herein by reference. Both methods provide a reliable, convenient and conventional protocol for screening of bacteria for the presence of retron-encoded RT and msDNAs.

In accordance with the RT extension method, the DNA portion of msDNA is specifically 32P radio labeled. Radio labeled from a total RNA preparation extracted from each bacteria strain to be screened. Twenty or more isolates of proteus mirabilia, Klebsiella pneumoniae, Salmonella species, rhizobial species, and enterococcal species were screened by this method. Lowmolecular-weight bands (Fig. 20) indicated the presence of small labeled DNAs after polyacrylamide gel electrophoresis and autoradiography of the labeling reaction mixes. In addition, half of each labeling reaction mix was also treated with RNase A, causing a shift to a faster-migrating band, indicating that the labeled DNA is also associated with RNA. This is hallmark of the msDNA molecule as discussed above. Four of the 23 P. mirabilia isolates screened produced msDNA, while only 1 of 21 K. pneumoniae isolates and 4 of 70 Salmonella isolates screened produced msDNA. msDNA was detected in any of the 30 or so enterococcal strains screened by this method. It was concluded that the bacterial genera which contain msDNA producing retron elements are representatives of three of the four major subdivision of the purple bacteria or Proteobacteria, as described above.

In accordance with this embodiment of the invention, it is noteworthy that the discovery of msDNA extends for the first time the distribution of retron-elements to a new phylogenetic division of the purple bacteria, namely, the alpha subdivision. A collection of 63 rhizobial isolates (shown in Table 1) were screened for the presence of msDNA by the RT extension method. Among the 63 isolates, msDNA were detected in 10 (16% - Fig. 20 and Fig. 21). However, all 10 positive isolates give strong, clearly labeled bands with a typical shaft of a fast-migrating band after treatment with RNase A, indicating the presence of RNA and DNA in the labeled molecule.

so. FIFTEENTH ST. UDELPHIA, PA 191022 The 10 retron-encoding rhizobial strains include both fast growing (rhizobium) and slow-growing (1915) 875-8883

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(Bradyrhizobium) rhizobia.

The RT extension method comprises treating a preparation of total RNA, extracted from a bacterial strain to be tested, with RT from a suitable source in the presence of the deoxynucleotides dATP, dTTP, dGTP and dCTP, one of which is radiolabeled, e.g., [α-32P] dCTP, electrophoresing the treated RNA preparation on a polyacrylamide gel and determining initially the presence or absence of msDNA in the bacterium of interest by detecting a band of radiolabeled DNA corresponding to the single-stranded DNA of msDNA. Typical examples of suitable sources of RT are avian myeloblastosis virus (AMV) and Moloney murine leukemia virus (Mo-MLV). Conceivably, the test could be automated.

Total RNA samples, which contain msDNA if present in the bacterium, are extracted from the bacterial strain of interest and prepared for RT extension as follows. Total RNA, prepared from a 5-ml culture from the bacterial strain, is added to 50 μ l of a reaction mixture containing: 50 mM tris-HCl (pH 8.3); 6 mM MgCl₂; 40 mM KCl; 5 mM DTT; 1 µm dATP, dTTP and dGTP; 0.04 μM dCTP; 0.2 μM [α ³²P] dCTP; and 10 units of AMV-RT (Boehringer Mannheim). The reaction mixture is incubated at 37°C for 30 minutes, then extracted with 50 µl of phenolchloroform (1:1) and precipitated with ethanol. The samples are subjected to electrophoresis on a 4% acrylamide -8 M urea gel with appropriate nucleotide size markers, e.g., the Klenow fragment of DNA polymerase I. If the labeled sample is digested with ribonuclease (RNase) A before it is placed on the gel, a single band corresponding to single-stranded DNA is detected, which is indicative of the presence of msDNA. An aliquot from each labeling reaction mixture is treated with 5 μg of RNase for 10 minutes at 37^{0} C just prior to electrophoresis to detect in the gel a shift to a faster - migrating species, indicating that each labeled DNA is also associated with RNA, which is the hallmark of the msDNA molecule.

Low-molecular weight bands in the gel indicate the presence of small labeled DNAs after polyacrylamide gel electrophoresis and autoradiography of the labeling reaction mixtures.

Multiple bands observed in some of the lanes of the gel even after RNase treatment may be due to incomplete extension by RT during the labeling reaction, or, alternatively, multiple SUITE 500
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The Yee method for screening bacteria for the presence of retrons which synthesize msDNAs involves purifying by a conventional phenol extraction procedure total chromosomal DNA from the desired bacteria to be screened, electrophoresis on a five percent preparation acrymalide gel and checking for a satellite band. The major satellite band is cut out to extract the material in the band to quantitate the material in the satellite band. Total chromosomal DNA is subjected to acrylamide gel electrophoresis, the gel is stained with a ethidium bromide and densitometric scanning is employed to quantitate the satellite DNA against the pBR322 standard. The method is described in better details in Yee cited above.

A collection of rhizobial isolates from the United States Department of Agriculture (USDA) Beltsville Rhizobium Culture Collection are screened for the presence of msDNA by the RT extension method. This collection represents isolates at different times, from different legume hosts and from different geographic locations. msDNAs are detected in 10 isolates. All 10 positive isolates give strong, clearly labeled bands of DNA, with a typical shift to a fast-migrating band after treatment with RNase A, indicating the presence of RNA and DNA in the labeled molecule. The 10 retron-encoding rhizobial strains include both fast-growing (Rhizobium) and slow-growing (Bradyrhizobium) rhizobia as follows: Rhizobium sp. (Acacia) 3002 and 3838, Bradyrhizobium sp. (Aeschynomene) 3516, Bradyrhizobium sp. (Albizia) 3004, Bradyrhizobium sp. (Erythrima) 3242, Rhizobium loti 3468 and 3503, Rhizobium trifolii 2048 and 2065 and Bradyrhizobium sp. (Vigna) 3447. See Figure 21

Total DNA from each of eight msDNA-producing strains clearly cross-hybridizes with a <u>nod</u> YAB (1.6 - kb <u>Eco</u> RI fragment) gene probe derived from <u>Bradyrhizobium</u> japonicum, confirming that these strains are members of the <u>Rhizobiaceae</u>.

In view of the diversity of retron elements in prokaryotic populations, it is not excluded that msDNA synthesizing retrons would be found in bacteria living in alkaline environments, such as in alkaline environments: <u>Plectonema nostocorum</u>, <u>Flavobacterium spp.</u>

Agrobacterium spp. Bacillus spp. Ectothiorhodospira spp.; in acidic environments: Thiobacillus thermophilica and thiooxidans, Thermoplasma acidophilus, Sulfolobus acidocaldarius, Cuanidium

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caldarius, Bacillus acidocaldarius; in very high temperature environment (thermophilic): Sulfolobus acaidocaldarius, Caldariella acidophila, Thermus aquaticus; in very low temperature (psychrotrophic): Vibrio marinus, Pseudomonas spp., Cytophaga spp., Flavobacterium spp.; in high salt environments (halophilic): Halobacterium cutirubrum and salinarium, Halococcus morrhuae, Danaliella viridis; in high barometric pressure (like deep sea - barophilic), which are believed to inhibit the gut of ocean bottom dwelling fish. By using one of the two screening tests identified above, one skilled in the art will readily determine whether any one of these bacteria contain retrons synthesizing msDNA. This may be particularly interesting for making evolutionary comparisons between homologous RT genes present in distantly related phytogenic strains.

A representative number of amino acid sequences of representative RTs were analyzed to determine similarities and differences. The following observations were made. The amino acid sequences of these bacterial RTs are shown in Figure 14. The individual nucleotide and amino acid sequences for each of the RTs are shown in Figures 2, 11 and 15 through 19.

From a comparison of these sequences, it is noted that there are 61 conserved positions in the RT domains as indicated by solid dots at the bottom of the sequences in Figure 14. It is further noted that all bacterial RTs possess the YXDD sequence. Several other residues are conserved including the LPQS sequence that is especially common in retroviral reverse transcriptases. The RT domains are divided into seven subdomains. For each subdomain, the consensus sequences for the seven bacterial RTs can be established, as shown at the bottom of the sequences in Figure 14. There are 18 extra residues (except 26 residues for RT-Ec67) between subdomains 2 and 3, in which there is a reasonably good consensus sequence.

It has been noted that the RTs of the present invention possess a number of common conserved sequences of nucleotides and amino acid residues.

The most common conserved sequence of amino acid residues noted is as follows: tyrosine, alanine or cysteine and two aspartic acid residues. This conserved sequence, common to all RTs of the present invention, is also known as the YXDD sequence.

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A second conserved sequence of amino acid residues noted is as follows: serine, x which is a hydrophobic residue selected from the group consisting of valine, phenylalanine leucine and isoleucine, x₁ which is a polar residue selected from the group consisting of threonine, asparagine, lysine and serine and x_2 which is a hydrophobic residue selected from the group consisting of tryptophan, phenylalanine and alaning.

A third conserved sequence of amino acid residues noted is as follows: asparagine, x which is a hydrophobic residue selected from the group consisting of alanine, leucine and phenylalanine and x_1 which is a hydrophobic residue selected from the group consisting of leucine, as shown in Sec. TD No. 45552 valine and isoleucine

A fourth conserved sequence of amino acid residues further noted is as follows: x which is a polar residue selected from the group consisting of arginine, glutamic acid, lysine, valine and glutamine, a second residue which is valine, a third residue which is threonine and a fourth residue which is glycine,

These conserved sequences are only a portion of the total number of common sequences of the RTs. For other conserved sequences held in common by the bacterial RTs reference is made to Figure 14.

The RTs of the other groups of bacteria described herein as capable of synthesizing msDNAs are likewise believed to have a similar profile of conserved nucleic acid and amino acid residue sequence similarities as shown in Figure 14 and discussed above. This observation also applies to the genus Nannocystis.

In accordance with the invention, it is contemplated that prokaryotic reverse transcriptase, which is essential for msDNA synthesis, may be responsible for host cell parasitic or selfish DNA synthesis. Additionally, it is thought that the prokaryotic reverse transcriptase molecule may be essential for synthesis of biological messengers and nucleic acid enzymes.

The msDNAs synthesized by the reverse transcriptase disclosed herein possess a highly stable RNA; it is capable of self-annealing and may serve as the primer and template for msDNA O. FIFTEENTH ST. Synthesis. The reverse transcriptases (RTs) disclosed herein may be used as diagnostic agents. It is

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also contemplated that the RTs of the invention can synthesize msDNAs which will contain specific selected DNA fragments that can hybridize with complementary ssDNA, or otherwise identify ssDNAs, sought for, thus being useful as probes.

The possibility for the msDNAs to behave like restriction enzymes (or have restriction-like enzyme activity) in being capable of cleaving DNAs, or cut off a segment of itself, cannot be excluded.

The following examples are provided for purposes of illustration only and are not to be viewed as a limitation of the scope of the invention. The following examples are illustrative of bacterial isolates screened and identified to contain msDNA by way of the present invention.

EXAMPLE 1

One of the rhizobial strains, <u>Rhizobium trifolii</u> USDA 2065 is identified as containing msDNA by the RT extension method by which msDNA from total RNA is specifically labeled with ³²P as follows.

Total RNA from a 5-ml culture of R. trifolii 2065 is added to a 50 μ l reaction mixture containing: 50 mM tris-HCl (pH 8.3); 6 mM Mg Cl₂; 40 mM KCl; 5 mM DTT; 1 μ m dATP, dTTP and dGTP; 0.04 μ Md CTP; 0.2 μ M [α^{32} P] dCTP; and 10 units of AMV-RT (Boehringer Mannheim). The reaction mixture is incubated at 37°C for 30 minutes, then extracted with 50 μ l of phenolchloroform (1:1) and precipitated with ethanol. The samples are subjected to electrophoresis on a 4% acrylamide-8 M urea gel with appropriate nucleotide size markers, such as the Msp I digest of pBR322 end-labeled with [α^{-32} P] dCTP and the Klenow fragment of DNA polymerase I. An aliquot of the reaction mixture containing R. trifolii RNA is treated with 5 μ g of RNase for 10 minutes at 37°C prior to electrophoresis to detect in the gel a shift to a faster-migrating species, which indicates that the 32 P-labeled DNA extended by RT is also associated with RNA, which clearly demonstrates the presence of msDNA.

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Low-molecular weight bands in the gel indicate the presence of small ³²P-labeled DNA after polyacrylamide gel electrophoresis and autoradiography. The labeled DNA is indicative of the presence of msDNA.

EXAMPLE 2

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By the method described above in Example 1, (a) <u>Proteus mirabilis</u> 1174b is found to synthesize msDNA by the retrons containing the RT; (b) <u>Klebsiella pneumoniae</u> 912b is found to synthesize msDNA by RT; (c) <u>Salmonella</u> sp. strain SARB-3 is found to synthesize msDNA by the retrons containing the by the retrons containing the RT; (d) <u>Nannocystis exedens</u> Nael is found to synthesize msDNA by RT; (e) <u>Bradyrhizobium</u> spp. 3447, 3516 and 3004 are also found to synthesize msDNA by the retrons containing the RT.

The following method, exemplified for <u>E. coli</u>, for the isolation and purification of bacterial RT is applicable to bacteria which are screened as positive for the presence of msDNA by the RT extension <u>in vitro</u> method.

EXAMPLE 3

Isolation and Purification of Bacterial Reverse Transcriptase.

The following is a description of a convenient method for isolating and purifying a bacterial RT.

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From 10 liters of a stationary phase culture of <u>E. coli</u> strain C2110 harboring plasmid pCl-1EP5b, cells are harvested, washed in 50 mM Tris (pH 8.0), and resuspended in lysozyme buffer (50 mM Tris (pH 7.5), 10% sucrose, 0.3 M NaCl, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride). Fresh lysozyme is added to a final concentration of 2 mg/ml. The suspension is incubated on ice for 15 minutes followed by a quick freeze at -70°C, then thawed on ice. Lysis is enhanced by the addition of 2 volumes of buffer M (50 mM Tris (pH 7.0), 1 mM dithiothreitol, 0.2% Nonidet P-40,

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10% glycerol, and 25 mM NaCl) followed by incubation on ice, then a quick freeze-thaw. A cleared lysate is obtained by centrifugation at 38,000 rpm in a 50Ti rotor for 30 minutes. The cleared lysate is fractionated by ammonium sulfate precipitation (0-50%, 50-70% and 70-90%), followed by dialysis overnight (4°C) for each fraction against buffer M. Ammonium sulfate fractions, 50-70% and 70-90%, show RT activity and are pooled, then applied to a DEAE-column (2.5 x 50 cm; DE52 Whatman) equilibrated with buffer M. The DE52 column is washed, and RT activity is eluted from the column at a range of 300 to 350 mM NaCl. The DE52 fractions showing RT activity are pooled, concentrated by membrane ultrafiltration (Amicon) and then loaded onto a Sephacryl S-300 column (Pharmacia LKB Biotechnology Inc., 1.5 x 75 cm) equilibrated with buffer M. The column is developed with the same buffer. Again, fractions from the S-300 column having RT activity are pooled and concentrated, and 0.7 ml is loaded onto a 16-30% glycerol density gradient. The glycerol gradients are set up and run as described previously (Viswanathan et al., 1989). The purified Ec67.RT (fractions 7, 8 and 9) is stored as separate glycerol fractions at -20°C.

When this protocol is applied to the msDNA bacterial synthesizing strains, the respective RTs are isolated and identified as shown above.

Another convenient method for isolating and purifying reverse transcriptase is published in Lampson B.C., S. Inouye and M. Inouye, "msDNA of Bacteria", <u>Progress in Nucleic Acid</u>

Research and Molecular Biology, Vol. 40, pages 1 et seq.

The invention has been described in detail with particular reference to the above embodiments. It will be understood, however, that variations and modifications can be affected within the spirit and scope of the invention.

CLAIMS

We claim:

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- 1. An isolated and purified bacterial reverse transcriptase (RT) which is capable of synthesizing msDNA, which RT comprises a conserved sequence of amino acid residues as follows: tyrosine, x which is alanine or cysteine, and two aspartic acid residues.
- The bacterial RT of claim I which comprises a second conserved sequence of amino acid residues as follows: serine, x which is a hydrophobic residue selected from the group consisting of valine, phenylalanine, leucine and isoleucine, x_1 which is a polar residue selected from the group consisting of threonine, asparagine, lysine and serine and x_2 which is a hydrophobic residue selected from the group consisting of tryptophan, phenylalanine and alanine.
- 3. The bacterial RT of claim 2 which comprises a third conserved sequence of amino acid residues as follows: asparagine, x which is a hydrophobic residue selected from the group consisting of alanine, leucine and phenylalanine and x which is a hydrophobic residue selected from the group consisting of leucine, valine and isoleucine.
- 4. The bacterial RT of claim 3 which comprises a fourth conserved sequence of amino acid residues as follows: x which is a polar residue selected from the group consisting of arginine, glutamic acid, lysine, valine and glutamine, a second residue which is valine, a third residue which is threonine and a fourth residue which is glycine.
- 5. The bacterial RT of claim 1 which has the common subdomains 1 through 7 shown in Table 5.

- 6. The bacterial RT of claim 1 wherein the conserved sequence is located in subdomain 5 shown in Table 5.
- 7. The bacterial RT of claim 6 which has a total of 61 conserved amino acid residues.
- 8. An isolated and purified bacterial RT which comprises a sequence of amino acid residues shown in Figure 14.
- 9. An isolated and purified bacterial RT from a bacterium which is capable of synthesizing an msDNA as determined by the reverse transcriptase extension in vitro screening test, which indicates the presence or absence of msDNA in the bacterium.
- The bacterial RT of claim 9 wherein the bacterium is selected from the group of genera consisting of Myxococcus, Escherichia, Proteus, Klebsiella, Flexabacter, Cytophaga, Stigmatella, Salmonella, Nannocystis, Rhizobium and Bradyrhizobium.
- determining the presence or absence of msDNA in the bacterium comprises treating a preparation of total RNA extracted from the bacterium with a reverse transcriptase (RT) in the presence of a radiolabeled deoxynucleotide, which RT, when msDNA is present in the total RNA of the bacterium, utilizes the DNA portion of the msDNA as a primer and the RNA portion of the msDNA as a template for radiolabeling the DNA portion of the msDNA, electrophoresing the treated RNA preparation and determining the presence of msDNA in the bacterium by detecting a band of radiolabeled DNA, said band being indicative of the presence of msDNA in the bacterium.

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ABSTRACT

The present invention relates to a prokaryotic reverse transcriptase enzyme. The enzyme is capable of synthesizing a hybrid DNA-RNA molecule called msDNA with the genes which synthesize the DNA and RNA portions of the molecule.

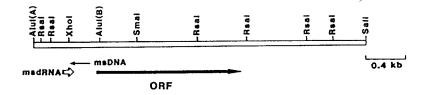


FIGURE 1

TCA TCC GCG CCG AGA CCC CCT CCT ACC TCC CCC CCG ACC CCC AGA CCC CCC TCG AGA CCC 60 TOT ACC DOG TIT COC TOG ATG GTC AGG TGG TGG CGG TGC AGT GGG GGC CGC GCA CGG GCT 120 COC CCC CTC ACC ACC CCC TCT CCT TCC ACT CCC ATG CCC AAG CCC CAC CCT ACT TCC 110 CCC CCC TCC AGA AGT TCC CCC CTC ACG CCT ACA TCC ACG CCG CCT CCC CAT TCC TCT AAA 240 CCC TTC AAC CAC GCC TCC GCC GCC ACG GCC GCC GCC GAC GAC AGC TCC GAC GAA GAG ACC 300 $\stackrel{3}{\sim}$ RNA CTC GAG CCC CCC ACC GCC CTT GCC CCC CTC CGC TTG GAA TGC AGG ACA GTC TCC GCA AGG 420 GAG CTC CCC CCC TCC CCC CAA CCC CAA CCC CAA CCC CAA CCC TAA CCC AAC CTT ACG TCC TGT GAG AGG CGT TCC TAG CCT GTT GTT GGC TCT CGC CCT AGG CAG TAG GGC CAG GGT GGG TAG GGG AGG CAA
ATC GGA CAA GAA CGG AGA GAG GGA GGA TGC GTG ATG CGG GTG CCA GCC ATC GGG TGG GTT まる AMC CAC AMC AMC AMC COC CAG COC ACC CAG COC COC COC CCC CTC AMG CCT CAG CCC CAC CAG 960 K Z K K K A Z A T Z R R A L K R Q A H E 100 CAC CTC CAC TCC CCC CAC CCC CCC CCC CAC CCC CAC CCC CCC AAC CCC TCC AAC CTC TCC AAC CTC CCC 1140 E L $_{150}^{0}$ S A E A L A K A L G L S V S K L R AND COD GAC GGC AGC AGC COC AGC ATT AGG TCC CCC AAG CCT GAG CTC AAC CCA GCC GAC 1260 K R D G S K R T I T S P $\frac{K}{200}$ P E L K A A Q CTC ACC TTC TOC TOC ACC AAC CCC AAC CAC CCC AAC CCC ACC CCC CC AAT GCG GGG GGC AAG CAC GCG GCG GCG GCC GCA GTC GCG GCC GAC GTC GTC GCC CAC GTC 1920 N A A C K D A P A A R V P R D V V R Q L CTC GCT CAG CTC AGG CAG CTC CAG TCC AGG CCG AGG CCG CCT CCG CAG CCG CAG TCA CGC 2100 L A Q L T E L E S T A S A A P Q A E CAG CCG CCG CGG GTA C

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VKLKPCHDGPKVKQ WPLTEEKIKALVEICTEMEKEGKISKIGPENPYNTPVFAIKKKDSTKWR
RPWARTPPKAPRNQ PVPFKPERLQALQHLVRKALEACHIEPYTG PCNNPVFPVKKA NGTWR
NVLYRIGSDNQYTQFTIPKKGKGVRTISAPTDRL KDIQRRICDLLSDCRDEIFAIRKI SNNYS
HIV
HTLV1
Ec-67
           RT
RT
                   AFHREVDTATHYVSWTIPKRDGSKRTITSPKPEL KAAQR WVLS
                                                                                                    NVV ERLP VHGAA
                    KLVDFRELNKRTQDFWEVQLGIPHPAGLKKK KSVTVLDVGDAYFSVPLDEDFRKYT
FIHDLRATNSLTIDLSSSSPGPPDLSSLPTTLAHLQTIDLRDAFFQIPLPKQFQPYF
FGFB RGKSIILNAYKHRGKQIILNIDLKDFFESFNFGRVRG YFLS NQDF
HIV
HTLV1 RT
Ec-67 RT
                    HGFV AGRSILTMALAHQGADVVVKVDLKDFFFSVTURRVKGLLRKGGLREGTSTLLSLLSTEAP
Mx-162 RT
                   FTIP SINNETPGIRYQYNVLPQCWKGSPAIFQS SMTKILEPFKKQNPDIVIYQ<del>YMDD</del>LYVC FTVP QQCMYGFGTRYAWKVLPQGFKNSPTLFEM QLAHILQPIRQAFFQCTILQYMDDILLA LN PVVATTLAKAACYN GTLPQGSPCSPIISNLICNIHDMRLAKLAKKY GCTYSRYADDITI REAVQFRCKLLHVAKGP RALPQGAPTSFGITNALCLKLDKRLSALAKRL GFTYTRYADDLTF
HTLV1 RT
Ec-67 RT
Mx-162 RT
                   HTLV1 RT
Ec-67 RT
Mx-162 RT
                    SWTVNDIQKLVGKLNWASQIYP
нтν
           RT
HTLV1 RT
                    RWALPELQALLGEIQWVSKGTP
Ec-67 RT
                    RCYYKKTRALAHALYRTGE YK
Mx-162 RT
                    PAARVPRDVVRQLRAAIHN RK
```

FIGURE 3

A

Mx-162

Mx 162	92	KAEATERRALKRQAHEAW-KATHVGHLGAGVHWAEDRL	128	
Mo-MLV	1143	TVLLTTPTALKVDGIAAWIHAAHVKAADPGGG-PSSRL 0 000 0 0 0 0 0	1179	
В				
Mx-162	411	${\tt GKDAPAARVPRDVVRQLRAAIHNRKKGKPGREGESLEQLK}$	GMAAFIHMTD-PAKGRAF-LAQLTELESTASAAPQAE	485
HIV	396	GKEGHSARQCR-APRRQGCWKCGKPGHIMTNCPD-R	-QAGFLGLGPWGKKPRNFPVAQVPQ-GLTPTAPP	461

(A) Sequence similarity of the region from residues 18 to 128 of the msDNA Mx162 RT (see Figure 2) with a carboxy-terminal region of integration

(B) Comparison of the sequence from residues 411 to 485 of the msDNA-Mx162 RT (see Figure 2) with the sequence from residues 396 to 461 of

18 PTPELTAPSSDAAAKREARRLAHEALLVRAKAIDEAGGADDWVQAQLVSKGLAVEDLD-FSSASEKDKKA-WKEKK 91

MO-MLV 1070 PDPDMTRVTNSPSLQAHLQALYLVQHEVW-RPL-AAAYQEQ-LDRPVVPHPYRVGDTVWVRRHQTKNLEPRWKGPY 1142

protein of Moloney murine leukemia virus (M-MuLV) (residues 1070 to 1179, Shinnick et al., 1981)

the $\it gag$ protein of human immunodeficiency virus (HIV, Ratner et al., 1985)

FIGURE 4

Α Mx-162 304 GP-RALPQGAPTSPGITHALCLKLDKRLSALAKRL-GFTYTFYADDLTF-SWTKAKQPKPRRTQRPPVAVL 371 Ec-67 Ec-86 HIV 150 YAWKVLPQGFKNSPTLFEM---QLAHLQPIRQAFPQCTILQYMDDILLAS--PSHEDLLLLSEATMASLI 215 HTLV1 303 LTWTRLPQGFKNSFTLFDE---ALHRDLADFRIQHPDLILLQYVDDLLLAA-TSELDCQQG-TRALL-QTL Mo-MLV 141 FQWKVLPQGHTCSPTICQL---VVGQVLEPLRLKHPSLCMLHYHDDLLLAA--SSHDGLEAAGEEVI-STL RSV 122 FAWRVLPQGFINSPALFER---ALQEPLRQVSAAFSQSLLVSWHDDILLYAS--PTEEQRSQCYQALA-ARL 186 BLV Mt.plasmid 288 IATHGVPQGASTSCGLATYNVL-----ELFLRY--DELIMYADDGIL-CRQDPSTPDFSVEEAGVVQEP 348 339 YEYLRHPFGLKNAP-ATFORCHN-DI----LRPLLINKHC-LVKLDDIIVFS-TSLDEHLQSLGLVFE--KL 399 17.6 GYPSY 284 YEFCRLPTGLRNASSIFQR---ALDDV---LREQI-GKICYVTVDDVIIFS--ENESDHYRHIDTVLK-CL 344 1032 CKLNKAIYGLKQAARCHFR-CIYI---LDKGNINENIYV-LLYVDDVVIAT--GDHTRHNNFKRYLME-KF 1112 Copia 990 CLLKKSLYGLKQSPRQWNA-CVYV-KQVSE-QEHLYL---LLKVDIMLIAG--KSKSEINKVKEQLSM-EF 1069 Tal-3 948 IRLKKSLYELKQS-GANWYE--EVRG-WSCVFKNSQV-TICLFVDLMVLFS--KNLNSNKRIIEKLKM-QY 1023 Ty912

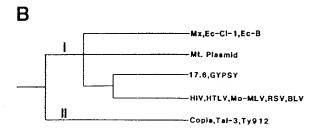
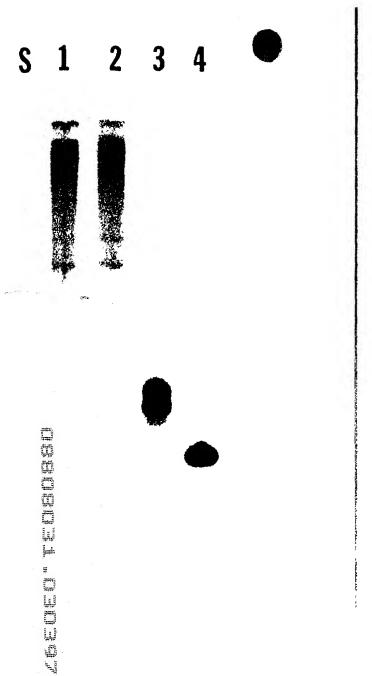


FIGURE 5



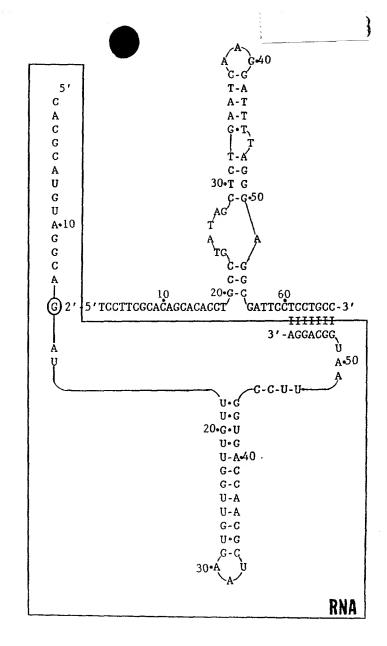
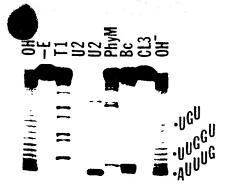
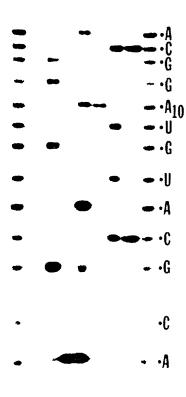


FIGURE 6

FIGURE 7





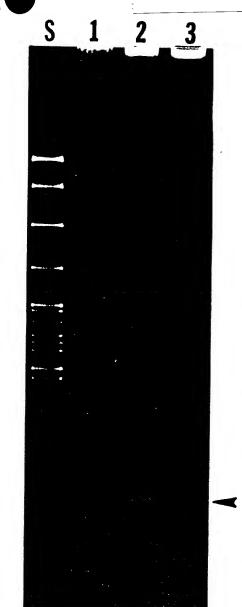
 $\begin{smallmatrix}&&&&&&&&&&&&&\\1&2&3&4&5&6&7&8&9\end{smallmatrix}$

23.0- ***

9.4 -6.6 -

4.4-

2.3-**2.**0-



FIGU-

FIGURE 9

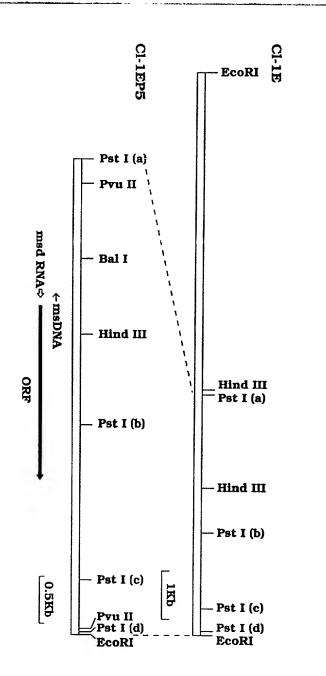


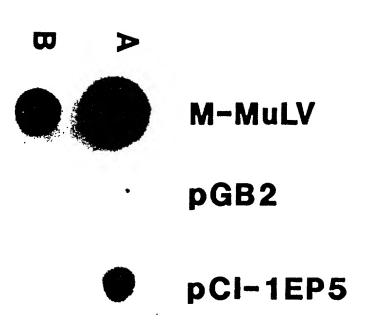
FIGURE 10

NG GAT COT CTT TGC TCA GAT CGG CCA GAA CTG GCG GGC TTT TGC TCA 120 TOT CAT GCA TGT GCA TGA AAA CCA CTG CAT AAA GCG GGC AGG CGT GGC GGG GAT AGG AGC 180 **□**NHA TIC ATA AND AGG CAT GTA GGG AGA TTT GTT GGT TGT GAA TCG CAA CCA GTG GGC TTA ATG ANG TAT TTG TGG GTA CAT GGG TGT ANA CAA CCA ACA CTT AGG GTT CGT CAC CGG AAT TAG GCA GGA GGA ATC GCC TCC CTA AAA TCC TTG ATT CAG AGC TAT ACG GCA CGT GTG CTG TGC 360
GGT GGT TAG GGG AGG GAT TTT AGG AAG TAA GTG TGC ATA TGC GGT CGA CAG GAG AGG ACA AAA ACA TCT AAA CTT GAC GCA CTT AGG GCT CGT ACT TCA CGT GAA GAC TTG GCT AAA 480 T K T S K L D A L R A A T S R E D L A K ATT TTA GAT ATT AAG TIG GTA TIT TTA ACT AAC CTT CTA TAT AGA ATC CGC TCG CAT AAT 340

1 L D I K L V F L T N V L Y R I G S D N CAN TAG AGT GAN TIT AGA ATA GCG AAG AAA GGG AAA GGG GTA AGG ACT ATT TGT GGA GGT Q Y T Q F T 1 P K K G K G V R T I S A P $^{+}$ GAG ATC TIT GGT ATA AGG AAA ATT AGT AAC AAC TAT TCC TIT GGT TIT GAG AGG GGA AAA 720 E I F A I R K I S H H Y S F G F E R G K C X TCA ATA ATC CTA AAT CCT TAT AAG CAT AGA CGG AAA CAA ATA ATA TTA AAT ATA CAT CTT 780 S I I L N A Y K H R G K Q I I L N I D L AAG GAT TIT TIT GAA 7 AGC TIT AAT TIT GGA CGA GTI AGA GGA TAT TIT GTI TCC AAT CAG X D F F B S F N F G R V R G Y F L S N Q GAT TIT TTA TTA AAT CCT GTG GTG GGA ACG ACG CTT GCA AAA GGT GCA TGC TAT AAT GGA 900 D F L L N P V V A T T L A K A A C Y N G 2150 GAT ATG AGA TTA GCT AAG CTG GCT AAA AAA TAT GGA IGT ACT TAT AGC AGA TA<u>T GCT GA</u>T 1020 D M R L A K L A K K Y G C T Y S R <u>C A D</u> GAT ATA AGA ATT TCT AGA AAT AAA AAT AGA TIT CCG TTA GAA ATG GCT ACT GTG CAA CCT 1080 CAA GGG GTT GTT TTG GGA AAA GTT TTG GTA AAA GAA ATA GAA AAG GTT GGA TTG GAA ATA 1140 E G V V L G K V L V K E I E N S G F E I AAT CAT TCA AAG ACT ACG CTT ACG TAT AAG ACA TCA ACG CAA CAA CTA ACG CGA CTT ACA 1200 N D S K T R L T $\frac{\chi}{2250}$ K T S R Q E V T G L T CAA TAT AAA GTG CCA GAT CAA AAT GGT GTT TTA GTT TCA GGA 1320 E Y K V P D E N G V L V S C $^{\circ}$ CCT CTG GAT AAA CTI GAG GGG ATG TIT GGT TIT ATT GAT CAA GTT CAT AAG TIT AAC AAT 1380 G L D K L E G H F G F I D Q V D K F H N ATA ANG ANA ANA CTG ANC ANG CAN CCT GAT AGA TAT GTA TTG ACT ANT GCG ACT TTG CAT 1440 I K K K L N K Q P D R Y V L T N A T L H CGT TIT AMA TTA AMG TIG AMI GCG CGA GAA AMA GCA TAI AGT AMA ITT ATT TAC TATA AMA 1500 G F K L K L H A R E K A Y S K P I Y Y K 2 TTT TTT CAT GCC AAC ACC TCT CCT ACG ATA ATT ACA CAA F F F H G N T C P T I I T E TTG TITT AGA GAA AAA ACA L F R E K T gag E TIT THA GAT CIT TOT GGG GGA ACT GCA ACT GCA AAA AAA TIT GTA GAG CGT TAT AAA AAA I 1740 F L D L S G G T A D L X X F V E R Y X N AAT TAT GCT TCT TAT TAT GCT TCT GTT CCA AAA CAG CCA GTG ATT ATG GTT CTT GAT AAT 1800 M Y A S Y Y G S V P K Q P V I M V L D M $^{\rm AA}$ V ₽ *450 GAT ACA GGT CCA AGC GAT TTA CTT AAT TTT CTG CGC AAT AMA GTT AAA ACC TGG CCA CAC 1860 D T G P S D L L N F L R N K V K S C P D TTA TAT ATA GTT 1920 L Y I V *500 CTC AGA CGA TTG AGT CCT TCC GGC GAA CAA AGT TCA ATG CAG GAT CTT TTC CCT AAA GAT 1980 L T P L S P S G E Q T S H E D L F P K D CAT TIT AMG CCA TIT TGT TGT ATT TTT CAT CCT ATA AMA CAT ATA AMG CAA CAT TAT AMA 2160 D F K A F C C I F D A I K D I K E H Y K TTA ATC TTA AAT AGG TAA TGA AGA GCC CTA AGG TTA TGA AGG CTA AGG CTG ATT TTT CCT 2220 TAA AAT TTA TAT GGT TTG AAT IGT AAT ATA TTA TCT TCA AGC CAT TTA TTT AAT TCC TCC 2280 ATC CTT TIC TOT AMO GOT ATT AMT TOG TTC CTC ACA AMC ACT AMA CTC GOT TIT TOG ACA 2340

CGC TGC CAT CAT GTC ATG GCG GC

V	RT	VKLKPGMDGPKVKQ WPLTEEKIKALVEICTEMEKEGKISKIGPENPYNTPVFAIKKKDSTKWR	239
LVl	RT	RPWARTPPKAPRNQ PVPFKPERLQALQHLVRKALEAGHIEPYTG PGNNPVFPVKKA NGTWR	75
DNA	RT	NVLYRIGSDNQYTQFTIPKKGKGVRTISAPTDRL KDIORRICDLLSDCRDEIFAIRKI SNNYS	94
		+ 0 • • 0 + • + • +	
v .	RT	KLVDFRELNKRTQDFWEVQLGIPHPAGLKKK KSVTVLDVGDAYFSVPLDEDFRKYTAFTIP SI	302
LV1		FIHDLRATNSLTIDLSSSSPGPPDLSSLPTTLAHLQTIDLRDAFFQIPLPKQFQPYFAFTVP QQ	139
DNA		FGFE RGKSIILNAYKHRGKQIILNIDLKDFFESFNFGRVRG YFLS NQDF LLN PVVA	
		0 • + •+ + +0 +• +•	130
V	RT	NNETPGIRYQYNVLPQGWKGSPAIFQS SMTKILEPFKKQNPDIVIYQYMDDLYVGS DLEIG	363
LV1	RT	CNYGPGTRYAWKVLPQGFKNSPTLFEM QLAHILQPIRQAFPQCTILQYMDDILLAS PSHE	199
ANG	RT	MMT 1771 1 OUR AMERICA	212
		· • • • + + 0 + 00 • • • 0	
¥	RT	QHRTKIEELRQHLLRWGLTTP DKKHQKEP PFLWMGYELHPDKWTVQPIVLPE KDSWTVNDI	424
LVl	RT	DLLLLSEATMASLISHGLPVS ENKTQQTPGTIKFLGQIISPNHLTYDAVPTVPI RSRWALPEL	262
ANC	RT		276
		0 + • • • • + +•	•
V	RT	QKLVGKLNWASQIYPGIK VRQLCKLLRGTKALTEVIPLTEEAELELAENREILKEPVHGVYYD	497
LVl	RT	QALLGEIQWVSKGTPTLRQPLHSLYCALQRHTDPRDQIYLNPSQVQSLVQLRQALSQNCRSRLVQ	327
ANG	RT	RALAHALYRTGE YKVPDE NGV LVSGGLDKLEGMFGFIDQVDKFNNIKKKLNKQ PDRYVL	
. :		0● + + + + + + 0 + 00 ● 0+0	
V	RT	PSKDLIA EIQKQGQGQWTYQIYQE PFKNLKTGKYARMRGAHTNDVKQLTEAVQKITT	544
LV1			
DNA	RT	TNATLHGFKLKL NAREKAY SKFIY YKFFHGNTCPTIITEGKTDRIYLKAALHSLET SYPEL	396
		0 • 00 + 00 + 0 0 + + + 0 + 00 0	
V	₽RT	ESIVIWGKTPKFKLPIQKETWETWWTEYWQATWI PE WEFV NTPPL VKLWYQ	595
LVl		LCQTIHHNISTQTFNQFIQTSDHPSVPILLHHSHRFKNLGAQTGELWNTFLKTAAPLAPVKALMP	456
DNA	RT	FREKTDSKKKEINLNIFKSNEKTKYFLDLSGGTADLKKFVERYKNNYASYYGSV PKQPVIMVLD	
		+ + 00 + 0 0 • 0•	
J	RT	LE KEPIV GAETFYVDGAANRETKLGKAGYVTNKGROK VV PLTNTTNO KTELOAIYLA	650
LV1	Contract Con	LE KEPIV GAETFYVDGAANRETKLGKAGYVTNKGRQK VV PLTNTTNQ KTELQAIYLA VFTLSP VIINTAPCLFSDGSTSRAAYILWDKQILSQRS FP LPPPHKSA Q RAELLGLLHGL	
DNA		NDTG PSDLLN FLRNKVKSCPDDVTEMRKMKYIHVFYNLYIVLTPLSPSGEQTSMEDLFPKDIL	
30.00		0 • 0 + + + 0+0 +0 • 0 • 0	<i>J</i> 2 <i>J</i>
Į	RT	LQDS GLE VNIVTDSQYAL QIIQA QPDKSESELVNQIIEQLIKKEKVYLAWVPAHKG	708
	RT	SSAR SWR CLNIFLDSKYLYHYLRTLALGTFQGRSSQAPFQA LLPRLLSRKVVYLHHVRSHTN	578
DNA	RT	DIKIDGKKFNKNNDGDSKTEYGKHI FSMR VV RDKKRKIDFKAFCCIFDA	572
		+ • ••• • • • • • • • • • • • • • • • •	·
Ţ	RT	IGGNEQVDKLVSAG	722
Νl		_	592
NA	RT		586



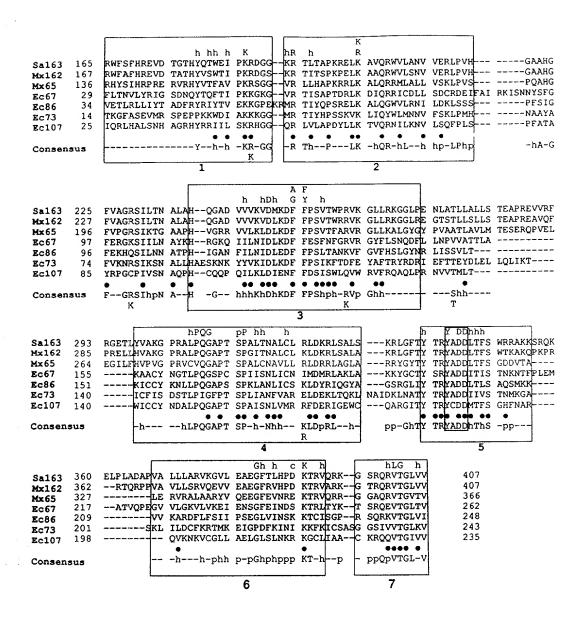
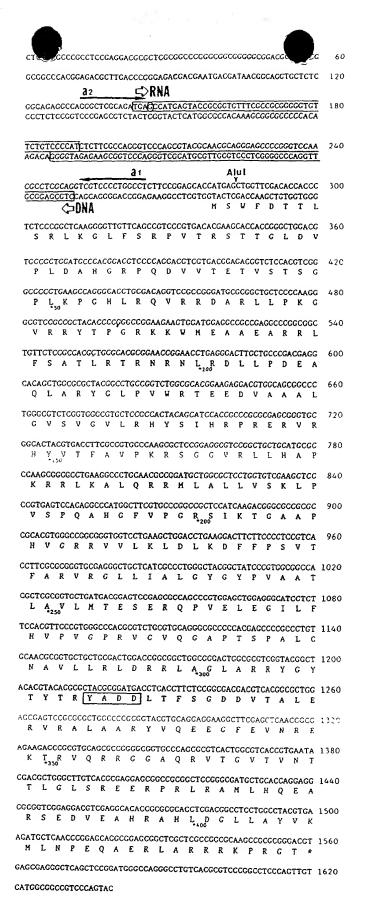


FIGURE 14



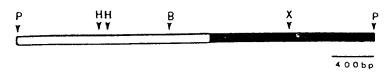
OC ACT TOO GGC GCT CGG GCT GCG CGA GGG CCC GTG CGA GCA CAT GAT GGC GCT GCG GCT GT COA GGT CCG GCA CCG CGC CGA GCA GGA AGC ACT GCG TCA GAC CCC CGC GGG CCG CCA 120 180 CT CAT CCG CGC GGA GAC GCG CTC CTA CGT GCG GCG CGA GCC CTC CGG CCA GGA GCA GCT TA COG COT CTC ATT GOA TOG GAA AUT GOT GGC GOT GGA GTG GGG CCC CCG CCA GGG GGA 240 TO COG COG GOA GAA GOT CTG GTT CGA CAC GGA CGC CGA GGC GOG CAC CGC CTA CTT CAC 300 OG OCT GGA GTC CTT GGC CGC GGA GGG ATA TAT CGA TGC GGC TGC TTC AAT GAT GTA GAA 360 420 AC GCA AGO CAO GGG GCC GGG GGC GGG CGG CGG AAA GGC AGG TGC GAC GGA ACG ACA GAC RNA T COT GOG AGC GAC CGA GAG AGG TOC CAA GCC ATC AGC CTC AGC GCC TOC AGC GCC AGA GAC AGC GCC TCC AGC GCT CTC TCC AGG GTT CGC TAG TCG GAG TCG CGC AGC TCG CCC TCT 480 540 T CTC TTC CCT CCG GTG ACT ACC TCT CCG GCC GGG GAG CTG AAC CAA CGA CGC AAC CGC ACC CGG GAG AAG GGA GGC GAC TCA TGG AGA GGC GCG CCC CTC GAC TTG GTT GCT GCG TTG GGG THE COO GGC CGG AGA GGT ACT CAC CGG AGG GGA GAG COG GTG AGG CTA CGG TGC CCC
A AAG GGG CCG GCC TCT CCA TGA GTG GGC TCC CCT CTC GGC CAC TCC GAT GGC AGG GGG 660 21 720 840 900 TC GAA GAE GCG GGC GGC ACG GAC GCC TGG GTG CGG CAG CAG CTG GTG GCC AAG GGC GTC L E A G G T D A M V R Q Q L V A K G V CG GGG GER GAG GTG GAC TTC GAG TCG CTC AGC GAC AAG CAG AAG GCC GCC TGG AAG GAG
A A 187 E V D F E S L S D K Q K A A W K E 1020 AG ANG ANG GOC CAG GOC ACC GAG CGG CGG CGG CAG ANG CGC CTG GCG TAG GAG K K K K A E A T E R R A Q K R L A W E 1100 AG GCC $\stackrel{\bullet}{AGG}$ CAC ATC CAC CAC CTG GGC GTG GGG GTG CAC TGG GAC GAG GGC GGA GGG CCG $\stackrel{\bullet}{K}$ A $\stackrel{\bullet}{T}$ $\stackrel{\bullet}{H}$ $\stackrel{\bullet}{I}$ $\stackrel{\bullet}{H}$ $\stackrel{\bullet}{I}$ $\stackrel{\bullet}{H}$ $\stackrel{\bullet}{I}$ $\stackrel{\bullet}{K}$ $\stackrel{\bullet}{$ TTC GAC GTG GCC GGG GGC GAG GAG CGG GCC AAG GCC AAC GGC TTG CCG GAG GGG P D V A G R E E R A X A X G L P E G TO TOO THE CAC COC CAG GTG GAC ACG GGC ACG CAC TAC CAG ACG TAG GAG ATT CCG AAG
F S F H R E V D T G T H Y Q T W E I P K GG GAC GOC GGC AAG CGG ACG CTC ACC GCG CCG AAG CGG GAG CTC AAAG CCC GTG CAA CGC R D G G K R T L T A P K R E L K A V Q R GG GTG CTC GCG AAC GTG GTG GAG CCG CTG CCG GTG CAC GGG GCC GCG CAC GGC TTC GTG

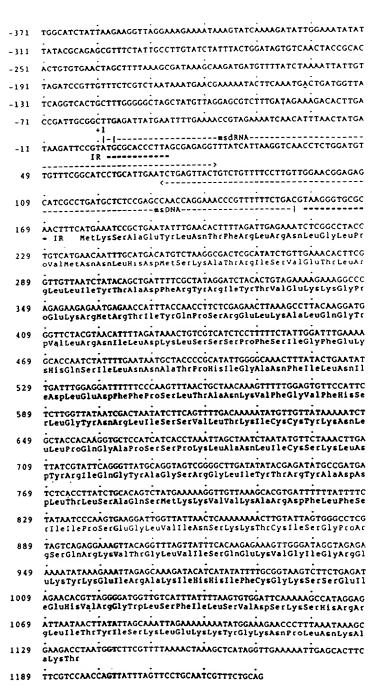
AMS GGA GGA CTC CCG GAG AMC CTG GGG AMG CTC CTG GGG CTG CTC TCC ACC GAG GCC CCG K G G L P E N L A T L L A L L S T E A P CGC GAG GTG GTG CGG TTC CGG GGA GAG ACG CTG TAC GTG GCC AAG GGC CCT CGC R E V V R F R G E T L Y V A R G P R CCC CAG GGG GCC CCC ACC TCT CCG GGG CTG ACG AAC GGG CTG TGC CTG CGG CTG GAC AAG P T S P A L T N A L C L R L D X 1800 ACG TTC TCC TGG CGG CGG GCG AAG AAG TCC ccc gre goe ete ete ese cse gre aag get gre ete gag goe gag get tie ace ete p v a l l l a r v r g v l e a e g f t l CAC CCG GAC AAG ACG CGG GTG CAG CGC AAG GGC AGC CGG CAG CCG GTG ACG GGG H P D K T R V Q R K G S R Q R V T G CTG CGC GCG GCG ATC CAC AAC CGG GAG CAG GGC AAC GCC GGC CCC ACC GGG GAG ACG CTG GAG CAG CTC AAG GGG CTC GGG GCC TTC CTT CAC ATG ACG GAC GGG GAG AAG GGC CGC GCC E Q \mathbb{C} \mathbb{C} 2160 TTC CTG CGA CGG CTG GAG GCC CTC GAG AAG CGC CAG ACC GCC TGA CCC TCA CTG GTC GTC F L R R L E A L E R R Q T A \sim CGG GGC ATC GCA GCG GGC GCC GGG ACG GAC CGT CAC CCC CCA GAT CTC CAT GCC ATG CTG 2280 GGG ATT CTG GGC GGT GAA GAA GAC TTC CCA GCC GAG ACG GAC GAA GCC CTG CGG ATC CGA TGA CTC CTC GCC CGG GGC GAT CTC CCG GAG GGG CAC CGT TCC GAC GTC CGT GCC ATT GCT CAC CCA GGG CTC CCG GCC CCA GCC TTG GGT GTC CGC CGA GAA GAA GAG CAG CCC GGA GAT CGC CGT CAG GTT CTC CGG CGA CGC ATC CTC GGG GCC CGG CGC CAA ATC CTT CAG CAG CAG GGT GCC CTT GGC GGT GCC ATC GCT GGA CCA CAG CTC CCG GCC GTG GAG GCT GTC ACT CGC GGC GAA GTA GAG CAT CCC ATT CAG CGC CTT GAT GGC GCT GGG CGC CGA GCT GTC CGG ACC COG CCA GAT GTC CTT CAC CCG GAC CGT GCC ATG CGA CGT GCC ATC GCT GAC CCA CAG CTC CCC CCC GAG CGC CGT GAG ATC ATG CGG ATA GAG GCC GGG GAA GAA GCG CAG CTG CTC GGA GAC GOT GOD TOT GGA GOA COA CAG GOT GGC CTC GCC TTC GTC ATT GTC GAG CAG GAA GAA CAG CAC COA GTC CGC CGC GGT GAA CGC GGA GAG GAA GTT GTC CTC GGG GCC CGT GAA GAC AGA COT GOT GOT GOA CAG COC CAG GOT GOG COA GAT GAA CAC CTC GTC ATT GAC GTT GGC CAC GAA GAA GAG CGC ATC GCC GAC CCG GGT GAG CCG GCG CGG GCT GGA GCT GCC GGG CAC

TTTTGAGAAG CGCCATACCA AACAGGGGAT ACAGACCAAC CTGACGCTGA AAGAGGALAG CTACCGCGAC TGCCTGCCGA AGTGCGACCA F \mathbb{R} \mathbb{R} CCCCCCAGCA ACATAMCCTC ACTCAGACCG GCAACAGCCG GTCTTTTCCT TTCTGGCCAT TGCCCACAGG TGAACAATCC ACTCTTGACC 10080 CTTCACCOTT TATTCACCCT TTATCACTAT GALLITATTA ATLLULACE AGAGGTELIC ACTUTALCA GTALLICCTG ALLIACTTI 10170 TTATCACCCC GOGCATOGGC CGACTOGACA GATCCAGAAC GAGCAAAAAT CACAAAGGTG ACGAGTCGAC TOTTCACTACT TCACCAACTC 10240 THATCH ACCACATE ATTAINING TAMPIATES ACCITATED THATCH AMERICAN THATCH AMERICAN TOTAL TAGGCCATCO COCATGAGTC ATGGTTTCGC CTAGTATTTI AGCTNICGCC CTCGTTCAGT TCGCTGAGCG CCCGCTGGGG GCCACCCGATC 10530 ATGGGGTAGC GCCTACTCAG TACCAAAGCG CATCATAAAA MCGATAGGGG CAGCAAGTCA AGGGACTGG CGCCGACCCC CGGTGGCTAG ACCUMENTAL TOGACTICS CALCAGAMA TOGACTITI MATECTATAC CATALAGATA TICTOCATCE TRACTCACT 10410
TCGCTTGACT ACCTGCACA ACCCACAMA ACCCACAMA TCGCACAMA TCGCA AMMANAT GGTACTGAGG TATCTAGAGC ARCCCGGTTA TTTTCATCAT TCGTTGANA GALCALAGTA AMTGTCCTG GTATGTAAA 10710 K K H G T E V 8 R A T A L P 8 8 F V E K K V K C P G H V K AMARTICUTC TITCTUTUTU GIOCTAACAA MACAATOGA GAACCATCAG CAAGACGATT GGAATTAATA AMITITETU AAGGTATTI 10800 K P V P L C G A K K K K G E P E A R R L E L I K P S E R_{FV} L GARTARCTOT CACTITITIC ITSCIGNACT ASTITICANA GARTRANGCA COCATGAIGA ATCATTATCT GATARTTAT TAGATATCGA 10190 N N C H F F L A E L V F K E L S T D E E S L S D N L L D I E AGCTGACTTA TCTANATTAG CTGATCATAT TATCATTGTT TTAGANAGTT ATTCATCTTT CACGGACTT GGTGCATTCG CATACAGCAA 10980
A D L S K L A D H I I I V L E S Y S S F T E L G A F A Y S K GCARTTACGC AMGARATTAN TANTAGTTAN CAMTACANA TTTATANATG AGRANTCATT TATANATATG GGRCCANTAN AGGCTATTAC 11070 Q L R K K L I I V N N T K F I N E K S F I N N G P I K A I T TCAGCAATCA CAACAATCTG GTCATTTCTT ACATTATAAA ATGACAGAAG GTATTGAAGG TATAGAGCOC TCTGATGGGA TTGGGGAATT 11160 Q Q S Q Q S G H Y L H Y K H T E G I E S I E R S D G I G E I ATTCGACCCC CTATATGATA TICTTTCTAA GAACGACAGA GCAATTICAA GAACTITAAA AAAAGAGG TTAGATCCTI CCAGTAACTI 11250 F D P L Y D I L S K N D R A I S R T L K K Z E L D P S S K F CANTANAGAC TEAGTACGAT TEATTCATCA COTABITETT GEATGTOGGC CTTTGCAACT TANTGAACTC ATCGAATTA TEACALLAT 11340 N K D B V R F I H D V I F V et C G P L Q L N E L I E I I T K I ATTIGGACA GAMGCCATT ACAMMMA TETTETAMAG CACTTGOTA TECTAATAGE TATTAGATA ATATCATGA CAMATGGGAT 11430
F.G. T. E. B. Y. K. K. L. L. K. H. L. G. I. L. I. A. I. R. I. I. S. C. T. H. G. I. TTATTATTCT TTGTATALIG ANTATTATT TALATATGAC TTGACATTG ACALCATATC ATCALTGTT ALAGTITIT TCCTCAAGAA 11520
Y Y S L Y K Z Y Y F K Y D F D I D W I S S W F K V F F L K N CAAGCCAGAA AGGATGAGGG TATATGAGAA TATATAGCCT AATTGATTCT CAGACATTGA TGACTAAGGG ATTTGCTTCT GAAGTAATGC 11610

K P I R N R V Y Z N I *

RT N R I Y S L I D S Q T L N T K G F A S E V N GATCACCTGA GCCGCCAAA AAATGGGATA TAGCTAAGAA AAAAGGAGGT ATGAGAACAA TTTATCACCC GTCATCAAAA GTTAAATTAA 11700 R S P P K K W D I A K X K G G W R T I I Y H P S S K V K L TTCAATATTG GTTAATGAT AATGITTTTT CGAAGCTCCC AATGCATAAT GCTGCATATG CATTTGTTAA AAACCGATCA ATAAAAAGCA 11790 I Q Y W L N K N V F S K L P N N N N A A Y A F V K N R S I K S ATGCTTTATT ACATGCCGAA TCAAAGAATA AGTATTATGT GAAAATAGAT CTCAAAGATT TTTTCCCTC AATAMATTT ACTGATTTG 11840 M A L L M A Z S K M K Y Y V K I D L K D F F P S I X F T D F AGTACGCATT CACTCGTTAT CGAGATCGCA TTGAATTTAC TACAGAATAT GATAAGGAGT TACTACAACT TATAAAACG ATCTGCTTTA 11970 E Y A F T R Y R D R I E F T T E Y D K E L L Q L I K T I C F TATCAGATAG CACTCTCCCT ATCGGGTTTC CTACATCTCC ATTANTIGCA AACTTTGTGG CAAGAGAACT TGATGAAAAA CTGACGCAAA 12060 I S D S T L P I G F P T S P L I A H F V A R Z L D E K L T Q AACTAAATGC AATTGATAAA CTTAATGCCA CTTATACACG ATATGCTGAT GATATTATTG TCTCTACAAA TATGAAAGGG GCTAGCAAAT 12150 K L N A T Y T R Y A D D I I V S T N N K G A S K TRANSFERGE TEGTETTARA AGRACANTGA ARGAGATTGG TECAGACTET AMARTARACA THARRATT TRAGGETTECT AGGETTECT L I L D C F K R T K K E I G P D F K I W I K K F K I C S A S GAGGAAGTAT AGTAGTTACC GGATTGAAAG TITGCCACGA TITTCATATT ACATTACATA GATCAATGA AGATAAAATA AGATTGCATC 12330 G G S I V V T G L K V C H D F H I T L H R S N R D N I R L H THICHTHIT ATCHARGGC ARATTARANG ATGRAGATCA TAMPARATIT TURGGTTATA TUGGTTATGC ARAGATATA GACCUCCATT 12420 L S L L S K G I L K D T D K M K L S G Y I A Y A K D I D P H TTTATACAAA ACTGAACAGA AAATATTTC AAGAATAAA ATGGATTCAG AATCTCCACA ACAAAGTTCA ATAACTTTA TATTTTCGAT 12510 F Y T K L N R K Y F Q E I K M I Q N L N N K V E $^{\circ}$ GCACCCCAAT AACTTCATTG ATTAAATTGG GAACAATATA GGCTTTTCAG GATGACCTAC ACTCTAGAGA ATGTGTATAC AAAAGTGTAT 12600 AAGTTATTTT CAAACCTATA TAAAATACAG CAAATCAAT GCATTGGCGG CATTTTACOA CTCCTGTGAT CTTCCGCCAA AATGCCTG

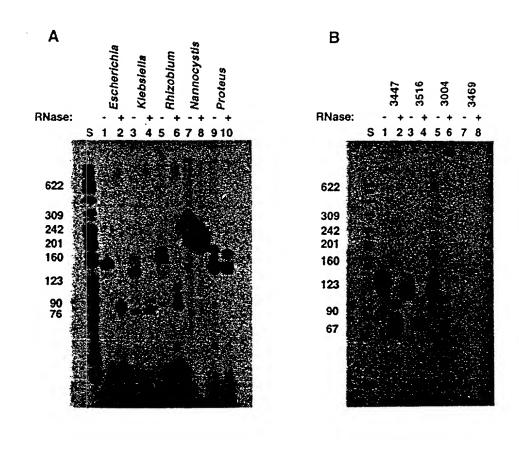








	60
cggcggttaaggcctatcgcgaagagttcggcgtttaaaAATATGCCCTGTGCAGGGTTT	120
TTGCTGTGCGCAGCGTGATGCGCTTCAAGATATCGTGTTAATCTGCTTT <u>GCCAGCAGTG</u> AACGACACGCGTCGCACTACGCGAAGTTCTATAGCACAATTAGACGAAAAGCGGTCGTCAC	180
CCANTAGEGTTTCCGGCCTTTTGTGCCGGGAGGGTCGCGAGTCGCTGACTTAACGCCAG	240
TAGTATGTCCATATACCCAAAGTCGCTTCATTGTACCTGAGTACGCTTCGCGTACGTCGC ATCATACAGGTATATGGGTTTCAGCGAAGTAACATGGACTCATGCGAAGCGCATGCAGCG	300
GCTGACGCGCTCAGTACAGTTACGCGCCTTCGGGATGGTTTAATGGTATTGCCGCTGTTG CGACTGCGCGAGTCATGTCAATGCGCGGAAGCCCTACCAAATTACCATAACGGCGACAAC	360
GCGCCTCTTTTGGCCGCCGTGATGTGGAGAGTGGAATGGATGCTACCCGGACAACCCTTC M D A T R T T L L	420
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	480
ATGCGCTCAGTAATCATGCCGGACGCCATTACCGACGCATTATTCTTTCT	540
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	600
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	660
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	720
TCGAAAACTTTTTCGATAGCATTAGCTGGTTACAGGTCTGGCGTGTGTTTCGCCAGGCCC E N F F D S I S W L Q V W R V F R Q A Q	780
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	840
CGCAGGGGGCACCAACTTCGCCAGCCATTTCCAATCTTGTGATGCGCCGTTTTGATGAAC Q G A P T S P A I S N L V M R R F D E R	900
GCATAGGGGAATGGTGTCAGGCTCGGGGAATTACCTACACCCGCTACTGCGATGACATGA I G E W C Q A R G I T Y T R \fbox{Y} C D D M T	960
CCTTTTCAGGTCACTTCAATGCCCGCCAGGTTAAAAATAAAGTGTGCGGATTGTTAGCGG F S G H F N A R Q V K N K V C G L L A E	1020
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1080
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1140
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1200
GTGAACTTGATCCTTCTGGCGATCTCCACGCACAGGCAACGGCGTATCTTTATGCTTTGC E L D P S G ·D L H A Q A T A Y L Y A L Q	1260
AGGGAAGAATAAACTGGTTATTGCAAAATCAACCCTGAGGATGAGGCCTTTCAACAGGCGA	1320
G R I N W L L Q I N P E D E A F Q Q A R	
GAGAGAGTGTAAAGCGAATGCTGGTTGCATGGTAAGAAAAGCGTCAGGCAĞACGTTTČTG E S V K R M L V A W *	1380
CCTGACCGTTTAGGGGAGAattactgcaactgcgcggcaattagcggccagcgggcgtca	1440
${\tt aaatcatccgtcgggcggtatttaaactcgctgcggacaaaacgtgacagcataccttca}$	1500
cagaaggccaggatctggcttgccagcagggtttcatcgg Oligo 2336	1540



RHIZOBIAL ISOLATES

Strain (legume host genus)	USDA strain no.	Geographic source (date)	msDNA produced
Rhizobium sp. (Acacia)	3002	Brazil (1959)	+
,	3003	Africa (1950)	
	3325	Morocco (1974)	•
	3838	? (1976)	+
Bradyrhizobium sp. (Aeschynomene)	3516	Florida (1972)	+
bradymicoomin sp. (rescriptioniene)	4362	1 Mida (1972)	т
D 1 15-15 -		3111 (2050)	
Bradyrhizobium sp. (Albizia)	3004	Maryland (1952)	+
Bradyrhizobium sp. (Apios)	3240	Maryland (1939)	
Bradyrhizobium sp. (Arachis)	3339	Thailand (1979)	
	3341	Hawaii (1978)	
Rhizobium sp. (Astragalus)	3854	Alaska (1962)	
Rhizobium sp. (Cajanus)	3472		
Bradyrhizobium sp. (Canavalia)	3317	Brazil (1974)	
Rhizobium sp. (Cicer)	3378 '		
our (order)	3379	Mexico (1963)	
Produkischium on (Consuille)			
Bradyrhizobium sp. (Coronilla)	3165	Virginia (1935)	
n . 1 11 . 11	3167	? (1961)	
Bradyrhizobium sp. (Crotalria)	3384	Brazil (1967)	
Bradyrhizobium sp. (Desmodium)	- 3225	Ecuador (1948)	
Bradyrhizobium sp. (Erythrina)	3241		
\$	3242	Maryland (1939)	+
Rhizobium fredii	191	China (1979)	·
		Illinois (1933)	
Rhizobium leguminosarum	2370		
	2429	Hawaii (1978)	
	2435	Holland (1955)	
	2480	Tennessee (1951)	
	2489	• •	
Rhizobium sp. (Lens)	2426		
	3404	Colombia (1979)	
Rhizobium loti	3084	Maryland (1946)	
Ouzooum tott			
	3468	New Zealand (1961)	+
	3469		
	3471		
	3503		+
	3669	California (1968)	
Bradyrhizobium sp. (Lotus)	3074	Minnesota (1954)	
oracymicociam sp. (Doras)	3470	California (1916)	
Dhimakium am (I umiuma)			
Rhizobium sp. (Lupinas)	3040	Florida (1940)	
Bradyrhizobium sp. (Lupinas)	3045	Florida (1946)	
Bradyrhizobium sp. (Macrotyloma)	3451	Zimbabwe (1960)	
Rhizobium medicago	1097	North Dakota (1948)	
Rhizobium meliloti	1011	Maryland (1933)	
	1021a	North Dakota (1948)	
Phizobium phanali		Washington (1948)	
Rhizobium phaseoli	2667	washington (1940)	
	2669	m 11 (0)	
	2674	Brazil (?)	
	2676	Colombia (1972)	
	3256	Illinois (1941)	
Rhizobium sp. (Robinia)	3436	` ,	
gradyrhizobium sp. (Stylosanthes)	3441	Brazil (?)	
augmizoolum sp. (Stylosumines)		Colombia (1976)	
27-2-1: . :0 !!!	3477		
Rhizobium trifolii	2046	Virgi nia (1934)	
	2048	Illinois (1934)	+
	2063	Florida (1939)	
	2065	Alabama (1952)	+
	2116	South Carolina (1944)	
	2134	? (1974)	
		: (17/ 7)	
	2145	0-1161- (2000)	
	2156	California (1920)	
25.5mm に 11.1 12.1	1177	Florida (1939)	
Rhizobium tropici	2744	Brazil (?)	
Rhizobium sp. (Trigonella) Rhizobium tropici Bradyrhizobium sp. (Vigna)	2744 3447	Brazil (?) Thailand (1979)	+

All strains are from the USDA Beltsville Rhizobium Culture Collection, provided by Peter van Berkum.
As defined by detection of radiolabeled msDNA by the RT extension method.

SEQUENCE LISTING

(1) GENERAL\ INFORMATION:

(i) APPI ICANT: Inouye, Sumiko Hsu, Mei-Yin

Eagle, Susan

Inouye, Masayori

- (ii) TITLE OF INVENTION: Prokaryotic Reverse Transcriptase
- (iii) NUMBER OF SEQUENCES: 45
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Weiser & Associates
 - (B) STREET: 230 South Fifteenth Street, Suite 500
 - (C) CITY: \Philadelphia
 - (D) STATE:\Pennsylvania
 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 19102
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: \IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: RatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/269,118
 - (B) FILING DATE: 30-JUN-1994
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Weiser, Gerard J.
 - (B) REGISTRATION NUMBER: 19,763
 - (C) REFERENCE/DOCKET NUMBER: 377.5888P
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 215-879-8383
 - (B) TELEFAX: 215-875-8 94
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2176 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS

(k) LOCATION: 640..2094

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

		(X1)	SEQ	OFINC	E DE	SCRI	PTIO	M: D	EQ 1	UM CL.	1:1:						
r	TCAT	CCGC	GC G	GACA	.CCCC	C TC	CTAC	GTGC	ccc	CCGA	.CGC	GGAG	AGCG	GC (GTGGA	.GACG	G- 60
ı	TGTA	.CCGC	GT T	TCCC	GGA	T GG	TCAC	CTGG	TGG	CGGT	'GGA	GTGG	GGCC	CCG (CGCAC	:GGGC	T 120
	CGCC	GCGT	CA C	CAGC	:GGCI	C TG	GTTC	GACT	' CGG	SATGO	GGA	AGCC	CCCC	GA (GCCTA	CTTC	G 180
i	CGCG	CCTC	GA G	AAGT	TGG	G GC	TGAC	CGGCI	' ACA	ATCGA	CGC	GGCC	TCG0	CA !	TTGGT	'CTAA	A 240
	CCCT	TCAA	CC A	.CGGC	TCGG	id ce	GCCAC	CGCGC	: GGC	CCGGC	AGG	ACAG	GTGC	CGA (CGAAC	AGAC	G 300
-	ACGA	.CGTG	CG C	TTCA	rcece	C GA	AGCAG	CCGA	GAG	GAGGT	CCG	GAGT	'GCA'I	CA (GCCTG	AGCG	C 360
	CTCG	AGCG	IGC G	GAGC	GGCG	T T	cecc	CGCTC	CGG	FTTGG	TAA	GCAG	GACI	ACT (CTCCG	CAAG	G 420
	TAGC	CTGT	TC I	TGGC	CTCTC	T CC	COTCO	CTAGG	CAC	CTAC	GCC	AGGG	TGGG	GTA (GCGGA	GCCA	A 480
The state	CGAC	GCCA	rac e	CCGT	TTAC	CC CF	rcqcc	CGGCC	GT#	AGTG(CTA	GGAG	iggg <i>i</i>	AGA (GCCGG	TGAG	G 540
Post Street	CTAC	CGTG	CC C	CAGG	TAAC	A TO	GTG	TGCT	TTC	CCCGG	GCCT	CCGT	CGAC	CTG	CTCGC	CGCCA	т 600
The street of	GTCC	CGTC	TT	CCATO	CGCCC	GC GC	CCCGC	CAZ	A GGT	rgcac	GAC A	ATG A Met T	CC (GCC :	AGG (Arg I	CTG .eu	654
*											-	1			J	5	
	GAC Asp	CCG Pro	TTC Phe	GTC Val	CCC Pro	GCA Ala	GCT Ala	TCG Ser	CCG Pro	CAG Gln 15	GCC Ala	GTG Val	CCC Pro	ACG Thr	CCC Pro 20	GAG Glu	702
And And Ann Ann	CTC Leu	ACC Thr	GCT Ala	CCG Pro 25	TCG Ser	TCA Ser	GAC Asp	GCG Ala	GCC Ala 30	GCG Ala	AAG Lys	CGT Arg	GAA Glu	GCC Ala 35	CGC Arg	CGG Arg	750
	CTC Leu	GCG Ala	CAC His 40	GAA Glu	GCG Ala	TTG Leu	CTC Leu	GTC Val 45	CGC Arg	GCG Ala	AAG Lys \	GCC Ala	ATC Ile 50	GAC Asp	GAA Glu	GCG Ala	798
	GGC Gly	GGC Gly 55	GCC Ala	GAC Asp	GAC Asp	TGG Trp	GTG Val 60	CAG Gln	GCG Ala	CAG Gln	CTC Leu	GTC Val 65	TCC Ser	AAG Lys	GGG Gly	CTC Leu	846
	GCG Ala 70	GTC Val	GAG Glu	GAC Asp	CTG Leu	GAC Asp 75	TTC Phe	TCC Ser	AGC Ser	GCC Ala	TCC Ser 80	GAG Glu	AAG Lys	GAC Asp	AAG Lys	AAG Lys 85	894
y t	GCC Ala	TGG Trp	AAG Lys	GAG Glu	AAG Lys 90	AAG Lys	AAG Lys	GCC Ala	GAG Glu	GCC Ala 95	ACC Thr	GAG Glu	CGC Arg	CGC Arg	GCG Ala 100	CTG Leu	942
	AAG	CGT	CAG	GCG	CAC	GAG	GCG	TGG	AAG	GCC	ACG	CAC	dTG	GGC	CAC	CTG	990

*	Lys	Arg	Gln	Ala 105	His	Glu	Ala	Trp	Lys 110	Ala	Thr	His	Val	Gly 115	His	Leu	
									GAC Asp								1038
									CGG Arg								1086
-									GCG Ala								1134
1									GAG Glu								1182
									GAC Asp 190								1230
									GCG Ala								1278
	GTC Val																1326
									CTG Leu								1374
And the state of t									TTC Phe								1422
-									GGC Gly 270								1470
									GAA Glu								1518
*									GCC Ala								1566
χ.									ATC Ile								1614

		_															
					CTG Leu 330												1662
	ACG Thr	CGC Arg	TAC Tyr	GCG Ala 345	GAC Asp	GAC Asp	CTG Leu	ACC Thr	TTC Phe 350	TCC Ser	TGG Trp	ACG Thr	AAG Lys	GCG Ala 355	AAG Lys	CAG Gln	1710
	CCC Pro	AAG Lys	CCG Pro 360	CGG Arg	CGG	ACG Thr	CAG Gln	CGT Arg 365	CCC Pro	CCC Pro	GTC Val	GCG Ala	GTC Val 370	CTC Leu	CTG Leu	TCT Ser	1758
	CGC Arg	GTC Val 375	CAG Gln	GAA Glu	GTG Val	GTG Val	GAG Glu 380	GCG Ala	GAG Glu	GGC Gly	TTC Phe	CGC Arg 385	GTG Val	CAC His	CCG Pro	GAC Asp	1806
	AAG Lys 390	ACG Thr	CGC Arg	GTC Val	GCC Ala	CGC Arg 395	AAG Lys	GGC Gly	ACG Thr	CGG Arg	CAG Gln 400	CGG Arg	GTC Val	ACC Thr	GGG Gly	CTC Leu 405	1854
onis then done	GTC Val	GTG Val	AAT Asn	GCG Ala	GCG Ala 410	GGC Gly	AAC Lys	GAC Asp	GCG Ala	CCC Pro 415	GCG Ala	GCC Ala	CGA Arg	GTC Val	CCG Pro 420	CGC Arg	1902
the fields object sense, to	GAC Asp	GTC Val	GTC Val	CGC Arg 425	CAG Gln	CTC Leu	CGC Arg	GCC Ala	GCC Ala \430	ATC Ile	CAC His	AAC Asn	CGG Arg	AAG Lys 435	AAG Lys	GGC Gly	1950
the state of the state of	AAG Lys	CCG Pro	GGC Gly 440	CGC Arg	GAG Glu	GGC Gly	GAG Glu	TCG Ser 445	CTC	GAG Glu	CAG Gln	CTC Leu	AAG Lys 450	GGC Gly	ATG Met	GCC Ala	1998
their their their their	GCC Ala	TTC Phe 455	ATC Ile	CAC His	ATG Met	ACG Thr	GAC Asp 460	CCG Pro	GCC Ala	AAG Lys	GGC Gly	CGC Arg 465	GCC Ala	TTC Phe	CTG Leu	GCT Ala	2046
TTTTTT.	CAG Gln 470	CTC Leu	ACG Thr	GAG Glu	CTC Leu	GAG Glu 475	TCC Ser	ACG Thr	GCG Ala	AGC Ser	ALA 480	GCT Ala	CCG Pro	CAG Gln	GCG Ala	GAG Glu 485	2094
	TGA	CGCT	CAG (CGCG	CGTC	CG T	CGCC	GACG'	T GC	CGCG	CGCC	AGC:	AACG	CCG (CATT	CAGCAA	2154
	CTC	CGTC	AGC (CGGC	GCGG(GT A	3										2176
												`					

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 263 amino acids

- (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi)\SEQUENCE DESCRIPTION: SEQ ID NO:2:

260

Val Lys Leu Lys Pro Gly Met Asp Gly Pro Lys Val Lys Gln Trp Pro Leu Thr Glu Glu Lys Ile Lys Ala Leu Val Glu Ile Cys Thr Glu Met Glu Lys Glu Gly Lys Ile Ser Lys Ile Gly Pro Glu Asn Pro Tyr Asn Thr Pro Val Phe 🔃 la Ile Lys Lys Lys Asp Ser Thr Lys Trp Arg Lys Leu Val Asp Phe Arg Glu Leu Asn Lys Arg Thr Gln Asp Phe Trp Glu Val Gln Leu Gly Ile Pho His Pro Ala Gly Leu Lys Lys Lys Ser Val Thr Val Leu Asp Val Apy Asp Ala Tyr Phe Ser Val Pro Leu Asp 110 105 100 Glu Asp Phe Arg Lys Tyr Thr Ala Phe Thr Ile Pro Ser Ile Asn Asn Glu Thr Pro Gly Ile Arg Tyr Gln\Tyr Asn Val Leu Pro Gln Gly Trp 135 Lys Gly Ser Pro Ala Ile Phe Gln Se χ Ser Met Thr Lys Ile Leu Glu 160 145 150 155 Pro Phe Lys Lys Gln Asn Pro Asp Ile Val Ile Tyr Gln Tyr Met Asp Asp Leu Tyr Val Gly Ser Asp Leu Glu Ile Gly Gln His Arg Thr Lys 185 Ile Glu Glu Leu Arg Gln His Leu Leu Arg Trp\Gly Leu Thr Thr Pro 195 Asp Lys Lys His Gln Lys Glu Pro Pro Phe Leu Tro Met Gly Tyr Glu 220 210 215 Leu His Pro Asp Lys Trp Thr Val Gln Pro Ile Val Læu Pro Glu Lys 235 Asp Ser Trp Thr Val Asn Asp Ile Gln Lys Leu Val Gly Lys Leu Asn 245 255 Trp Ala Ser Gln Ile Tyr Pro

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 263 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
- Arg Pro Trp Ala Arg Thr Pro Pro Lys Ala Pro Arg Asn Gln Pro Val
- Pro Phe Lys Pro Glu Arg Leu Gln Ala Leu Gln His Leu Val Arg Lys
- Ala Leu Glu Ala Gly His The Glu Pro Tyr Thr Gly Pro Gly Asn Asn 35
- Pro Val Phe Pro Val Lys Lys Ala Asn Gly Thr Trp Arg Phe Ile His 50 55
- Asp Leu Arg Ala Thr Asn Ser Leu Thr Ile Asp Leu Ser Ser Ser Ser 65 70 75 80
- Pro Gly Pro Pro Asp Leu Ser Ser Leu Pro Thr Thr Leu Ala His Leu 85 90 95
- Gln Thr Ile Asp Leu Arg Asp Ala Phe Phe Gln Ile Pro Leu Pro Lys
- Gln Phe Gln Pro Tyr Phe Ala Phe Thr Val Pro Gln Gln Cys Asn Tyr 115 120 125
- Gly Pro Gly Thr Arg Tyr Ala Trp Lys Val\Leu Pro Gln Gly Phe Lys
 130 135
- Asn Ser Pro Thr Leu Phe Glu Met Gln Leu Ala His Ile Leu Gln Pro 145 150 160
- Ile Arg Gln Ala Phe Pro Gln Cys Thr Ile Leu Öln Tyr Met Asp Asp
 165 170 175
- Ile Leu Leu Ala Ser Pro Ser His Glu Asp Leu Leu Leu Ser Glu
 180 185 190
- Ala Thr Met Ala Ser Leu Ile Ser His Gly Leu Pro Val Ser Glu Asn
 195 200 205
- Lys Thr Gln Gln Thr Pro Gly Thr Ile Lys Phe Leu Gly Gln Ile Ile 210 215 220

Ser Pro Asn His Leu Thr Tyr Asp Ala Val Pro Thr Val Pro Ile Arg 225 230 235 240

Ser Arg Trp Ala Leu Pro Glu Leu Gln Ala Leu Leu Gly Glu Ile Gln 245 250 255

Trp Val Ser Lys Gly Thr Pro

- (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH:\259 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Asn Val Leu Tyr Arg Ile Gly Ser Asp Asn Gln Tyr Thr Gln Phe Thr

Ile Pro Lys Lys Gly Lys Gly Wal Arg Thr Ile Ser Ala Pro Thr Asp

Arg Leu Lys Asp Ile Gln Arg Arg \text{\text{Vle Cys Asp Leu Leu Ser Asp Cys}}
35 40 45

Arg Asp Glu Ile Phe Ala Ile Arg Lys Ile Ser Asn Asn Tyr Ser Phe 50 55

Gly Phe Glu Arg Gly Lys Ser Ile Ile Leu Asn Ala Tyr Lys His Arg
65 70 80

Gly Lys Gln Ile Ile Leu Asn Ile Asp Leu Lys Asp Phe Phe Glu Ser 85 90 95

Phe Asn Phe Gly Arg Val Arg Gly Tyr Phe Leu\Ser Asn Gln Asp Phe
100 105 110

Leu Leu Asn Pro Val Val Ala Thr Thr Leu Ala Lys Ala Ala Cys Tyr

Asn Gly Thr Leu Pro Gln Gly Ser Pro Cys Ser Pro 1 le Ile Ser Asn 130 135 140

Leu Ile Cys Asn Ile Met Asp Met Arg Leu Ala Lys Leu Ala Lys Lys 145 150 155 160

Tyr Gly Cys Thr Tyr Ser Arg Tyr Ala Asp Asp Ile Thr Ile Ser Thr 165 170 175 Asn Lys Asn Thr Phe Pro Leu Glu Met Ala Thr Val Gln Pro Glu Gly 180 185 190

Val Val Leu Gly Lys Val Leu Val Lys Glu Ile Glu Asn Ser Gly Phe 195 200 205

Glu Ile Asn Asp Ser Lys Thr Arg Leu Thr Tyr Lys Thr Ser Arg Gln 210 215 220

Glu Val Thr Cly Leu Thr Val Asn Arg Ile Val Asn Ile Asp Arg Cys 235 240

Tyr Tyr Lys Lys Thr Arg Ala Leu Ala His Ala Leu Tyr Arg Thr Gly 250 255

Glu Tyr Lys

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 266 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Ala Phe His Arg Glu Val Asp Thr Ala Thr His Tyr Val Ser Trp Thr 1 5

Ile Pro Lys Arg Asp Gly Ser Lys Arg Thr Ile Thr Ser Pro Lys Pro 20 25 30

Glu Leu Lys Ala Ala Gln Arg Trp Val Leu Ser Asn Val Val Glu Arg
35 40 45

Leu Pro Val His Gly Ala Ala His Gly Phe Val Ala Gly Arg Ser Ile
50 55 60

Leu Thr Asn Ala Leu Ala His Gln Gly Ala Asp Val Val Lys Val 65 70 75 80

Asp Leu Lys Asp Phe Phe Pro Ser Val Thr Trp Arg Arg Val Lys Gly 85 90

Leu Leu Arg Lys Gly Gly Leu Arg Glu Gly Thr Ser Thr Leu Leu Ser

Leu Leu Ser Thr Glu Ala Pro Arg Glu Ala Val Gln Phe Arg Gl χ Lys 115 120 125

Leu Leu His Val Ala Lys Gly Pro Arg Ala Leu Pro Gln Gly Ala Pro 130 135 140

Thr Ser Pro Gly Ile Thr Asn Ala Leu Cys Leu Lys Leu Asp Lys Arg 145 150 155 160

Leu Ser Ala Neu Ala Lys Arg Leu Gly Phe Thr Tyr Thr Arg Tyr Ala 165 170 175

Asp Asp Leu Thr Phe Ser Trp Thr Lys Ala Lys Gln Pro Lys Pro Arg
180 185 190

Arg Thr Gln Arg Pro Val Ala Val Leu Leu Ser Arg Val Gln Glu
195 200 205

Val Val Glu Ala Glu Oly Phe Arg Val His Pro Asp Lys Thr Arg Val 210 220

Ala Arg Lys Gly Thr Arg Gln Arg Val Thr Gly Leu Val Val Asn Ala 225 230 230 235

Ala Gly Lys Asp Ala Pro Ala Ala Arg Val Pro Arg Asp Val Val Arg 255

Gln Leu Arg Ala Ala Ile His Asn Arg Lys 265

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 111 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Pro Thr Pro Glu Leu Thr Ala Pro Ser Ser Asp Ala Ala Ala Lys Arg
1 10 15

Glu Ala Arg Arg Leu Ala His Glu Ala Leu Leu Val Arg Ala Lys Ala
20 25 30

Ile Asp Glu Ala Gly Gly Ala Asp Asp Trp Val Gln Ala Gln Leu Val

Ser Lys Gly Leu Ala Val Glu Asp Leu Asp Phe Ser Set Ala Ser Glu
50 60

Lys Asp Lys Lys Ala Trp Lys Glu Lys Lys Lys Ala Glu Ala Thr Glu 65 70 75 80

Arg Arg Ala Leu Lys Arg Gln Ala His Glu Ala Trp Lys Ala Thr His 90 95

Val Gly His Leu Gly Ala Gly Val His Trp Ala Glu Asp Arg Leu 100 105 110

- (2) INFORMATION FOR SEQ ID NO:7:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 110 amino acids
 - (B) TYPE:\amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Pro Asp Pro Asp Met The Arg Val The Asn Ser Pro Ser Leu Gln Ala
1 10 15

His Leu Gln Ala Leu Tyr Deu Val Gln His Glu Val Trp Arg Pro Leu 20 25 30

Ala Ala Tyr Gln Glu Gln Leu Asp Arg Pro Val Val Pro His Pro 35

Tyr Arg Val Gly Asp Thr Val Tro Val Arg Arg His Gln Thr Lys Asn 50 60

Leu Glu Pro Arg Trp Lys Gly Pro Tyr Thr Val Leu Leu Thr Thr Pro 65 70 75 80

Thr Ala Leu Lys Val Asp Gly Ile Ala Ala Trp Ile His Ala Ala His 85 90 95

Val Lys Ala Ala Asp Pro Gly Gly Gly Pro Ser Ser Arg Leu
100 105

- (2) INFORMATION FOR SEQ ID NO:8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 75 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
 - Gly Lys Asp Ala Pro Ala Ala Arg Val Pro Arg Asp Val Val Arg Gln

Leu Arg Ala Ala Ile His Asn Arg Lys Lys Gly Lys Pro Gly Arg Glu 25 30

Gly Glu Ser Leu Glu Gln Leu Lys Gly Met Ala Ala Phe Ile His Met

Thr Asp Pro Ala Lys Gly Arg Ala Phe Leu Ala Gln Leu Thr Glu Leu 50 60

Glu Ser Thr Ala Ser Ala Ala Pro Gln Ala Glu 65 70 75

- (2) INFORMATION FOR SEQ ID NO:9:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 66 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY \ linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Gly Lys Glu Gly His Ser Ala Arg Gln Cys Arg Ala Pro Arg Gln
1 10 15

Gly Cys Trp Lys Cys Gly Lys Pro Gly His Ile Met Thr Asn Cys Pro 25

Asp Arg Gln Ala Gly Phe Leu Gly Leu Gly Pro Trp Gly Lys Lys Pro 35 40 45

Arg Asn Phe Pro Val Ala Gln Val Pro Gln Gly Leu Thr Pro Thr Ala 50 55 60

Pro Pro 65

- (2) INFORMATION FOR SEQ ID NO:10:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 68 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Gly Pro Arg Ala Leu Pro Gln Gly Ala Pro Thr Ser Pro Gly Ile Thr 1 5 10 15

Asn Ala Leu Cys Leu Lys Leu Asp Lys Arg Leu Ser Ala Leu Ala Lys 20 25 30

Arg Leu Gly Phe Thr Tyr Thr Arg Tyr Ala Asp Asp Leu Thr Phe Ser 35 40 45

Trp Thr Lys Ala Lys Gln Pro Lys Pro Arg Arg Thr Gln Arg Pro Pro 50 55 60

Val Ala Val Leu 65

- (2) INFORMATION FOR SEQ \ID NO:11:
 - (i) SEQUENCE CHARACTARISTICS:
 - (A) LENGTH: 68 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Tyr Asn Gly Thr Leu Pro Gln Gly Ser Pro Cys Ser Pro Ile Ile Ser 10 15

Asn Leu Ile Cys Asn Ile Met Asp\Met Arg Leu Ala Lys Leu Ala Lys 20 30

Lys Tyr Gly Cys Thr Tyr Ser Arg Tyr Ala Asp Asp Ile Thr Ile Ser 35 40 45

Thr Asn Lys Asn Thr Phe Pro Leu Glu Met Ala Thr Val Gln Pro Glu 50 55 60

Gly Val Val Leu 65

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 68 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Tyr Lys Asn Leu Leu Pro Gln Gly Ala Pro Ser Ser Pro Lys Leu Ala 1 5 10 15

Asn Leu tle Cys Ser Lys Leu Asp Tyr Arg Ile Gln Gly Tyr Ala Gly 25 30

Ser Arg Gly Leu Ile Tyr Thr Arg Tyr Ala Asp Asp Leu Thr Leu Ser

Ala Gln Ser Met Lys Lys Val Val Lys Ala Arg Asp Phe Leu Phe Ser 50 55 60

Ile Ile Pro Ser

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 67\amino acids
 - (B) TYPE: amind acid
 - (D) TOPOLOGY: 14near
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION:\SEQ ID NO:13:

Tyr Gln Tyr Asn Val Leu Pro Gln Gly Trp Lys Gly Ser Pro Ala Ile 1 5 10 15

Phe Gln Ser Ser Met Thr Lys I Leu Glu Pro Phe Lys Lys Gln Asn 20 25 30

Pro Asp Ile Val Ile Tyr Gln Tyr Met Asp Asp Leu Tyr Val Gly Ser

Asp Leu Glu Ile Gly Gln His Arg Thr Lys Ile Glu Glu Leu Arg Gln 50 60

His Leu Leu 65

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 66 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Tyr Ala Trp Lys Val Leu Pro Gln Gly Phe Lys Asn Ser Pro Thr Leu 1 5 10 15

Phe Glu Met Gln Leu Ala His Ile Leu Gln Pro Ile Arg Gln Ala Phe 20 25 30

Pro Gln Cys Thr Ile Leu Gln Tyr Met Asp Asp Ile Leu Leu Ala Ser 35 40 45

Pro Ser His Glu Asp Leu Leu Leu Ser Glu Ala Thr Met Ala Ser 50 55 60

Leu Ile 65

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 65 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Leu Thr Trp Thr Arg Leu Pro Gln Gly Phe Lys Asn Ser Pro Thr Leu 1 5 10 15

Phe Asp Glu Ala Leu His Arg Asp Leu Ala Asp Phe Arg Ile Gln His 20 25 30

Pro Asp Leu Ile Leu Leu Gln Tyr Val Asp Asp Leu Leu Leu Ala Ala 35 40 45

Thr Ser Glu Leu Asp Cys Gln Gln Gly Thr Arg Ala Leu Leu Gln Thr 50 55 60

Leu 65

- (2) INFORMATION FOR SEQ ID NO:16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 65 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Phe Gln Trp Lys Val Leu Pro Gln Gly Met Thr Cys Ser Pro Thr Ile 1 5 10 15

Cys Gln Leu Val Val Gly Gln Val Leu Glu Pro Leu Arg Leu Lys His 20 25 30

Pro Ser Leu Cys Met Leu His Tyr Met Asp Asp Leu Leu Leu Ala Ala 35 40 45

Ser Ser His Asp Gly Leu Glu Ala Ala Gly Glu Glu Val Ile Ser Thr 50 55 60

Leu 65

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 65 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Phe Ala Trp Arg Val Leu Pro Gln Gly Phe Ile Asn Ser Pro Ala Leu 1 5 10 15

Phe Glu Arg Ala Leu Gln Glu Pro Leu Arg Gln Val Ser Ala Ala Phe 20 25 30

Ser Gln Ser Leu Leu Val Ser Tyr Met Asp Asp Ile Leu Tyr Ala Ser 35 40 45

Pro Thr Glu Glu Gln Arg Ser Gln Cys Tyr Gln Ala Leu Ala Ala Arg 50 55 60

Leu 65

- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 61 amino acids
 - (B) TYPE: amino acid
 - (D) TOFOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Ile Ala Thr Asn Gly Val Pro Gln Gly Ala Ser Thr Ser Cys Gly Leu 1 5 10 15

Ala Thr Tyr Asn Val Leu Glu Leu Phe Leu Arg Tyr Asp Glu Leu Ile 20 25 30

Met Tyr Ala Asp Asp Gly Ile Leu Cys Arg Gln Asp Pro Ser Thr Pro 35 40 45

Asp Phe Ser Val Glu Glu Ala Gly Val Val Gln Glu Pro 50 55 60

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 61 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Tyr Glu Tyr Leu Arg Met Pro Phe Gly Leu Lys Asn Ala Pro Ala Thr 1 5 10 15

Phe Gln Arg Cys Met Asn Asp Ile Leu Arg Pro Leu Leu Asn Lys His
20 25 30

Cys Leu Val Tyr Leu Asp Asp Ile Ile Val Phe Ser Thr Ser Leu Asp 35 40 45

Glu His Leu Gln Ser Leu Gly Leu Val Phe Glu Lys Leu 50 55 60

- (2) INFORMATION FOR SEQ ID NO:20:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 61 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Tyr Glu Phe Cys Arg Leu Pro Phe Gly Leu Arg Asn Ala Ser Ser Ile 1 5 10 15

Phe Gln Arg Ala Leu Asp Asp Val Leu Arg Glu Gln Ile Gly Lys Ile 20 25 30

Cys Tyr Val Tyr Val Asp Asp Val Ile Ile Phe Ser Glu Asn Glu Ser 35 40 45

Asp His Val Arg His Ile Asp Thr Val Leu Lys Cys Leu 50 55 60

- (2) INFORMATION FOR SEQ ID NO:21:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 63 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Cys Lys Leu Asn Lys Ala Ile Tyr Gly Leu Lys Gln Ala Ala Arg Cys
1 10 15

Trp Phe Arg Cys Ile Tyr Ile Leu Asp Lys Gly Asn Ile Asn Glu Asn 20 25 30

Ile Tyr Val Leu Leu Tyr Val Asp Asp Val Val Ile Ala Thr Gly Asp 35 40 45

Met Thr Arg Met Asn Asn Phe Lys Arg Tyr Leu Met Glu Lys Phe 50 55 60

- (2) INFORMATION FOR SEQ ID NO:22:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 62 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Cys Leu Leu Lys Lys Ser Leu Tyr Gly Leu Lys Gln Ser Pro Arg Gln 1 5 10 15

Trp Asn Ala Cys Val Tyr Val Lys Gln Val Ser Glu Gln Glu His Leu 20 25 30

Tyr Leu Leu Tyr Val Asp Asp Met Leu Ile Ala Gly Lys Ser Lys 35 40 45

Ser Glu Ile Asn Lys Val Lys Glu Gln Leu Ser Met Glu Phe 50 55 60

- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 63 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Ile Arg Leu Lys Lys Ser Leu Tyr Glu Leu Lys Gln Ser Gly Ala Asn 10 15

Trp Tyr Glu Glu Val Arg Gly Trp Ser Cys Val Phe Lys Asn Ser Gln 20 25 30

Val Thr Ile Cys Leu Phe Val Asp Asp Met Val Leu Phe Ser Lys Asn 35 40 45

Leu Asn Ser Asn Lys Arg Ile Ile Glu Lys Leu Lys Met Gln Tyr 50 55 60

- (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 58 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ix) FEATURE:
 - (A) NAME/KEY: misc feature
 - (B) LOCATION: 15
 - (D) OTHER INFORMATION: /note= "The 2' position of this nucleotide is linked to the 5' position of nucleotide number 1 of SEQ ID NO: 25 of this application."
 - (ix) FEATURE:
 - (A) NAME/KEY: misc binding
 - (B) LOCATION: 52..58

			STRANDEDNESS: single TOPOLOGY: linear
l	(ix)	(B)	RE: NAME/KEY: misc_feature LOCATION: 1 OTHER INFORMATION: /note= "The 5' position of this nucleotide is linked to the 2' position of nucleotide number 15 of SEQ ID NO: 24 of this application."
	(ix)	(B)	NAME/KEY: misc_binding LOCATION: 6167 OTHER INFORMATION: /note= "This region can hydrogen bond to nucleotides 52-58 of SEQ ID NO: 24 of the application."
TCC			NCE DESCRIPTION: SEQ ID NO:25:
TCC	TGCC		
(2)	INFO	RMATIC	ON FOR SEQ ID NO:26:

(D) OTHER INFORMATION: /note= "This region can hydrogen

CACGCAUGUA GGCAGAUUUG UUGGUUGUGA AUCGCAACCA GUGGCCUUAA UGGCAGGA

application."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

(i) SEQUENCE CHARACTERISTICS:

(D) TOPOLOGY: linear

(A) NAME/KEY: CDS

(ix) FEATURE:

(A) LENGTH: 2423 base pairs

(B) TYPE: nucleic acid(C) STRANDEDNESS: double

(B) LOCATION: 418..2175

(A) LENGTH: 67 base pairs (B) TYPE: nucleic acid

bond to nucleotides 61-67 of SEQ ID NO: 25 of this

58

this

60

67

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

		(XI)	SEQ	OBMC	.E DE	IJ CKI	FIIO	14.	11.7 T	D 110							
	TGGC	CATT	'NA G	ATAC	GGAT	TT TI	'CACT	TCCI	' TGA	CAGT	'GCA	TGAC	TATG	CT G	CATG	AAATN	60
	GCAT	GATC	GA T	TGAG	GATC	G TO	TTTG	CTCA	GAT	'CCGC	CAG	AACT	'GGCG	GG C	TTTT	GCTCA	120
	TGTC.	ATGC	AT G	TGCA	TGAA	A AC	CACT	GCAT	' AAA	.GCGG	GCA	GGCG	TGGC	GG G	GATA	.CGAGC	180
	GCGC	GCTA	TC A	CCGA	TAAA	'A GO	CAAA	ATAC	TTC	TGGA	AAA	CAGA	AAGT	TG A	AGTG	ATATG	240
	TTCA	TAAA	CA C	GCAT	GTAG	G CA	GATT	TGTT	GGT	TGTG	TAA	CGCA	ACCA	GT G	GCCT	TAATG	300
	GCAG	GAGG	L AA	CGCC	TCCC	CT AA	AATC	CTTC	aTT	CAGA	AGCT	ATAC	GGCA	.GG I	GTGC	TGTGC	360
	GAAG	GAGI	GC C	CTGCA	TGCG	r T	CTCC	TTGG	CCI	TTTT	TCC	TCTO	GGAT	'GA A	GAAG	AA	417
	ATG Met	ACA Thr	AAA Lys	ACA Thr	TCT Ser 5	AAA Lys	CTT Leu	GAC Asp	GCA Ala	CTT Leu 10	AGG Arg	GCT Ala	GCT Ala	ACT Thr	TCA Ser 15	CGT Arg	465
	GAA Glu	GAC Asp	TTG Leu	GCT Ala 20	AAA Lys	ATT Ile	TTA Leu	GAT Asp	ATT Ile 25	AAG Lys	TTG Leu	GTA Val	TTT Phe	TTA Leu 30	ACT Thr	AAC Asn	513
The state state of	GTT Val	CTA Leu	TAT Tyr 35	AGA Arg	ATC Ile	GGC Gly	TCG Ser	GAT Asp 40	AAT Asn	CAA Gln	TAC Tyr	ACT Thr	CAA Gln 45	TTT Phe	ACA Thr	ATA Ile	561
the standard the standard to	CCG Pro	AAG Lys 50	AAA Lys	GGA Gly	AAA Lys	GGG Gly	GTA Val 55	AGG Arg	ACT Thr	ATT Ile	TCT Ser	GCA Ala 60	CCT Pro	ACA Thr	GAC Asp	CGG Arg	609
Land Same West Co.	TTG Leu 65	AAG Lys	GAC Asp	ATC Ile	CAA Gln	CGA Arg 70	AGA Arg	ATA Ile	TGT Cys	GAC Asp	TTA Leu 75	CTT Leu	TCT Ser	GAT Asp	TGT Cys	AGA Arg 80	657
The state of the s	GAT Asp	GAG Glu	ATC Ile	TTT Phe	GCT Ala 85	ATA Ile	AGG Arg	Lys	Ile	Ser	Asn	AAC Asn	Tyr	Ser	Pne	GGT Gly	705
	TTT Phe	GAG Glu	AGG Arg	GGA Gly 100	AAA Lys	TCA Ser	ATA Ile	ATC Ile	CTA Leu 105	AAT Asn	GCT Ala	TAT Tyr	AAG Lys	CAT His 110	AGA Arg	GGC Gly	753
	AAA Lys	CAA Gln	ATA Ile 115	ATA Ile	TTA Leu	AAT Asn	ATA Ile	GAT Asp 120	CTT Leu	AAG Lys	GAT Asp	TTT Phe	TTT Phe 125	GAA Glu	AGC Ser	TTT Phe	801
	AAT Asn	TTT Phe 130	Gly	CGA Arg	GTT Val	AGA Arg	GGA Gly 135	TAT Tyr	TTT Phe	CTT Leu	TCC Ser	AAT Asn 140	CAG Gln	GAT Asp	TTT Phe	TTA Leu	849
	TTA Leu	AAT Asn	CCT Pro	GTG Val	GTG Val	GCA Ala	ACG Thr	ACA Thr	CTT Leu	GCA Ala	AAA Lys	GCT Ala	GCA Ala	TGC Cys	TAT Tyr	AAT Asn	897

	145					150					155					160		
	GGA Gly	ACC Thr	CTC Leu	CCC Pro	CAA Gln 165	GGA Gly	AGT Ser	CCA Pro	TGT Cys	TCT Ser 170	CCT Pro	ATT Ile	ATC Ile	TCA Ser	AAT Asn 175	CTA Leu	945	
	ATT Ile	TGC Cys	AAT Asn	ATT Ile 180	ATG Met	GAT Asp	ATG Met	AGA Arg	TTA Leu 185	GCT Ala	AAG Lys	CTG Leu	GCT Ala	AAA Lys 190	AAA Lys	TAT Tyr	993	
	GGA Gly	TGT Cys	ACT Thr 195	TAT Tyr	AGC Ser	AGA Arg	TAT Tyr	GCT Ala 200	GAT Asp	GAT Asp	ATA Ile	ACA Thr	ATT Ile 205	TCT Ser	ACA Thr	AAT Asn	1041	
	AAA Lys	AAT Asn 210	ACA Thr	TTT Phe	CCG Pro	TTA Leu	GAA Glu 215	ATG Met	GCT Ala	ACT Thr	GTG Val	CAA Gln 220	CCT Pro	GAA Glu	GGG Gly	GTT Val	1089	
, {	Val 225	TTG Leu	GGA Gly	AAA Lys	GTT Val	TTG Leu 230	GTA Val	AAA Lys	GAA Glu	ATA Ile	GAA Glu 235	AAC Asn	TCT Ser	GGA Gly	TTC Phe	GAA Glu 240	1137	
	M ATA M Ile	AAT Asn	GAT Asp	TCA Ser	AAG Lys 245	ACT Thr	AGG Arg	CTT Leu	ACG Thr	TAT Tyr 250	AAG Lys	ACA Thr	TCA Ser	AGG Arg	CAA Gln 255	GAA Glu	1185	
*	GTA Val	ACG Thr	GGA Gly	CTT Leu 260	ACA Thr	GTT Val	AAC Asn	AGA Arg	ATC Ile 265	GTT Val	AAT Asn	ATT Ile	GAT Asp	AGA Arg 270	TGT Cys	TAT Tyr	1233	
	TAT U Tyr	AAA Lys	AAA Lys 275	ACT Thr	CGG Arg	GCG Ala	TTG Leu	GCA Ala 280	CAT His	GCT Ala	TTG Leu	TAT Tyr	CGT Arg 285	ACA Thr	GGT Gly	GAA Glu	1281	
(U TAT	AAA Lys 290	GTG Val	CCA Pro	GAT Asp	GAA Glu	AAT Asn 295	GGT Gly	GTT Val	TTA Leu	GTT Val	TCA Ser 300	GGA Gly	GGT Gly	CTG Leu	GAT Asp	1329	
	AAA Lys 305	CTT Leu	GAG Glu	GGG Gly	ATG Met	TTT Phe 310	GGT Gly	TTT Phe	ATT Ile	GAT Asp	CAA Gln 315	GTT Val	GAT Asp	AAG Lys	TTT Phe	AAC Asn 320	1377	
	AAT Asn	ATA Ile	AAG Lys	AAA Lys	AAA Lys 325	CTG Leu	AAC Asn	AAG Lys	CAA Gln	CCT Pro 330	GAT Asp	aga Arg	TAT Tyr	GTA Val	TTG Leu 335	ACT Thr	1425	
	AAT Asn	GCG Ala	ACT Thr	TTG Leu 340	CAT	GGT Gly	TTT Phe	AAA Lys	TTA Leu 345	Lys	TTG Leu	AAT Asn	GCG Ala	CGA Arg 350	GAA Glu	AAA Lys	1473	
·	GCA Ala	TAT Tyr	AGT Ser 355	Lys	TTT Phe	ATT Ile	TAC Tyr	TAT Tyr 360	Lys	TTT Phe	TTT Phe	CAT His	GGC Gly 365	Asn	ACC Thr	TGT Cys	1521	

												ATA Ile 380					1569
												TTG Leu					1617
												ATA Ile					1665
												GGA Gly					1713
al												GCT Ala					1761
												GAT Asp 460					1809
	_											GTT Val					1857
Willer H Charles many	GAC											ATT Ile					1905
Sales Sales	AAT Asn	TTA Leu	TAT Tyr	ATA Ile 500	GTT Val	CTC Leu	ACA Thr	CCA Pro	TTG Leu 505	AGT Ser	CCT Pro	TCC Ser	GGC Gly	GAA Glu 510	CAA Gln	ACT Thr	1953
Pro de												GAT Asp					2001
												TCA Ser 540					2049
												GAT Asp					2097
												GCT Ala					2145
						TTA Leu					TAAT	rgaa(CAG (CCT	AACGT	T	2195

ATGAACGCTA	AGGCTGATTT	TTCGTTAAAA	TTTATATGGT	TTGAATTGTA	ATATATTATC	2255
TTCAAGCCAT	TTATTTAATT	CCTGCATCCT	TTTCTGTAAG	GGTATTAATT	CGTTCCTCAC	2315
AAACACTAAA	CTCGCTTTTT	CCACATCCCC	AAACCCCCCT	AACATTATTC	GGCATAATCC	2375
CCATCATTTG	CGGTGGCACA	CGATGCGCTG	CCATCATGTC	ATCGCGGC		2423

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 546 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Val Lys Leu Lys Pro Gly Met Asp Gly Pro Lys Val Lys Gln Trp Pro 1 5 10 15

Leu Thr Glu Glu Lys Ile Lys Ala Leu Val Glu Ile Cys Thr Glu Met 20 25 30

Glu Lys Glu Gly Lys Ile Ser Lys Ile Gly Pro Glu Asn Pro Tyr Asn 35 40 45

Thr Pro Val Phe Ala Ile Lys Lys Lys Asp Ser Thr Lys Trp Arg Lys 50 55 60

Leu Val Asp Phe Arg Glu Leu Asn Lys Arg Thr Gln Asp Phe Trp Glu 65 70 75 80

Val Gln Leu Gly Ile Pro His Pro Ala Gly Leu Lys Lys Lys Ser 85 90 95

Val Thr Val Leu Asp Val Gly Asp Ala Tyr Phe Ser Val Pro Leu Asp
100 105 110

Glu Asp Phe Arg Lys Tyr Thr Ala Phe Thr Ile Pro Ser Ile Asn Asn 115 120 125

Glu Thr Pro Gly Ile Arg Tyr Gln Tyr Asn Val Leu Pro Gln Gly Trp 130 135 140

Lys Gly Ser Pro Ala Ile Phe Gln Ser Ser Met Thr Lys Ile Leu Glu 145 150 155 160 Pro Phe Lys Lys Gln Asn Pro Asp Ile Val Ile Tyr Gln Tyr Met Asp Asp Leu Tyr Val Gly Ser Asp Leu Glu Ile Gly Gln His Arg Thr Lys Ile Glu Glu Leu Arg Gln His Leu Leu Arg Trp Gly Leu Thr Thr Pro Asp Lys Lys His Gln Lys Glu Pro Pro Phe Leu Trp Met Gly Tyr Glu Leu His Pro Asp Lys Trp Thr Val Gln Pro Ile Val Leu Pro Glu Lys Asp Ser Trp Thr Val Asn Asp Ile Gln Lys Leu Val Gly Lys Leu Asn Trp Ala Ser Gln Ile Tyr Pro Gly Ile Lys Val Arg Gln Leu Cys Lys Leu Leu Arg Gly Thr Lys Ala Leu Thr Glu Val Ile Pro Leu Thr Glu Glu Ala Glu Leu Glu Leu Ala Glu Asn Arg Glu Ile Leu Lys Glu Pro Val His Gly Val Tyr Tyr Asp Pro Ser Lys Asp Leu Ile Ala Glu Ile Gln Lys Gln Gly Gln Gly Gln Trp Thr Tyr Gln Ile Tyr Gln Glu Pro Phe Lys Asn Leu Lys Thr Gly Lys Tyr Ala Arg Met Arg Gly Ala His Thr Asn Asp Val Lys Gln Leu Thr Glu Ala Val Gln Lys Ile Thr Thr Glu Ser Ile Val Ile Trp Gly Lys Thr Pro Lys Phe Lys Leu Pro Ile Gln Lys Glu Thr Trp Glu Thr Trp Trp Thr Glu Tyr Trp Gln Ala Thr Trp Ile Pro Glu Trp Glu Phe Val Asn Thr Pro Pro Leu Val Lys Leu Trp Tyr Gln Leu Glu Lys Glu Pro Ile Val Gly Ala Glu Thr Phe Tyr Val Asp Gly Ala Ala Asn Arg Glu Thr Lys Leu Gly Lys Ala Gly Tyr

Val Thr Asn Lys Gly Arg Gln Lys Val Val Pro Leu Thr Asn Thr Thr 450 455 460

Asn Gln Lys Thr Glu Leu Gln Ala Ile Tyr Leu Ala Leu Gln Asp Ser 465 470 475 480

Gly Leu Glu Val Asn Ile Val Thr Asp Ser Gln Tyr Ala Leu Gln Ile 485 490 495

Ile Gln Ala Gln Pro Asp Lys Ser Glu Ser Glu Leu Val Asn Gln Ile 500 505 510

Ile Glu Gln Leu Ile Lys Lys Glu Lys Val Tyr Leu Ala Trp Val Pro 515 520 525

Ala His Lys Gly Ile Gly Gly Asn Glu Gln Val Asp Lys Leu Val Ser 530 535

Ala Gly 545

- (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 578 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Arg Pro Trp Ala Arg Thr Pro Pro Lys Ala Pro Arg Asn Gln Pro Val 1 5 10 15

Pro Phe Lys Pro Glu Arg Leu Gln Ala Leu Gln His Leu Val Arg Lys 20 25 30

Ala Leu Glu Ala Gly His Ile Glu Pro Tyr Thr Gly Pro Gly Asn Asn 35 40 45

Pro Val Phe Pro Val Lys Lys Ala Asn Gly Thr Trp Arg Phe Ile His 50 55 60

Asp Leu Arg Ala Thr Asn Ser Leu Thr Ile Asp Leu Ser Ser Ser Ser 65 70 75 80

Pro Gly Pro Pro Asp Leu Ser Ser Leu Pro Thr Thr Leu Ala His Leu 85 90 95

Gln Thr Ile Asp Leu Arg Asp Ala Phe Phe Gln Ile Pro Leu Pro Lys 100 105 110 Gln Phe Gln Pro Tyr Phe Ala Phe Thr Val Pro Gln Gln Cys Asn Tyr Gly Pro Gly Thr Arg Tyr Ala Trp Lys Val Leu Pro Gln Gly Phe Lys Asn Ser Pro Thr Leu Phe Glu Met Gln Leu Ala His Ile Leu Gln Pro Ile Arg Gln Ala Phe Pro Gln Cys Thr Ile Leu Gln Tyr Met Asp Asp Ile Leu Leu Ala Ser Pro Ser His Glu Asp Leu Leu Leu Ser Glu Ala Thr Met Ala Ser Leu Ile Ser His Gly Leu Pro Val Ser Glu Asn Lys Thr Gln Gln Thr Pro Gly Thr Ile Lys Phe Leu Gly Gln Ile Ile Ser Pro Asn His Leu Thr Tyr Asp Ala Val Pro Thr Val Pro Ile Arg Ser Arg Trp Ala Leu Pro Glu Leu Gln Ala Leu Leu Gly Glu Ile Gln Trp Val Ser Lys Gly Thr Pro Thr Leu Arg Gln Pro Leu His Ser Leu Tyr Cys Ala Leu Gln Arg His Thr Asp Pro Arg Asp Gln Ile Tyr Leu Asn Pro Ser Gln Val Gln Ser Leu Val Gln Leu Arg Gln Ala Leu Ser Gln Asn Cys Arg Ser Arg Leu Val Gln Thr Leu Pro Leu Leu Gly Ala Ile Met Leu Thr Leu Thr Gly Thr Thr Thr Val Val Phe Gln Ser Lys Glu Gln Trp Pro Leu Val Trp Leu His Ala Pro Leu Pro His Thr Ser Gln Cys Pro Trp Gly Gln Leu Leu Ala Ser Ala Val Leu Leu Leu Asp Lys Tyr Thr Leu Gln Ser Tyr Gly Leu Leu Cys Gln Thr Ile His His Asn Ile Ser Thr Gln Thr Phe Asn Gln Phe Ile Gln Thr Ser Asp His

. 1 1

Pro Ser Val Pro Ile Leu Leu His His Ser His Arg Phe Lys Asn Leu 405 410 415

Gly Ala Gln Thr Gly Glu Leu Trp Asn Thr Phe Leu Lys Thr Ala Ala 420 425 430

Pro Leu Ala Pro Val Lys Ala Leu Met Pro Val Phe Thr Leu Ser Pro 435 440 445

Val Ile Ile Asn Thr Ala Pro Cys Leu Phe Ser Asp Gly Ser Thr Ser 450 455 460

Arg Ala Ala Tyr Ile Leu Trp Asp Lys Gln Ile Leu Ser Gln Arg Ser 465 470 475 480

Phe Pro Leu Pro Pro Pro His Lys Ser Ala Gln Arg Ala Glu Leu Leu 485 490 495

Gly Leu Leu His Gly Leu Ser Ser Ala Arg Ser Trp Arg Cys Leu Asn 500 505 510

Ile Phe Leu Asp Ser Lys Tyr Leu Tyr His Tyr Leu Arg Thr Leu Ala 515 520 525

Leu Gly Thr Phe Gln Gly Arg Ser Ser Gln Ala Pro Phe Gln Ala Leu 530 535 540

Leu Pro Arg Leu Leu Ser Arg Lys Val Val Tyr Leu His His Val Arg 545 550 555

Ser His Thr Asn Leu Pro Asp Pro Ile Ser Arg Leu Asn Ala Leu Thr 565 570 575

Asp Ala

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 555 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Asn Val Leu Tyr Arg Ile Gly Ser Asp Asn Gln Tyr Thr Gln Phe Thr 1 5 10 15

Ile Pro Lys Lys Gly Lys Gly Val Arg Thr Ile Ser Ala Pro Thr Asp 20 25 30

Arg Leu Lys Asp Ile Gln Arg Arg Ile Cys Asp Leu Leu Ser Asp Cys Arg Asp Glu Ile Phe Ala Ile Arg Lys Ile Ser Asn Asn Tyr Ser Phe Gly Phe Glu Arg Gly Lys Ser Ile Ile Leu Asn Ala Tyr Lys His Arg Gly Lys Gln Ile Ile Leu Asn Ile Asp Leu Lys Asp Phe Phe Glu Ser Phe Asn Phe Gly Arg Val Arg Gly Tyr Phe Leu Ser Asn Gln Asp Phe 100 Leu Leu Asn Pro Val Val Ala Thr Thr Leu Ala Lys Ala Ala Cys Tyr 120 Asn Gly Thr Leu Pro Gln Gly Ser Pro Cys Ser Pro Ile Ile Ser Asn Leu Ile Cys Asn Ile Met Asp Met Arg Leu Ala Lys Leu Ala Lys Lys 155 145 Tyr Gly Cys Thr Tyr Ser Arg Tyr Ala Asp Asp Ile Thr Ile Ser Thr 170 Asn Lys Asn Thr Phe Pro Leu Glu Met Ala Thr Val Gln Pro Glu Gly 185 Val Val Leu Gly Lys Val Leu Val Lys Glu Ile Glu Asn Ser Gly Phe 195 Glu Ile Asn Asp Ser Lys Thr Arg Leu Thr Tyr Lys Thr Ser Arg Gln 210 215 Glu Val Thr Gly Leu Thr Val Asn Arg Ile Val Asn Ile Asp Arg Cys Tyr Tyr Lys Lys Thr Arq Ala Leu Ala His Ala Leu Tyr Arg Thr Gly Glu Tyr Lys Val Pro Asp Glu Asn Gly Val Leu Val Ser Gly Gly Leu 260 265 Asp Lys Leu Glu Gly Met Phe Gly Phe Ile Asp Gln Val Asp Lys Phe 280 Asn Asn Ile Lys Lys Lys Leu Asn Lys Gln Pro Asp Arg Tyr Val Leu 290 295 Thr Asn Ala Thr Leu His Gly Phe Lys Leu Lys Leu Asn Ala Arq Glu 305 310

Lys Ala Tyr Ser Lys Phe Ile Tyr Tyr Lys Phe Phe His Gly Asn Thr Cys Pro Thr Ile Ile Thr Glu Gly Lys Thr Asp Arg Ile Tyr Leu Lys 340 Ala Ala Leu His Ser Leu Glu Thr Ser Tyr Pro Glu Leu Phe Arg Glu 360 Lys Thr Asp Ser Lys Lys Glu Ile Asn Leu Asn Ile Phe Lys Ser Asn Glu Lys Thr Lys Tyr Phe Leu Asp Leu Ser Gly Gly Thr Ala Asp 390 Leu Lys Lys Phe Val Glu Arg Tyr Lys Asn Asn Tyr Ala Ser Tyr Tyr 410 Gly Ser Val Pro Lys Gln Pro Val Ile Met Val Leu Asp Asn Asp Thr 425 Gly Pro Ser Asp Leu Leu Asn Phe Leu Arg Asn Lys Val Lys Ser Cys 435 Pro Asp Asp Val Thr Glu Met Arg Lys Met Lys Tyr Ile His Val Phe 455 Tyr Asn Leu Tyr Ile Val Leu Thr Pro Leu Ser Pro Ser Gly Glu Gln 470 475 Thr Ser Met Glu Asp Leu Phe Pro Lys Asp Ile Leu Asp Ile Lys Ile 485 490 495 Asp Gly Lys Lys Phe Asn Lys Asn Asn Asp Gly Asp Ser Lys Thr Glu 500 505 Tyr Gly Lys His Ile Phe Ser Met Arg Val Val Arg Asp Lys Lys Arg 520 Lys Ile Asp Phe Lys Ala Phe Cys Cys Ile Phe Asp Ala Ile Lys Asp 530 Ile Lys Glu His Tyr Lys Leu Met Leu Asn Ser

555

(2) INFORMATION FOR SEQ ID NO:30:

545

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 243 amino acids

550

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Arg Trp Phe Ser Phe His Arg Glu Val Asp Thr Gly Thr His Tyr Gln
1 10 15

Thr Trp Glu Ile Pro Lys Arg Asp Gly Gly Lys Arg Thr Leu Thr Ala 20 25 30

Pro Lys Arg Glu Leu Lys Ala Val Gln Arg Trp Val Leu Ala Asn Val 35 40 45

Val Glu Arg Leu Pro Val His Gly Ala Ala His Gly Phe Val Ala Gly 50 55 60

Arg Ser Ile Leu Thr Asn Ala Leu Ala His Gln Gly Ala Asp Val Val 65 70 75 80

Val Lys Val Asp Met Lys Asp Phe Phe Pro Ser Val Thr Trp Pro Arg 85 90 95

Val Lys Gly Leu Leu Arg Lys Gly Gly Leu Pro Glu Asn Leu Ala Thr

Leu Leu Ala Leu Leu Ser Thr Glu Ala Pro Arg Glu Val Val Arg Phe
115 120 125

Arg Gly Glu Thr Leu Tyr Val Ala Lys Gly Pro Arg Ala Leu Pro Gln
130 135 140

Gly Ala Pro Thr Ser Pro Ala Leu Thr Asn Ala Leu Cys Leu Arg Leu 145 150 155 160

Asp Lys Arg Leu Ser Ala Leu Ser Lys Arg Leu Gly Phe Thr Tyr Thr 165 170 175

Arg Tyr Ala Asp Asp Leu Thr Phe Ser Trp Arg Arg Ala Lys Lys Ser 180 185 190

Arg Gln Lys Glu Leu Pro Leu Ala Asp Ala Pro Val Ala Leu Leu Leu 195 200 205

Ala Arg Val Lys Gly Val Leu Glu Ala Glu Gly Phe Thr Leu His Pro 210 215 220

Asp Lys Thr Arg Val Gln Arg Lys Gly Ser Arg Gln Arg Val Thr Gly 225 230 235 240

Leu Val Val

- (2) INFORMATION FOR SEQ ID NO:31:
 - (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 241 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Arg Trp Phe Ala Phe His Arg Glu Val Asp Thr Ala Thr His Tyr Val 1 5 10 15

Ser Trp Thr Ile Pro Lys Arg Asp Gly Ser Lys Arg Thr Ile Thr Ser 20 25 30

Pro Lys Pro Glu Leu Lys Ala Ala Gln Arg Trp Val Leu Ser Asn Val 35 40 45

Val Glu Arg Leu Pro Val His Gly Ala Ala His Gly Phe Val Ala Gly
50 55 60

Arg Ser Ile Leu Thr Asn Ala Leu Ala His Gln Gly Ala Asp Val Val 65 70 75 80

Val Lys Val Asp Leu Lys Asp Phe Phe Pro Ser Val Thr Trp Arg Arg 85 90 95

Val Lys Gly Leu Arg Lys Gly Gly Leu Arg Glu Gly Thr Ser Thr
100 105 110

Leu Leu Ser Leu Leu Ser Thr Glu Ala Pro Arg Glu Ala Val Gln Phe
115 120 125

Pro Arg Glu Leu Leu His Val Ala Lys Gly Pro Arg Ala Leu Pro Gln
130 135 140

Gly Ala Pro Thr Ser Pro Gly Ile Thr Asn Ala Leu Cys Leu Lys Leu 145 150 155 160

Asp Lys Arg Leu Ser Ala Leu Ala Lys Arg Leu Gly Phe Thr Tyr Thr
165 170 175

Arg Tyr Ala Asp Asp Leu Thr Phe Ser Trp Thr Lys Ala Lys Gln Pro 180 185 190

Lys Pro Arg Arg Thr Gln Arg Pro Pro Val Ala Val Leu Leu Ser Arg 195 200 205

Val Gln Glu Val Val Glu Ala Glu Gly Phe Arg Val His Pro Asp Lys 210 215 220

Thr Arg Val Ala Arg Lys Gly Thr Arg Gln Arg Val Thr Gly Leu Val 225 230 235 240

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEOUENCE CHARACTERISTICS:
 - (A) LENGTH: 231 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Arg His Tyr Ser Ile His Arg Pro Arg Glu Arg Val Arg His Tyr Val

1 10 15

Thr Phe Ala Val Pro Lys Arg Ser Gly Gly Val Arg Leu Leu His Ala 20 25 30

Pro Lys Arg Arg Leu Lys Ala Leu Gln Arg Arg Met Leu Ala Leu Leu 35 40 45

Val Ser Lys Leu Pro Val Ser Pro Gln Ala His Gly Phe Val Pro Gly 50 55 60

Arg Ser Ile Lys Thr Gly Ala Ala Pro His Val Gly Arg Arg Val Val 65 70 75 80

Leu Lys Leu Asp Leu Lys Asp Phe Phe Pro Ser Val Thr Phe Ala Arg
85 90 95

Val Arg Gly Leu Leu Lys Ala Leu Gly Tyr Gly Tyr Pro Val Ala Ala 100 105 110

Thr Leu Ala Val Leu Met Thr Glu Ser Glu Arg Gln Pro Val Glu Leu 115 120 125

Glu Gly Ile Leu Phe His Val Pro Val Gly Pro Arg Val Cys Val Gln
130 135 140

Gly Ala Pro Thr Ser Pro Ala Leu Cys Asn Ala Val Leu Leu Arg Leu 145 150 155 160

Asp Arg Arg Leu Ala Gly Leu Ala Arg Arg Tyr Gly Tyr Thr Tyr Thr 165 170 175

Arg Tyr Ala Asp Asp Leu Thr Phe Ser Gly Asp Asp Val Thr Ala Leu 180 185 190

Glu Arg Val Arg Ala Leu Ala Ala Arg Tyr Val Gln Glu Glu Gly Phe 195 200 205 Glu Val Asn Arg Glu Lys Thr Arg Val Gln Arg Arg Gly Gly Ala Gln 210 215 220

Arg Val Thr Gly Val Thr Val 225 230

- (2) INFORMATION FOR SEQ ID NO:33:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 234 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Phe Leu Thr Asn Val Leu Tyr Arg Ile Gly Ser Asp Asn Gln Tyr Thr

Gln Phe Thr Ile Pro Lys Lys Gly Lys Gly Val Arg Thr Ile Ser Ala 20 25 30

Pro Thr Asp Arg Leu Lys Asp Ile Gln Arg Arg Ile Cys Asp Leu Leu 35 40 45

Ser Asp Cys Arg Asp Glu Ile Phe Ala Ile Arg Lys Ile Ser Asn Asn 50 55 60

Tyr Ser Phe Gly Phe Glu Arg Gly Lys Ser Ile Ile Leu Asn Ala Tyr 65 70 75 80

Lys His Arg Gly Lys Gln Ile Ile Leu Asn Ile Asp Leu Lys Asp Phe 85 90 95

Phe Glu Ser Phe Asn Phe Gly Arg Val Arg Gly Tyr Phe Leu Ser Asn 100 105 110

Gln Asp Phe Leu Leu Asn Pro Val Val Ala Thr Thr Leu Ala Lys Ala 115 120 125

Ala Cys Tyr Asn Gly Thr Leu Pro Gln Gly Ser Pro Cys Ser Pro Ile 130 135 140

Ile Ser Asn Leu Ile Cys Asn Ile Met Asp Met Arg Leu Ala Lys Leu 145 150 155 160

Ala Lys Lys Tyr Gly Cys Thr Tyr Ser Arg Tyr Ala Asp Asp Ile Thr 165 170 175

Ile Ser Thr Asn Lys Asn Thr Phe Pro Leu Glu Met Ala Thr Val Gln 180 185 190

Pro Glu Gly Val Val Leu Gly Lys Val Leu Val Lys Glu Ile Glu Asn 195 200 205

Ser Gly Phe Glu Ile Asn Asp Ser Lys Thr Arg Leu Thr Tyr Lys Thr 210 220

Ser Arg Gln Glu Val Thr Gly Leu Thr Val 225 230

- (2) INFORMATION FOR SEQ ID NO:34:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 215 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
 - Val Glu Thr Leu Arg Leu Leu Ile Tyr Thr Ala Asp Phe Arg Tyr Arg
 - Ile Tyr Thr Val Glu Lys Lys Gly Pro Glu Lys Arg Met Arg Thr Ile 20 25 30
 - Tyr Gln Pro Ser Arg Glu Leu Lys Ala Leu Gln Gly Trp Val Leu Arg
 35 40 45
 - Asn Ile Leu Asp Lys Leu Ser Ser Ser Pro Phe Ser Ile Gly Phe Glu 50 55 60
 - Lys His Gln Ser Ile Leu Asn Asn Ala Thr Pro His Ile Gly Ala Asn 65 70 75 80
 - Phe Ile Leu Asn Ile Asp Leu Glu Asp Phe Phe Pro Ser Leu Thr Ala 85 90 95
 - Asn Lys Val Phe Gly Val Phe His Ser Leu Gly Tyr Asn Arg Leu Ile 100 105 110
 - Ser Ser Val Leu Thr Lys Ile Cys Cys Tyr Lys Asn Leu Leu Pro Gln
 115 120 125
 - Gly Ala Pro Ser Ser Pro Lys Leu Ala Asn Leu Ile Cys Ser Lys Leu 130 135 140
 - Asp Tyr Arg Ile Gln Gly Tyr Ala Gly Ser Arg Gly Leu Ile Tyr Thr 145 150 155 160
 - Arg Tyr Ala Asp Asp Leu Thr Leu Ser Ala Gln Ser Met Lys Lys Val 165 170 175

Val Lys Ala Arg Asp Phe Leu Phe Ser Ile Ile Pro Ser Glu Gly Leu 180 185 190

Val Ile Asn Ser Lys Lys Thr Cys Ile Ser Gly Pro Arg Ser Gln Arg 195 200 205

Lys Val Thr Gly Leu Val Ile 210 215

- (2) INFORMATION FOR SEQ ID NO:35:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 230 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Thr Lys Gly Phe Ala Ser Glu Val Met Arg Ser Pro Glu Pro Pro Lys
1 10 15

Lys Trp Asp Ile Ala Lys Lys Lys Gly Gly Met Arg Thr Ile Tyr His 20 25 30

Pro Ser Ser Lys Val Lys Leu Ile Gln Tyr Trp Leu Met Asn Asn Val

Phe Ser Lys Leu Pro Met His Asn Ala Ala Tyr Ala Phe Val Lys Asn 50 55

Arg Ser Ile Lys Ser Asn Ala Leu Leu His Ala Glu Ser Lys Asn Lys 65 70 75 80

Tyr Tyr Val Lys Ile Asp Leu Lys Asp Phe Phe Pro Ser Ile Lys Phe 85 90 95

Thr Asp Phe Glu Tyr Ala Phe Thr Arg Tyr Arg Asp Arg Ile Glu Phe 100 105 110

Thr Thr Glu Tyr Asp Leu Glu Leu Leu Gln Leu Ile Lys Thr Ile Cys 115 120 125

Phe Ile Ser Asp Ser Thr Leu Pro Ile Gly Phe Pro Thr Ser Pro Leu 130 135 140

Ile Ala Asn Phe Val Ala Arg Glu Leu Asp Glu Lys Leu Thr Gln Lys 145 150 155 160

Leu Asn Ala Ile Asp Lys Leu Asn Ala Thr Tyr Thr Arg Tyr Ala Asp 165 170 175 Asp Ile Ile Val Ser Thr Asn Met Lys Gly Ala Ser Lys Leu Ile Leu 180 185 190

Asp Cys Phe Lys Arg Thr Met Lys Glu Ile Gly Pro Asp Phe Lys Ile 195 200 205

Asn Ile Lys Lys Phe Lys Ile Cys Ser Ala Ser Gly Gly Ser Ile Val 210 215 220

Val Thr Gly Leu Lys Val 225 230

- (2) INFORMATION FOR SEQ ID NO:36:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 211 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Ile Gln Arg Leu His Ala Leu Ser Asn His Ala Gly Arg His Tyr Arg

1 10 15

Arg Ile Ile Leu Ser Lys Arg His Gly Gly Gln Arg Leu Val Leu Ala 20 25 30

Pro Asp Tyr Leu Leu Lys Thr Val Gln Arg Asn Ile Leu Lys Asn Val 35 40 45

Leu Ser Gln Phe Pro Leu Ser Pro Phe Ala Thr Ala Tyr Arg Pro Gly 50 55 60

Cys Pro Ile Val Ser Asn Ala Gln Pro His Cys Gln Gln Pro Gln Ile 65 70 75 80

Leu Lys Leu Asp Ile Glu Asn Phe Phe Asp Ser Ile Ser Trp Leu Gln
85 90 95

Val Trp Arg Val Phe Arg Gln Ala Gln Leu Pro Arg Asn Val Val Thr
100 105 110

Met Leu Thr Trp Ile Cys Cys Tyr Asn Asp Ala Leu Pro Gln Gly Ala 115 120 125

Pro Thr Ser Pro Ala Ile Ser Asn Leu Val Met Arg Arg Phe Asp Glu
130 135 140

Arg Ile Gly Glu Trp Cys Gln Ala Arg Gly Ile Thr Tyr Thr Arg Tyr 145 150 155 160

		55					60					65					
Į	GC Arg 70	CGC Arg	TAC Tyr	ACC Thr	CCG Pro	GGC Gly 75	CGG Arg	AAG Lys	AAG Lys	TGG Trp	ATG Met 80	GAG Glu	GCC Ala	GCC Ala	GAG Glu	GCC Ala 85	533
Į	CGG Arg	CGG Arg	CTG Leu	TTC Phe	TCC Ser 90	GCC Ala	ACG Thr	CTG Leu	CGC Arg	ACG Thr 95	CGG Arg	AAC Asn	CGG Arg	AAC Asn	CTG Leu 100	AGG Arg	581
Į	Asp	TTG Leu	CTG Leu	CCC Pro 105	GAC Asp	GAG Glu	GCA Ala	CAG Gln	CTG Leu 110	GCG Ala	CGC Arg	TAC Tyr	GGC Gly	CTG Leu 115	CCG Pro	GTC Val	629
7) -	rgg Frp	CGC Arg	ACG Thr 120	GAA Glu	GAG Glu	GAC Asp	GTG Val	GCA Ala 125	GCG Ala	GCC Ala	CTG Leu	GGC Gly	GTC Val 130	TCG Ser	GTG Val	GGC Gly	677
· Piling	Val	Leu 135	Arg	His	TAC Tyr	Ser	Ile 140	His	Arg	Pro	Arg	G1u 145	Arg	vaı	arg	HIS	725
	TAC Tyr 150	GTG Val	ACC Thr	TTC Phe	GCC Ala	GTG Val 155	CCC Pro	AAG Lys	CGC Arg	TCC Ser	GGA Gly 160	GGC Gly	GTC Val	CGG Arg	CTG Leu	CTG Leu 165	773
Total (САТ	GCG	CCC	AAG	CGG Arg 170	CGC	CTG	AAG	GCC	CTG	CAA	CGC	CGG	ATG	CTG	GCG	821
	CTC Leu	CTG Leu	GTG Val	TCG Ser 185	AAG Lys	CTC Leu	CCC Pro	GTG Val	AGT Ser 190	CCA Pro	CAG Gln	GCC Ala	CAT His	GGC Gly 195	TTC Phe	GTG Val	869
Marie The Mines	CCC Pro	GGC Gly	CGC Arg 200	TCC Ser	ATC Ile	AAG Lys	ACG Thr	GGC Gly 205	GCC Ala	GCG Ala	CCG Pro	CAC His	GTG Val 210	GTÄ	CGG Arg	CGG Arg	917
	GTG Val	GTC Val 215	CTG Leu	AAG Lys	CTG Leu	GAC Asp	CTG Leu 220	AAG Lys	GAC Asp	TTC Phe	TTC Phe	CCC Pro 225	ser	GTC Val	ACC Thr	TTC Phe	965
	GCG Ala 230	CGG Arg	GTG Val	CGA Arg	GGG Gly	CTG Leu 235	CTC Leu	ATC Ile	GCC Ala	CTG Leu	GGC Gly 240	Tyr	GGC Gly	TAT Tyr	CCC Pro	GTG Val 245	1013
	GCG Ala	GCC Ala	ACG Thr	CTC Leu	GCG Ala 250	Val	CTG Leu	ATG Met	ACG Thr	GAG Glu 255	TCC Ser	GAG Glu	CGC Arg	CAG Gln	CCC Pro 260	GTG Val	1061
	GAG Glu	CTG Leu	GAG Glu	GGC Gly 265	Ile	CTC Leu	TTC Phe	CAC His	GTT Val 270	Pro	GTG Val	GGC Gly	CCA Pro	CGC Arg 275	vaı	TGC Cys	1109

Cys Asp Asp Met Thr Phe Ser Gly His Phe Asn Ala Arg Gln Val Lys 175

Asn Lys Val Cys Gly Leu Leu Ala Glu Leu Gly Leu Ser Leu Asn Lys 185

Arg Lys Gly Cys Leu Ile Ala Ala Cys Lys Arg Gln Gln Val Thr Gly 195

Ile Val Val 210

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1640 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 279..1559

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

60	GCGGCG	ACGCG	CGG A	3GGG	GCG	GCG	CCCGC	C GG(CTCG	C'GCG(G A	CCGA	GCCT(CCC (CGAG	CTC
120	GCTCTC	AGGT0	GGC I	CAAC	CGA	ATGA	ACGAI	A CGA	GGAG2	CCCG	rg a	CGCT:	GAGA(ACG (GCCC	GCG
180	GGTGT	GCGGG	GCC (rTTC(GTG:	CGCG	FTAC	r gao	GCCA!	ATGA	CA G	CTCG	AGGG	GCC 1	AGAG	GGG
240	GTCCAA	CCGGG	GCC (GGA	GCA	CAAC	racg(G CG	CCCA	GGGT(CC A	rtcg	rcrc:	CCA '	TCC	TCT(
293		rc GA ne As					AGCA	C GG1	CTTC(GCCT	CT G	rccc	GTCG"	CAG (CTCG(CGC(
341	CGA Arg															
389	CAG Gln															
437	GGG Gly															
485	GTC Val												CAG Gln			

		CAG Gln															1157
		CTG Leu 295															1205
		ACG Thr															1253
		CTG Leu															1301
.1		TTC Phe															1349
	Ala	CAG Gln															1397
		GAG Glu 375															1445
		GAG Glu															1493
The state of the s		GTG Val															1541
1		AAG Lys					TGA	GCGA(GGG (CTCAC	GCTC	CG G1	ATGG(GCCA(ġ.		1589
	GGC	CTGT	CAC (GCGT(CCCG	GC C	rccc2	AGTT	G TC	ATGG(CGGC	CGT	CCCA	STA (J		1640

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3060 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 763..2202

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

	CCCZ	ACTT(CCG (GCGC:	rcgg(C TO	GCGC	GAGGG	G CC	CGTG	CGAG	CAC	ATGA:	rgg	CGCT	GCGGCT	60
	CGT	CCAG	GTC (CGGCI	ACCG	CG C	CGAG	CAGGA	A AG	CACT	GCGT	CAG	ACCC	CCG	CGGG	CCGCCA	120
	GCT	CATC	CGC (GCGGI	AGAC	GC G	CTCC	racgi	GC(GCG(CGAG	CCC	rccg	SCC.	AGGA	GCAGGT	180
	GTA	CCGC	GTC '	TCAT:	rgga:	rg go	GAAA(GTGGI	GG(CGGT	GGAG	TGG	GCC	CCC ·	GCCA	egggga	240
	GTC	CCGC	CGG (CAGA	AGCT	CT GO	GTTC	GACAC	C GGZ	ACGC(CGAG	GCG	CGCA	CCG	CCTA	CTTCAC	300
	GCG	CCTG	GAG '	TCCT:	rggco	CG C	GGAG	GGATA	A TAT	rcga:	rgcg	GCT	GCTT(CAA '	TGAT	STAGAA	360
A	CAC	GCAA	GCC I	ACGG(GCC(GC G	GGCG(CGCGG	G CGC	GAAA(GCA	GGT	GCGA(CGG .	AACG	ACAGAC	420
V_	ACTO	CGTG	CGA (GCGA	CCGAC	BA G	AGGT	CCCAP	A GC	CATC	AGCC	TCAC	3CGC(CTC	GAGC	GCGAGA	480
	GCG	GCGT	rgc (GCCG	CTCTC	G T	rgaa:	FTGCF	A GGZ	ACAC	rctc	CGC	AAGG"	rag	CCTG	TCTTG	540
	GCT	CTCT:	rcc (CTCC	GGTG <i>I</i>	AG T	ACCT	CTCCC	GC(CGGG	GAGC	TGA	ACCAZ	ACG .	ACGC	ACCGC	600
	CGT	rtcc	CCG (GCCG	GAGAC	G T	ACTC	ACCGG	G AGO	GGA	GAGC	CGG:	rgag(GCT .	ACCG'	rgcccc	660
	AGG"	rgag <i>i</i>	AAG (GTGG:	rgcc1	T C	GGC(CTCCC	C TCC	GACC(GCTC	GCG	CTCCC	STC ·	GCCC	FGCCCT	720
Half day ton	GCC:	rcgc(CCC (ccca	ACCTT	TG C	rcac(CGGCG	G CCI	AGGA(GCCG				GCC A Ala I		774
A May Long															GCG Ala		822
															GCC Ala 35		870
															GGC Gly		918
															GCG Ala		966
															GCC Ala		1014
	AAG	GAG	AAG	AAG	AAG	GCC	GAG	GCC	ACC	GAG	CGG	CGC	GCG	CAG	AAG	CGC	1062

	Lys 85	Glu	Lys	Lys	Lys	Ala 90	Glu	Ala	Thr	Glu	Arg 95	Arg	Ala	Gln	Lys	Arg 100	
									ACG Thr								1110
									GGG Gly 125								1158
									AAC Asn								1206
									CTG Leu								1254
									GTG Val								1302
									Gly								1350
									CAG Gln 205								1398
- 55									GCC Ala								1446
						Asn			GCC Ala								1494
									TTC Phe								1542
									GGA Gly								1590
									GCC Ala 285								1638
									AAG Lys								1686

							GCG Ala 315									CTG Leu	1734	
							CTG Leu										1782	
							ACG Thr										1830	
							CTG Leu										1878	
							CTG Leu										1926	
	GAC Asp	AAG Lys 390	ACG Thr	CGG Arg	GTG Val	CAG Gln	CGC Arg 395	AAG Lys	GGC Gly	AGC Ser	CGG Arg	CAG Gln 400	CGG Arg	GTG Val	ACG Thr	GGG Gly	1974	
	CTC Leu 405																2022	
							CTG Leu										2070	
							GGG Gly										2118	
·																CTG Leu	2166	
							GAG Glu 475						TGA	CCCT	CAC		2212	
	TGGT	rcgr	CCG (GGC1	ATCGO	CA GO	CGGGC	CGCCC	G GGF	ACGGI	ACCG	TCAC	cccc	CCA (BATC	CCATG	2272	
	CCAT	rgctc	GG (TTAE	CTGGC	SC GC	STGA	AGAAG	ACI	TCCC	CAGC	CGAG	SACGO	BAC (GAAG	CCCTGC	2332	
	GGAT	rccgi	ATG A	ACTC(CTCGC	cc co	GGGG	CGATO	C TCC	CCGGA	AGGG	GCAC	CCGTT	rcc (BACG"	CCGTG	2392	
	CCAT	rtgct	rca (CCCAC	GGC7	TC CC	CGGCC	CCCAC	G CCI	TGGC	STGT	CCGC	CCGAC	AA (SAAG2	AGCAGC	2452	
	CCG	GAGAT	rgg (CCGT	CAGGT	T CI	rccg(GCGA	C GCF	ATCCI	rcgg	GGC	CCGG	CGC (CAAAT	CCTTC	2512	

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AGCAGCAGGG	TGCCCTTGGC	GGTGCCATCG	CTGGACCACA	GCTCCCGGCC	GTGGAGGCTG	2572
TCACTCGCGG	CGAAGTAGAG	CATCCCATTC	AGCGCCTTGA	TGGCGCTGGG	CGCCGAGCTG	2632
TCCGGACCCG	GCCAGATGTC	CTTCACCCGG	ACCGTGCCAT	GCGACGTGCC	ATCGCTGACC	2692
CACAGCTCCT	CGCCCTCGGG	CTGGCCCCAG	AACTCGGGCT	CGCCTCCCCC	GGCGCTGAAG	2752
AAGATCTTCC	CCCCGAGCGC	CGTGAGATCA	TGCGGATAGA	GGCCGGGGAA	GAAGCGCAGC	2812
TGCTCGGAGA	CGGTGCCTCT	GGAGCACCAC	AGGCTGGCCT	CGCCTTCGTC	ATTGTCGAGC	2872
AGGAAGAAGA	GCACCGAGTC	CGCCGCGGTG	AACGCGGAGA	GGAAGTTGTC	CTCGGGGCCC	2932
GTGAAGACAG	ACGTGGTGCT	GGACAGCCCC	AGGCTGCGCC	AGATGAACAC	CTCGTCATTG	2992
ACGTTGGCCA	CGAAGAAGAG	CGCATCGCCG	ACCCGGGTGA	GCCGGCGCGG	GCTGGAGCTG	3052
CCGGGCAC						3060

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2788 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 2..103
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 707..1654
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1644..2591
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:
- T TTC GAG AAG CGC CAT ACC AAA CAG GGG ATA CAG ACC AAC CTG ACG
 Phe Glu Lys Arg His Thr Lys Gln Gly Ile Gln Thr Asn Leu Thr
 1 10 15
- CTG AAA GAG GAA AGC TAC GGC GAC TGG CTG CCG AAG TGC GAC GAC CCC
 Leu Lys Glu Glu Ser Tyr Gly Asp Trp Leu Pro Lys Cys Asp Asp Pro
 20 25 30

143

GCA GCA ACA TAACCTCACT CAGACCGGCA ACAGCCGGTC TTTTCCTTTC

Ala Ala Thr

		TGGC	CATT	GC C	ACAA	GGTG	A AC	AATC	CACT	GTT	CACC	CTT	CACC	GTTT.	AT T	CACC	CTTTA	203
		TCAC	TATG	AA A	TAT	TAAT	A AA	AAAC	CAGA	GGT	GAAC	AGT	GTGA	ACAG	TA A	AACC	TGAAA	263
		AAAC	TTTT	TA T	'CACC	CCGC	G CA	TCGC	CCGA	CTG	GACA	GAT	CCAG	AACG	AG C	AAAA	ATCAC	323
		AAAG	GTGA	CG A	.GTCG	ACTG	T TC	ACTC	TTCA	CCA	ACTC	ATC	ACCA	.CCTA	AC C	ACAT	GATAT	383
		AAAA	TGAT	AA A	TAAT	'CGAG	G TG	AACA	GTTA	AAT	GCAA	AAA	AACT	TTTT	CT C	AGCT	CTTGG	443
		ATAA	AAGA	AA A	AATT	TTCA	.C AT	'CAA'I	'AGCT	TTC	CTCT	TGA	ATCC	TCTT	GA G	GTTT	ATGAG	503
		AGCG	TAAC	AG A	.GCCA	AACC	T AG	CAT'I	TATT	' GGG	AATT	TAG	CCCA	TCGC	GC A	TGAG	TCATG	563
		GTTT	CGCC	TA G	TATT	TTAG	C TA	TGCC	CGTC	GTT	'CAGT	TCG	CTGA	.GCGG	CG G	CTGG	GGGCC	623
N	1	ACCG	ATCA	.GC G	AACT	GATC	G AC	GTGC	TCAA	GTA	.GGTT	TGG	CTCT	TTTA	GT C	CTCT	'ACCAT	683
- \\ /	A				AGGA			G AT		'G AC	T CA	G CI	'A AA	AA AA	A AA	T GG	T	733
	25.25	ACT Thr 10	GAG Glu	GTA Val	TCT Ser	AGA Arg	GCA Ala 15	ACC Thr	GCG Ala	TTA Leu	TTT Phe	TCA Ser 20	TCA Ser	TTC Phe	GTT Val	GAA Glu	AAG Lys 25	781
		AAC Asn	AAA Lys	GTA Val	AAA Lys	TGT Cys 30	CCT Pro	GGT Gly	AAT Asn	GTA Val	AAA Lys 35	AAA Lys	TTC Phe	GTC Val	TTT Phe	CTG Leu 40	TGT Cys	829
	Marie The State of	GGT Gly	GCT Ala	AAC Asn	AAA Lys 45	AAC Asn	AAT Asn	GGA Gly	GAA Glu	CCA Pro 50	TCA Ser	GCA Ala	AGA Arg	CGA Arg	TTG Leu 55	GAA Glu	TTA Leu	877
		ATA Ile	AAT Asn	TTT Phe 60	TCT Ser	GAA Glu	Arg	Tyr	Leu	AAT Asn	AAC Asn	TGT Cys	CAC His	TTT Phe 70	TTT Phe	CTT Leu	GCT Ala	925
		GAA Glu	Leu	GTT Val	TTC Phe	AAA Lys	GAA Glu	TTA Leu 80	AGC Ser	ACC Thr	GAT Asp	GAA Glu	GAA Glu 85	TCA Ser	TTA Leu	TCT Ser	GAT Asp	973
		AAT Asn 90	TTA Leu	TTA Leu	GAT Asp	ATC Ile	GAA Glu 95	GCT Ala	GAC Asp	TTA Leu	TCT Ser	AAA Lys 100	TTA Leu	GCT Ala	GAT Asp	CAT His	ATT Ile 105	1021
		ATC Ile	ATT Ile	GTT Val	TTA Leu	GAA Glu 110	AGT Ser	TAT Tyr	TCA Ser	TCT Ser	TTC Phe 115	ACG Thr	GAA Glu	CTT Leu	GGT Gly	GCA Ala 120	TTC Phe	1069
		GCA	TAC	AGC	AAG	CAA	TTA	CGC	AAG	AAA	TTA	ATA	ATA	GTT	AAC	AAT	ACA	1117

	Ala	Tyr	Ser	Lys 125	Gln	Leu	Arg	Lys	Lys 130	Leu	Ile	Ile	Val	Asn 135	Asn	Thr	
	AAA Lys	TTT Phe	ATA Ile 140	AAT Asn	GAG Glu	AAA Lys	TCA Ser	TTT Phe 145	ATA Ile	AAT Asn	ATG Met	GGA Gly	CCA Pro 150	ATA Ile	AAG Lys	GCT Ala	1165
÷	ATT Ile	ACT Thr 155	Gln	CAA Gln	TCA Ser	CAA Gln	CAA Gln 160	TCT Ser	GGT Gly	CAT His	TTC Phe	TTA Leu 165	CAT His	TAT Tyr	AAA Lys	ATG Met	1213
	ACA Thr 170	GAA Glu	GGT Gly	ATT Ile	GAA Glu	AGT Ser 175	ATA Ile	GAG Glu	CGC Arg	TCT Ser	GAT Asp 180	GGG Gly	ATT Ile	GGC Gly	GAA Glu	ATA Ile 185	1261
	TTC Phe	GAC Asp	CCC Pro	CTA Leu	TAT Tyr 190	GAT Asp	ATT Ile	CTT Leu	TCT Ser	AAG Lys 195	AAC Asn	GAC Asp	AGA Arg	GCA Ala	ATT Ile 200	TCA Ser	1309
Me	Arg	ACT Thr	TTA Leu	AAA Lys 205	AAA Lys	GAA Glu	GAG Glu	TTA Leu	GAT Asp 210	CCT Pro	TCC Ser	AGT Ser	AAC Asn	TTC Phe 215	AAT Asn	AAA Lys	1357
	Asp	TCA Ser	GTA Val 220	CGA Arg	TTT Phe	ATT Ile	CAT His	GAC Asp 225	GTA Val	ATT Ile	TTT Phe	GTA Val	TGT Cys 230	GGT Gly	CCT Pro	TTG Leu	1405
und udu # 40	CAA Gln	CTT Leu 235	Asn	GAA Glu	CTC Leu	ATC Ile	GAA Glu 240	ATA Ile	ATC Ile	ACA Thr	AAA Lys	ATA Ile 245	Phe	GGC Gly	ACA Thr	GAA Glu	1453
	AGC Ser 250	His	TAC Tyr	AAA Lys	AAA Lys	AAT Asn 255	CTT Leu	CTA Leu	AAG Lys	CAC	CTT Leu 260	. Gly	ATT	CTA Leu	ATA Ile	GCT Ala 265	1501
. in	ידידע 🗓	AGA Arg	ATA Ile	ATA Ile	TCA Ser 270	Cys	ACA Thr	AAT Asn	GGG Gly	ATT Ile 275	Tyr	TAT Tyr	TCT Ser	TTG Leu	TAT Tyr 280	AAA Lys	1549
	GAA Glu	TAT Tyr	TAT Tyr	TTT Phe 285	: Lys	TAT Tyr	GAC Asp	TTI Phe	GAC Asp 290	ıle	' GAC : Asp	AAC Asn	ATA	T'CA Ser 295	Ser	ATG Met	1597
W.	TTT Phe	AAA Lys	GTT Val	. Phe	TTC Phe	: CTC : Leu	: AAG Lys	AAC Asr 305	ı Lys	CCA Pro	GAA Glu	AGG Arg	310	: Arg	GTA Val	A TAT Tyr	1645
			ı Ile		SCCTA	ATT	GATT	CTC	AGA C	CATTO	atg <i>i</i>	AC TA	AGGG	PTTA S	7		1694
	GCT	TCTC	AAG	TAAT	GCGA	ATC A	CCTG	GAGC	CG CC	AAAZ	LAAA	r ggd	CATA	TAGC	TAAC	SAAAAA	1754
	GGA	GGTA	ATGA	GAAC	CAATT	TA T	CACC	CCGT	CA TO	CAAAZ	AGTT	AA A	TAAT	TTCA	ATA	TTGGTTA	1814

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ATGAATAATG TTTTTTCGAA GCTCCCAATG CATAATGCTG CATATGCATT TGTTAAAAAC 1874 CGATCAATAA AAAGCAATGC TTTATTACAT GCCGAATCAA AGAATAAGTA TTATGTGAAA 1934 ATAGATCTCA AAGATTTTTT CCCTTCAATA AAATTTACTG ATTTTGAGTA CGCATTCACT 1994 CGTTATCGAG ATCGCATTGA ATTTACTACA GAATATGATA AGGAGTTACT ACAACTTATA 2054 AAAACGATCT GCTTTATATC AGATAGCACT CTCCCTATCG GGTTTCCTAC ATCTCCATTA 2114 ATTGCAAACT TTGTGGCAAG AGAACTTGAT GAAAAACTGA CGCAAAAACT AAATGCAATT 2174 GATAAACTTA ATGCCACTTA TACACGATAT GCTGATGATA TTATTGTCTC TACAAATATG 2234 AAAGGGGCTA GCAAATTAAT TCTGGATTGT TTTAAAAGAA CAATGAAAGA GATTGGTCCA 2294 GACTTTAAAA TTAACATTAA AAAATTTAAG ATTTGTAGTG CTTCGGGAGG AAGTATAGTA 2354 GTTACCGGAT TGAAAGTTTG CCACGATTTT CATATTACAT TACATAGATC AATGAAAGAT 2414 AAAATAAGAT TGCATCTTTC TCTTTTATCA AAGGGCATAT TAAAAGATGA AGATCATAAT 2474 AAACTTTCTG GTTATATTGC TTATGCAAAA GATATAGACC CTCATTTTTA TACAAAACTG 2534 AACAGAAAT ATTTTCAAGA AATAAAATGG ATTCAGAATC TCCACAACAA AGTTGAATAA 2594 ACTTTATATT TTGGATGCAC CCCAATAACT TCATTGATTA AATTGGGAAC AATATAGGCT 2654 TTTCAGGATG ACCTACACTC TAGAGAATGT GTATACAAAA GTGTATAAGT TATTTTCAAA 2714 2774 CCTATATAAA ATACAGCAAA ATCAATGCAT TGGCGGCATT TTACCACTCC TGTGATCTTC CGCCAAAATG CCTC 2788

- (2) INFORMATION FOR SEQ ID NO:40:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 316 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Met Arg Ile Tyr Ser Leu Ile Asp Ser Gln Thr Leu Met Thr Lys Gly
1 10 15

Phe Ala Ser Glu Val Met Arg Ser Pro Glu Pro Pro Lys Lys Trp Asp 20 25 30

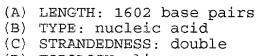
Ile Ala Lys Lys Gly Gly Met Arg Thr Ile Tyr His Pro Ser Ser 35 40 45

Lys Val Lys Leu Ile Gln Tyr Trp Leu Met Asn Asn Val Phe Ser Lys Leu Pro Met His Asn Ala Ala Tyr Ala Phe Val Lys Asn Arg Ser Ile Lys Ser Asn Ala Leu Leu His Ala Glu Ser Lys Asn Lys Tyr Tyr Val 90 Lys Ile Asp Leu Lys Asp Phe Phe Pro Ser Ile Lys Phe Thr Asp Phe 105 Glu Tyr Ala Phe Thr Arg Tyr Arg Asp Arg Ile Glu Phe Thr Thr Glu 125 120 115 Tyr Asp Lys Glu Leu Leu Gln Leu Ile Lys Thr Ile Cys Phe Ile Ser 135 Asp Ser Thr Leu Pro Ile Gly Phe Pro Thr Ser Pro Leu Ile Ala Asn 155 150 145 Phe Val Ala Arg Glu Leu Asp Glu Lys Leu Thr Gln Lys Leu Asn Ala 170 165 Ile Asp Lys Leu Asn Ala Thr Tyr Thr Arg Tyr Ala Asp Asp Ile Ile 180 Val Ser Thr Asn Met Lys Gly Ala Ser Lys Leu Ile Leu Asp Cys Phe 200 Lys Arg Thr Met Lys Glu Ile Gly Pro Asp Phe Lys Ile Asn Ile Lys 210 Lys Phe Lys Ile Cys Ser Ala Ser Gly Gly Ser Ile Val Val Thr Gly 225 230 Leu Lys Val Cys His Asp Phe His Ile Thr Leu His Arg Ser Met Lys 250 Asp Lys Ile Arg Leu His Leu Ser Leu Leu Ser Lys Gly Ile Leu Lys 265 Asp Glu Asp His Asn Lys Leu Ser Gly Tyr Ile Ala Tyr Ala Lys Asp 280 275 Ile Asp Pro His Phe Tyr Thr Lys Leu Asn Arg Lys Tyr Phe Gln Glu 290

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

Ile Lys Trp Ile Gln Asn Leu His Asn Lys Val Glu 310



- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 548..1507

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

TGGCATCTAT TAAGAAGGTT AGGAAAGAAA ATAAAGTATC AAAAGATATT GGAAATATAT	60
TATACGCAGA GCGTTTCTAT TGCCTTGTAT CTATTTACTG GATAGTGTCA ACTACCGCAC	120
ACTGTGTGAA CTAGCTTTTA AAGCGATAAA GCAAGATGAT GTTTTATCTA AAATTATTGT	180
TAGATCCGTT GTTTCTCGTC TAATAAATGA ACGAAAAATA CTTCAAATGA CTGATGGTTA	240
TCAGGTCACT GCTTTGGGGG CTAGCTATGT TAGGAGCGTC TTTGATAGAA AGACACTTGA	300
CCGATTGCGG CTTGAGATTA TGAATTTTGA AAACCGTAGA AAATCAACAT TTAACTATGA	360
TAAGATTCCG TATGCGCACC CTTAGCGAGA GGTTTATCAT TAAGGTCAAC CTCTGGATGT	420
TGTTTCGGCA TCCTGCATTG AATCTGAGTT ACTGTCTGTT TTCCTTGTTG GAACGGAGAG	480
CATCGCCTGA TGCTCTCCGA GCCAACCAGG AAACCCGTTT TTTCTGACGT AAGGGTGCGC	540
AACTTTC ATG AAA TCC GCT GAA TAT TTG AAC ACT TTT AGA TTG AGA AAT Met Lys Ser Ala Glu Tyr Leu Asn Thr Phe Arg Leu Arg Asn 1 5 10	589
CTC GGC CTA CCT GTC ATG AAC AAT TTG CAT GAC ATG TCT AAG GCG ACT Leu Gly Leu Pro Val Met Asn Asn Leu His Asp Met Ser Lys Ala Thr 15 20 25 30	637
CGC ATA TCT GTT GAA ACA CTT CGG TTG TTA ATC TAT ACA GCT GAT TTT Arg Ile Ser Val Glu Thr Leu Arg Leu Leu Ile Tyr Thr Ala Asp Phe 35 40 45	685
CGC TAT AGG ATC TAC ACT GTA GAA AAG AAA GGC CCA GAG AAG AGA ATG Arg Tyr Arg Ile Tyr Thr Val Glu Lys Lys Gly Pro Glu Lys Arg Met 50 55 60	733
AGA ACC ATT TAC CAA CCT TCT CGA GAA CTT AAA GCC TTA CAA GGA TGG Arg Thr Ile Tyr Gln Pro Ser Arg Glu Leu Lys Ala Leu Gln Gly Trp 65 70 75	781
GTT CTA CGT AAC ATT TTA GAT AAA CTG TCG TCA TCT CCT TTT TCT ATT Val Leu Arg Asn Ile Leu Asp Lys Leu Ser Ser Pro Phe Ser Ile 80 85 90	829

					CAA Gln 100											877
					CTG Leu											925
					GTT Val											973
					GTT Val											1021
					CCA Pro											1069
TCT Ser 175	AAA Lys	CTT Leu	GAT Asp	TAT Tyr	CGT Arg 180	ATT Ile	CAG Gln	GGT Gly	TAT Tyr	GCA Ala 185	GGT Gly	AGT Ser	CGG Arg	GGC Gly	TTG Leu 190	1117
					GCC Ala											1165
					GCA Ala											1213
					AAC Asn											1261
					ACA Thr											1309
ATA Ile 255	GGT Gly	AGA Arg	GAA Glu	AAA Lys	TAT Tyr 260	AAA Lys	GAA Glu	ATT Ile	AGA Arg	GCA Ala 265	AAG Lys	ATA Ile	CAT His	CAT His	ATA Ile 270	1357
					TCT Ser											1405
					GAT Asp											1453
					AAA Lys											1501

AAG ACC TAATGGTCTT CGTTTTAAAA CTAAAGCTCA TAGGTTGAAA AATTGAGCAC Lys Thr 320

315

1602

1557

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1540 base pairs

TTCTTCGTCC AACCAGTTAT TTAGTTCCTG CAATCGTTTC TGCAG

310

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ix) FEATURE:

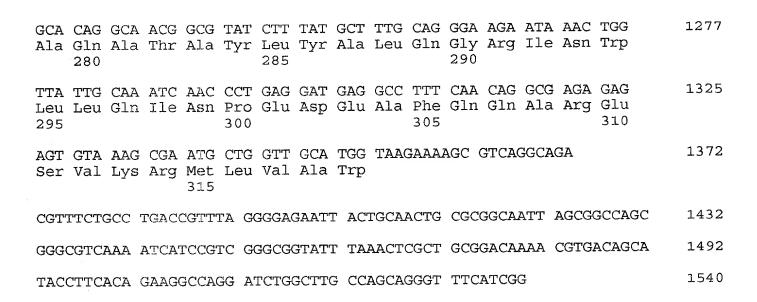
305

- (A) NAME/KEY: CDS
- (B) LOCATION: 396..1352

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

TCACCCTGAA AGACCTGATT GCTTACCTGG AAGAGAAGCC GGAAATGGCG GAACATCTGG	60
CGGCGGTTAA GGCCTATCGC GAAGAGTTCG GCGTTTAAAA ATATGCGCTG TGCAGGGTTT	120
TTGCTGTGCG CAGCGTGATG CGCTTCAAGA TATCGTGTTA ATCTGCTTTC GCCAGCAGTG	180
GCAATAGCGT TTCCGGCCTT TTGTGCCGGG AGGGTCGGCG AGTCGCTGAC TTAACGCCAG	240
TAGTATGTCC ATATACCCAA AGTCGCTTCA TTGTACCTGA GTACGCTTCG CGTACGTCGC	300
GCTGACGCGC TCAGTACAGT TACGCGCCTT CGGGATGGTT TAATGGTATT GCCGCTGTTG	360
GCGCCTCTTT TGGCCGCCGT GATGTGGAGA GTGGA ATG GAT GCT ACC CGG ACA Met Asp Ala Thr Arg Thr 1 5	413
ACC CTT CTG GCG CTC GAT TTG TTC GGC TCG CCG GGC TGG AGC GCC GAT Thr Leu Leu Ala Leu Asp Leu Phe Gly Ser Pro Gly Trp Ser Ala Asp 10 15 20	461
AAA GAA ATA CAG CGA CTG CAT GCG CTC AGT AAT CAT GCC GGA CGC CAT Lys Glu Ile Gln Arg Leu His Ala Leu Ser Asn His Ala Gly Arg His 25 30 35	509
TAC CGA CGC ATT ATT CTT TCT AAA CGC CAC GGT GGT CAG CGG CTG GTG Tyr Arg Arg Ile Ile Leu Ser Lys Arg His Gly Gly Gln Arg Leu Val 40 45 50	557
TTA GCC CCT GAT TAC TTG CTC AAA ACC GTA CAG CGC AAC ATT CTT AAG	605

Leu 55	Ala	Pro	Asp	Tyr	Leu 60	Leu	Lys	Thr	Val	Gln 65	Arg	Asn	Ile	Leu	Lys 70	,
AAC Asn	GTC Val	CTT Leu	TCA Ser	CAA Gln 75	TTT Phe	CCG Pro	CTT Leu	TCC Ser	CCT Pro 80	TTT Phe	GCT Ala	ACA Thr	GCC Ala	TAC Tyr 85	CGA Arg	653
CCA Pro	GGT Gly	TGC Cys	CCA Pro 90	ATC Ile	GTC Val	AGC Ser	AAC Asn	GCG Ala 95	CAG Gln	CCA Pro	CAC His	TGC Cys	CAA Gln 100	CAG Gln	CCG Pro	701
CAG Gln	ATC Ile	CTG Leu 105	AAA Lys	CTC Leu	GAT Asp	ATC Ile	GAA Glu 110	AAC Asn	TTT Phe	TTC Phe	GAT Asp	AGC Ser 115	ATT Ile	AGC Ser	TGG Trp	749
TTA Leu	CAG Gln 120	GTC Val	TGG Trp	CGT Arg	GTG Val	TTT Phe 125	CGC Arg	CAG Gln	GCC Ala	CAG Gln	TTG Leu 130	CCA Pro	CGT Arg	AAT Asn	GTG Val	797
GTA Val 135	ACC Thr	ATG Met	CTG Leu	ACC Thr	TGG Trp 140	ATT Ile	TGT Cys	TGT Cys	TAT Tyr	AAC Asn 145	GAC Asp	GCG Ala	TTA Leu	CCG Pro	CAG Gln 150	845
GGG Gly	GCA Ala	CCA Pro	ACT Thr	TCG Ser 155	CCA Pro	GCC Ala	ATT Ile	TCC Ser	AAT Asn 160	CTT Leu	GTG Val	ATG Met	CGC Arg	CGT Arg 165	TTT Phe	893
																941
CGC Arg	TAC Tyr	TGC Cys 185	GAT Asp	GAC Asp	ATG Met	ACC Thr	TTT Phe 190	TCA Ser	GGT Gly	CAC His	TTC Phe	AAT Asn 195	GCC Ala	CGC Arg	CAG Gln	989
GTT Val	AAA Lys 200	AAT Asn	AAA Lys	GTG Val	TGC Cys	GGA Gly 205	TTG Leu	TTA Leu	GCG Ala	GAG Glu	CTG Leu 210	GGC Gly	CTG Leu	AGC Ser	CTC Leu	1037
AAT Asn 215	AAA Lys	CGC Arg	AAA Lys	GGC Gly	TGC Cys 220	CTG Leu	ATA Ile	GCT Ala	GCC Ala	TGT Cys 225	AAG Lys	CGC Arg	CAG Gln	CAA Gln	GTA Val 230	1085
																1133
																1181
																1229
	AAC ASN CCA Pro CAG Gln TTA Leu GTA Val 135 GGG Gly GAT ASP CGC Arg GTT Val AAT ASN ACC Thr CGG Arg TCG	AAC GTC Asn Val CCA GGT Pro Gly CAG ATC Gln 120 GTA ACC Val Thr 135 GGG GCA Gly Ala GAT GAA Asp Glu CGC TAC Arg Tyr GTT AAA Lys 200 AAT AAA Asn Lys 215 ACC GGG Thr Gly CGG GCG Arg Ala TCG CAT	AAC GTC CTT Asn Val Leu los CGG GCA ATC CYS CAS ATC CYS CST AAA AAT Lys Asn 200 ATT Thr Gly lie CGG GCG ATT Thr Gly lie CGG GCG ATT Thr Gly lie CGG ATG ALG CGC ATG Thr GLy lie CGG ATG ALG CGC ATG Thr GLy lie CGG ACG ACG ACG ACG ACG ACG ACG ACG ACG	AAC GTC CTT TCA ASN Val Leu Ser CCA GGT TGC CCA Pro Gly Cys Pro 90 CAG ATC CTG AAA Gln lle Leu Lys 105 TTA CAG GTC TGG Leu Gln Val Trp 120 GTA ACC ATG CTG Val Thr Met Leu 135 GGG GCA CCA ACT Gly Ala Pro Thr GAT GAA AGG ATG GIU ARG ASP Glu ARG 170 CGC TAC TGC GAT ARG Tyr Cys Asp 185 GTT AAA AAT AAA Val Lys 200 AAT AAA AAT AAA ASN Lys 215 ACC GGG ATT GTT Thr Gly Ile Val CGG GCG CTG CGT Arg Leu Ser	AAC GTC CTT TCA CAA ASN Val Leu Ser Gln 75 CCA GGT TGC CCA ATC Pro Gly Cys Pro 1le 90 CAG ATC CTG AAA CTC Leu 105 TTA CAG GTC TGG CGT Arg 120 GTA ACC ATG CTG ACC Thr 135 GGG GCA CCA ACT TCG Thr 135 GAT GAA CGC ATA GGG Gly Ala Pro Thr Ser 155 GAT GAA CGC ATG Gly 170 CGC TAC TGC GAT GAC ASN ASP 185 GTT AAA AAT AAA GTG Cyal Lys Asn Lys Val 200 AAT AAA CGC AAA GGC AAA GGC ASN Lys Asn Lys Cyal Cyal Cyal Cyal Cyal Cyal Cyal Cyal	AAC GTC CTT TCA CAA TTT GLA GLA GTC Pro Gly Cys Pro lle Val Ser Gla GTC GTG AAA CTC GAT GLA Leu Lys Leu Asp 105 TTA CAG GTC TGG CGT GTG AAG CTC GAT Leu Gla Val Trp Arg Val 120 GTA ACC ATG CTG AAC TCG CCA Gly Ala Pro Thr Ser Pro 155 GAT GAA CGC ATG GIG GAA ASP Glu Arg Ile Gly Glu 170 CGC TAC TGC GAT GAC GAA ASP ASP ASP ASP ASP ASP ASP ASP ASP A	AAC GTC CTT TCA CAA TTT CCG Asn Val Leu Ser Gln Phe Pro 75 CCA GGT TGC CCA ATC GTC AGC Pro Gly Cys Pro 11e Val Ser 90 CAG ATC CTG AAA CTC GAT ATC GIn 11e Leu Lys Leu Asp 11e 105 TTA CAG GTC TGG CGT GTG TTT ACT CAC ATC Val Thr Trp 11e 140 GGG GCA CCA ACT TCG CCA ACT TTT Trp 11e 140 GGG GCA CCA ACT TCG CCA GCC Pro Ala 155 GAT AAA ACC ATG TH GGG GAA TGC ATG ACC ATG ATG TH ACC ATG TH ACC ATG TH ACC ATG TH ACC ATG ATG ASD ASD ASD ASD ASD ACC ATG ACC ACC ATG ACC ACC ATG ACC ACC ACC ACC ACC ACC ACC ACC ACC AC	AAC GTC CTT TCA CAA TTT CCG CTT ASN VAI Leu Ser Gln Phe Pro Leu CCA GGT TGC CCA ATC GTC AGC ASN Pho Gln Leu Leu Lys Leu Asp Leu CGA ATC GTC AGC ASN Pho Leu Leu CGT AAA CTC GAT ATC GAA TTD CGC ACT ATC CGT AAC ATC CGT AAC ATC CGT ACC ACC ATC CGT ACC ACC ATC CGT ACC ACC ACC ACC ACC ACC ACC ACC ACC AC	AAC GTC CTT TCA CAA TTT CCG CTT TCC CAG GGT TGC CYPRO Gly Cys Pro 10 10 10 10 10 10 10 10 10 10 10 10 10	AAC GTC CTT TCA CAA TTT CCG CTT TCC CCT Asn Val Leu Ser Gln Phe Pro Leu Ser Pro 80 CCA GGT TGC CCA ATC GTC ASC ASC AAC GCG CAG Pro Gly Cys Pro 11e Val Ser Asn Ala Gln 95 CAG ATC CTG AAA CTC GAT ATC GLU ASN Phe 110 TTA CAG GTC TGG CGT TCG CGT ATC TTT CGC CAG ASN ASN Phe 120 CTA ACC ATG CTG ACC TTT TCT ATC TCG ATC ACC ACC ACC ACC ACC ACC ACC ACC ACC	AAC GTC CTT TCA CAA TTT CCG CTT TCC CCT TTT Asn Val Leu Ser Gln Phe Pro Leu Ser Pro Phe 80 Pro Gly Cys Pro Ile Val Ser Asn Ala Gln Pro 95 Pro Gly Cys Pro Ile Val Ser Asn Ala Gln Pro 95 Pro 120 Pro 1	AAC GTC CTT TCA CAA TTT CCG CTT TCC CCA GCT TTT GCT Asn Val Leu Ser Gln Phe Pro Leu Ser Pro Phe Ala 80 CCA GGT TGC CCA ATC GTC AGC ASC ASC ASC GGC CAG CCA CAC Pro Gly Cys Pro Ile Val Ser Asn Ala GIn Pro His 90 CAG ATC CTG AAA CTC GAT ATC GTC ASC ASC ASC Phe Pro Gly Cys Pro Ile Val Ser Asn Ala GIn Pro His 91 CAG ATC CTG AAA CTC GAT ATC GAT ATC GAT ASC Phe Phe Asp 110 TTA CAG GTC TGG CGT GTG TTT CGC CAG GCC CAG TTG Leu Gln Val Trp Arg Val Phe Arg Gln Ala Gln Leu 130 GTA ACC ATG CTG ACC TGG ATT TGT TGT TAT AAC GAC Val Thr Met Leu Thr Thy 140 GGG GCA CCA ACT TCG CCA GCC ASC TTG ASN ASN ASN ASN Glu Arg Ile Gly Glu Trp Cys Gln Ala Arg Gly 170 GGC TAC TGC GAT GAC AST GG GAT TTC CAG GCT CGG GGA ASN ASN ASN ASN ASN ASN ASN ASN ASN AS	AAC GTC CTT TCA CAA TTT CCG CTT TCC CTT TTT GCT ACA ASN Val Leu Ser Gln Phe Pro Leu Ser Pro Phe Ala Thr 80 CCA GGT TGC CCA ATC GTC ASN ASN CAI Leu Ser Gln Phe Pro Leu Ser Pro Pro Phe Ala Thr 80 CCA GGT TGC CCA ATC GTC ASN ASN ALA GLN Pro His Cys 90 Ile Val Ser ASN ALA GLN Pro His Cys 95 CAG ATC CTG AAA CTC GAT ATC GLN ASN Phe Pro ASP Ser 110 Ile Leu Lys Leu ASP Ile Glu ASN Phe Phe ASP Ser 110 Ile Leu Lys Leu ASP Ile Glu ASN Phe Pro ASP Ser 115 TTA CAG GTC TGG CTG GTG ATC TTT ACA CAG GTC TGG ATT TTT TTC GAT ACC ATG CTG ACC ATG ACC ATG CTG ACC ACG GTG TTG CCA ACT TTT TTP ILE Cys Cys Tyr ASN ASP Ala 135 Ile GLU ASP ILE CYS CYS TYR ASN ASP ALA 135 Ile GLU ASP ILE CYS CYS TYR ASN ASP ALA 135 Ile GLU ASP ILE CYS CYS TYR ASN ASP ALA 135 Ile GLU ASP ILE CYS CYS TYR ASN ASP ALA 135 Ile GLU ASP	AAC GTC CTT TCA CAA TTT CCG CTT TCC CCT TTT GCT ACA GCC Asn Val Leu Ser Gln Phe Pro Leu Ser Pro Phe Ala Thr Ala 75	AAC GTC CTT TCA CAA TTT CCG CTT TCC CCT TTT GCT ACA GCC TAC ASN VAL Leu Ser Cln cln pro Leu Ser Pro Phe Ala Thr Ala Tyr 75	AAC GTC CTT TCA CAA TTT CCG CTT TCC CTT TTT GCT ACA GCC TAC CGA ASN VAI Leu Ser Gln Phe Pro Leu Ser Pro Phe Ala Thr Ala Tyr Arg 85 CCA GGT TGC CCA ATC GTC AGC AAC GCG CAG CCA CAC TGC CAA CAG CCG Pro Gly Cys Pro Ile Val Ser Asn Ala Gln Pro His Cys Gln Gln Pro 90 CAG ATC CTG AAA CTC GAT ATC GAA AAC TTT TTC GAT AGC ATT AGC TGG Gln Ile Leu Lys Leu Asp Ile Glu Asn Phe Phe Asp Ser Ile Ser Trp 1105 TTA CAG GTC TGG GGT GTG TTT TGC CAG GCC CAG TTG CCA CGT AAT GTG Gln Val Trp Arg Val Phe Arg Gln Ala Gln Leu Pro Arg Asn Val 125 GTA ACC ATG CTG ACC TGG ATT TTT TTC GAT AGC ATT AGC TGG Val Thr Met Leu Thr Trp Ile Cys Cys Tyr Asn Asp Ala Leu Pro Gln 135 GGG GCA CCA ACT TCG CCA GCC ATT TCC AAT CTC AAT CTG ATG CGC GT TTT GIT AGC ASp Glu Arg Ile Gly Glu Trp Cys Gln Ala Arg Gly Ile Thr Tyr Thr 1705 GGG GCA CCA ACT GGG GAT GGC ATG TGC CAG GCT CGG GGA ATT ACC TAC ACC Asp Glu Arg Ile Gly Glu Trp Cys Gln Ala Arg Gly Ile Thr Tyr Thr 180 GTT AAA AAT AAA GTG GG GGA TTG TTA GCG GAG GCC CGC CAG TTG CCA ASp Glu Arg Ile Gly Glu Trp Cys Gln Ala Ala Arg Gly Ile Thr Tyr Thr 180 GTT AAA AAT AAA GTG GG GGA TTG TTA GCG GAG CTC CTC AAT ARG GCC CAG Arg Tyr Cys Asp Asp Met Thr Phe Ser Gly His Phe Asn Ala Arg Gln Clu Leu Gly Leu Ser Leu 200 AAT AAA CGC AAA GGC TTC CTG AAG CCT TTT TCA GGT CAC TTC AAT GCC CAG Arg Tyr Cys Asp Asp Met Thr Phe Ser Gly His Phe Asn Ala Arg Gln Gln Val 2205 AAT AAA CGC AAA GGC TTC CTG ATG CTG TTT TAA AAC AAT CTG AAT GCC CTC CTG AAT ARG GCC CTC CTG AAT AAA CGC AAA AAC TTG TAA AAA AT AAA GTG TAC CTG AAT TTG TAA AAA AAT AAA GTG TAC CTG AAT AAA CTG AAA AAT AAA CTG AAA AAT AAA CTG CTG CTG AAA AAA TAA AAT AAA CTG CTG CTG AAA AAA TAA AAA CGC AAA AAA TAA AAA TAA AAA TAA AAA TAA CTG CTG CTG AAA AAA TAA AAA CGC AAA AAA TAA CTG CTG CTG AAA AAA TAA AAA CGC AAA AAA TAA CAC CTG AAA AAA TAA AAA CGC AAA AAA TAA CAC CTG AAA AAA TAA AAA CGC AAA AAA TAA CAC CTG AAA AAA TAA AAA TAA AAA TAA CAC CTG CTG AAA AAA TAA CTA



(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUÉNCE DESCRIPTION: SEQ ID NO:43:

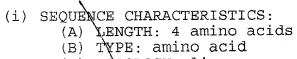
Tyr Xaa Asp Asp

1
4

- (2) INFORMATION FOR SEQ ID NO:44:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Ser Xaa Xaa Xaa 1 4

(2) INFORMATION FOR SEQ ID NO:45:



- (D) TOPOLOGY: linear

(ii) MOLECULE YYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

Zaa Val Thr Gly

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for scanning.		(Document title)	

Scanned copy is best available. Drawings are dark, and thinks are lines in specification and SEQUENCE